

FINAL

**Brown Trout
Laboratory Reproduction Studies
Conducted in Support of Development of a
Site-Specific Selenium Criterion**

**Revised
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Prepared for:

J.R. Simplot Company

Prepared by:



2500 55th Street, Suite 200
Boulder, CO 80301

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LIST OF ACRONYMS

AOC	Administrative Order on Consent
CAS	Columbia Analytical Services
CF	Cranio Facial
CI	Confidence Interval
dw	Dry Weight
EC/EC _x	Effect Concentration
ED	Edematous Tissue
FD	Finfold
GH-AAS	Gaseous Hydride Atomic Absorption
GSI	Graduated Severity Index
HS	Hoopes Spring
IDEQ	Idaho Department of Environmental Quality
ODA	Overburden Disposal Area
OLS	Ordinary Least Squares
SC	Saratoga National Fish Hatchery
SI	Site Investigation
SK	Skeletal
SPC	Spring Creek Trout Hatchery
SSSC	Site-Specific Selenium Criterion
TM	Technical Memorandum
TRAP	Toxicity Relationship Analysis Program
USFS	United States Forest Service
USEPA	United States Environmental Protection Agency

1.0 INTRODUCTION

A chronic site-specific selenium criterion (SSSC) is being developed for Hoopes Spring South Fork Sage Creek (SFSC) and the downstream receiving waters including Sage Creek and Crow Creek upstream of the Idaho and Wyoming State Line. Hoopes Spring is located in Sage Valley near the J.R. Simplot Company (Simplot) Smoky Canyon phosphate mine in Southeastern Idaho (Figure 1). In accordance with the Administrative Order on Consent (AOC) entered into by Simplot, Idaho Department of Environmental Quality (IDEQ), the US Forest Service (USFS), and the US Environmental Protection Agency (USEPA), a Site Investigation (SI) was conducted at the mine site in 2003 and 2004. Investigations to date have identified elevated concentrations of selenium in surface water being discharged via Hoopes Spring and South Fork Sage Creek Springs, which ultimately discharges to lower Sage Creek. Selenium released from overburden disposal areas (ODAs) at the mine has the potential to migrate vertically downward into the Wells Formation aquifer. Groundwater from the Wells Formation aquifer discharges at Hoopes Spring and South Fork Sage Creek Springs.

Source control actions implemented at the ODAs will limit infiltration to the Wells Formation, but will not immediately reduce selenium concentrations discharged from the Wells Formation via Hoopes Spring and South Fork Sage Creek Springs. In the interim, modification of the selenium surface water quality standard is being investigated.

Field monitoring studies are in progress to characterize the exposure environment, the aquatic community, and the physical habitat. Activities for the field monitoring studies are documented in the April 2007 Work Plan - Field Monitoring Studies for Developing a Site-Specific Selenium Criterion (NewFields 2007a).

This report for the study of brown trout (*Salmo trutta*) reproduction is presented as the first of two laboratory studies to evaluate the effects of ambient selenium concentrations in aqueous and dietary media on reproductive success of trout from the site. The second study involves a similar scope, but uses Yellowstone cutthroat trout (*Oncorhynchus clarki*). The laboratory studies were developed to complement information available from literature and an extensive field monitoring program for the study area. These laboratory studies are designed to assess potential effects of selenium accumulated in tissue of wild-caught adult brown trout on reproductive success, especially the development of young fish from fertilization through swim-up stages of development. The study design was presented in an October 17, 2007 Technical Memorandum - Methods for Testing Adult Brown Trout Reproductive Success. Laboratory portions of this testing were carried out at ENSR's environmental toxicology laboratory in Ft. Collins, Colorado by Dr. Rami Naddy. The deformities assessment was performed by Dr. Kevin Bestgen at Colorado State University's Larval Fish Laboratory.

The approach for the brown trout laboratory reproduction studies was based in part on the following published works:

- Kennedy et al. (2000). The effect of bioaccumulated selenium on mortalities and deformities in the eggs, larvae, and fry of a wild population of cutthroat trout (*Oncorhynchus clarki lewisii*);
- Holm et al. (2003). An assessment of the development and survival of rainbow trout (*Oncorhynchus mykiss*) and brook trout (*Salvelinus fontinalis*) exposed to elevated selenium in an area of active coal mining;
- Holm et al. (2005). Developmental effects of bioaccumulated selenium in eggs and larvae of two salmonid species; and
- Hardy (2005). Effects of dietary selenium on cutthroat trout (*Oncorhynchus clarki*) growth and reproductive performance.

Based on these and other works, the reproductive success of fish exposed to selenium via diet and water was identified as a highly sensitive endpoint. The approach also reflects the following understanding of the current state of the science regarding selenium toxicity:

- Chronic effects of selenium exposure to fish are due primarily to diet. Chronic toxicity is based on the magnitude and duration of exposure, as well as bio-uptake in the food web. The USEPA (2004) draft criteria document for selenium did not consider or use tests in which aqueous only exposure was tested. EPA states, “[b]ecause diet controls selenium chronic toxicity in the environment and water-only exposures require unrealistic aqueous concentrations in order to elicit a chronic response, only studies in which test organisms were exposed to selenium in their diet alone or in their diet and water were considered in the derivation of a chronic value.”
- Fish appear to be the most sensitive aquatic biota in the area of interest to chronic exposure and toxicity from selenium (Coyle et al. 1993; Hamilton et al. 1990; Hermanutz et al. 1996) (as cited in USEPA 2004).
- Reproductive success is the most sensitive biological end point for assessing selenium toxicity to fish (Lemly 1985a,b, 1992; Gillespie and Baumann 1986; Schultz and Hermanutz 1990; Coyle et al. 1993) (as cited in Lemly 1993).
- Selenium impacts on reproductive success in fish are strongly correlated to selenium content in eggs (Parametrix 2009)¹. Selenium in eggs is derived from maternal tissue, and is well correlated with whole body tissue selenium concentrations in maternal adults.

¹ Parametrix 2009 is a compilation document that reviews a number of important studies in the selenium literature. The conclusions drawn are based on the scientific evidence from numerous studies suggesting ovary or egg concentrations are the best tissue to correlate to effects. This position is also supported by USEPA in their revision of the National Selenium Criteria as relayed to the SSSC Work Group by Dr. Charles Delos.

- To date, three species of trout (i.e., brook, rainbow, and cutthroat) have been tested for bioaccumulation in adults and effects on development of young (Holm et al. 2003, 2005; Kennedy et al. 2000; Hardy 2005; Rudolph et al. 2008). No published literature has been found that indicates brown trout have undergone such testing to assess potential effects.

Because of the site-specific nature of selenium exposure and toxicity, wild-caught, reproducing fish from the study and reference areas are the best measure of current and potential impacts within this watershed. Brown trout are present in Hoopes Spring, Sage Creek, and Crow Creek. It is one of two predominant trout species (the other being Yellowstone cutthroat trout) found in these creeks and, although introduced, is recreationally important. Brown trout are by far the most abundant of the two trout species and are known to reproduce throughout the study area and therefore were used as a test species.

1.1 Background

Sage Creek downstream of Hoopes Spring regularly exceeds the chronic water quality standard for selenium. The frequency and magnitude of the exceedences decline downstream with tributary inflows. Only infrequent exceedences have been observed in Crow Creek immediately downstream of Sage Creek. While concentrations of selenium may exceed the surface water standard, it is not an explicit indication that the aquatic community is impaired. National surface water quality criteria adopted by states as standards do not always take into account the types of species present, nor the site-specific conditions, such as aqueous chemistry which may confound toxicity. Many factors influence the in-stream toxicity of selenium including the bioavailability of the form of selenium, tolerance of resident species (e.g., acclimation), and/or other factors that may enhance or ameliorate toxicity.

The brown trout adult reproduction testing used gravid adult wild fish captured at various locations from the study area (Figure 2), as well as hatchery fish for laboratory controls. Maternal transfer is believed to be one of the key factors influencing reproductive toxicity. Wild pre-spawn brown trout were collected from locations that represent the range of observed surface water selenium concentrations (NewFields 2007b). Aqueous and dietary selenium concentrations translate into a range of exposure conditions resulting in different body-burden loadings for parental fish, specifically adult female trout. It was anticipated that tissue concentrations in parental fish would confirm this. Gametes from the adult wild fish were collected and fertilized to evaluate reproduction. Although young were not exposed to aqueous selenium, they consumed any protein-bound organic selenium that was present in the yolk and passed on to the egg via parental exposure. The range and gradient of the selenium exposures, well-defined source area, exceedence of water quality standards and observations of thriving fish populations present a unique situation to examine selenium exposure and effects.

1.2 Objectives

The objectives of the testing presented herein are as follows:

- Document the range of selenium concentrations in wild parental fish due to in-situ integrated exposure of diet and water;
- Document the selenium concentrations in eggs produced by adults from different locations in the study area;
- Develop a relationship between selenium concentrations in maternal whole body tissue and egg tissue; and
- Develop relationships between egg tissue concentrations and measures of reproductive success and viability of young.

2.0 METHODS

The methods for testing adult brown trout reproductive success, including the study design plan and analysis details for the assessment of selenium exposure, were presented in a SSSC Workgroup – reviewed Technical Memorandum (TM) dated October 17, 2007 (NewFields 2007b). The methods for fish collection, egg collection and fertilization, and laboratory methods are briefly summarized below along with any deviations from the planned methods. ENSR conducted the reproduction testing for brown trout. Columbia Analytical Services (CAS) (Kelso, Washington) conducted the analytical chemistry for selenium concentrations in tissue. Appendix A describes the reproduction studies.

2.1 Wild Fish Collection

Because there is limited information on selenium toxicity to brown trout, there was some uncertainty as to where within the previously observed tissue concentration range reproductive effects might occur. To address whether the laboratory study would adequately cover the range of parental tissue concentrations expected in the system, brown trout tissue data for selenium were compiled for the stream segments of interest to examine the range of variability. The mean and its confidence interval (CI) suggest that the data are somewhat variable (mean (CI) - 13.27 ± 1.995). Since the system is comprised of small streams that have already been sampled numerous times, concern for the system is that over sampling can adversely influence the number of fish locally within the system, especially gravid females. Correspondingly, the goal was to identify the number of fish for collection that would provide a reasonably high probability of spanning the tissue concentration range of interest. The representativeness of the study is ultimately determined by capturing the range of effects and not the total number of fish. Next, the sample size to capture the range of tissue variability was estimated. How many samples are needed to cover the range of population of data (i.e., tissue concentrations), including at least one or more sample(s) that represent the upper 10th percentile? The following formula from Gilbert (1987) was used:

$$\alpha = 1 - (1 - p)^n$$

When rearranged to solve for n, it looks as follows:

$$n = \ln(1 - \alpha) / \ln(1 - p)$$

where:

α = Probability of at least one sample representing the upper p^{th} percentile;

p = target percent in number format; and

n = number of samples.

For this assessment, alpha was set at 0.05 and 0.1, while p was set at a range of percentile values. At a 95 percent confidence level (i.e., $\alpha = 0.05$), 29 samples should ensure that at least one or more samples would represent the upper 10th percentile (i.e., 90th). These 29 samples (i.e., adult females) were to be divided among the four exposure areas within the Crow Creek drainage. Rather than round down to seven fish per location, the number of fish was rounded up to 8 fish per location, which equaled 32 fish for the four locations. Based on Workgroup comments, the recommendation was to include additional fish samples for the reference area, thus an additional eight fish were added as targets for collection from the reference location. In total, for the five locations, it was estimated that collection of about 32 fish from the Crow Creek locations and 8 fish from the reference location (i.e., $n=40$) should capture the upper 10th percentile of the data distribution.

The upper 10th percentile was chosen because there will naturally be extremes in any environmental data, thus attempting to capture the entire range is not practical and over sampling of the spawning-age adults could easily occur in an effort to obtain fish with the highest tissue residues. Use of the 90th percentile captures a large proportion of the data. Based on the data currently available, the range of concentrations is such that capturing the 90th percentile, or upper 10th percentile, will result in capturing fish with 20 mg/kg dry weight (dw) selenium or more. Section 3.2.1 describes the initial data used to derive the upper 10th percentile and shows how the tissue concentrations from the adults collected for the reproduction study fit within that distribution. Section 4.7 describes the number of fish collected, and that the number of fish was adequate for the intended purpose of the study.

2.1.1 October Fish Collection

From October 22 to October 27, 2007, ripe and running brown trout were collected by electro-fishing at eight locations (Appendix B) (Table 1). In Crow Creek and Sage Creek, target locations included those where brown trout redds had been previously identified during Fall 2006. Other areas within these locations where brown trout might be expected to congregate and spawn due to favorable conditions, such as water depth, velocity, and substrate were also included (Figure 2).

Reference locations on Montpelier Creek and Stump Creek were also sampled for spawning brown trout. Snowslide Creek, near its confluence with Montpelier Creek, was evaluated and deemed to have inadequate flow and water depth.

Target age of fish for use in this study was 3+ years old. Fish of this age typically range from approximately 230 to 300 mm in length. Fish greater than 230 mm were collected, weighed and measured for length.

Adult brown trout were held instream near the location of capture in flow through “live cars”. Fish were held until October 30th, and were fed a commercial fish food pellet daily beginning October 27th. From October 27th to October 30th, fish were checked daily for signs of ripening. Ultimately, eggs could not be expressed from any of the retained females and all fish were released.

Water quality data were collected at each of the locations where brown trout were collected and subsequently held for possible spawning. In-situ field parameters, including water temperature, pH, conductivity, and dissolved oxygen were recorded (Table 2). A single water quality grab sample was also collected from each of these locations for analysis of dissolved and total selenium (Table 3 and Table 4).

2.1.2 November Fish Collection

From November 12 to 16, 2007, many of the same reaches sampled in October were sampled again for the presence of ripe and running brown trout (Table 5). Ripe or potentially ripe males and females were again held on-site in flow through “live cars” until they could be checked again for eggs or milt depending on gender. Collection methods were similar to those described above. No additional water quality data for selenium or ambient conditions were collected in November.

Eggs (from adult female trout) and milt (from adult male trout) were collected in the field November 13-15, 2007 for conduct of the reproduction tests. Fish were anesthetized using MS-222 to loss of equilibrium. Fish weight and length were then measured to the nearest 0.1 g and 1 mm, respectively. Trout were blotted dry, particularly the area around the urogenital opening to remove excess water that might contribute to premature water hardening of the eggs. The milt from several males at each location was expressed using a downward squeezing force, ventrally. Milt from several males was collected into a single plastic bag and stored on ice until added to individual egg batches from all females collected from that location. Eggs from each female were stripped from the vent in a similar fashion as the milt was collected from the males. Eggs from a single female were stripped into a pre-cleaned stainless steel bowl. Any blood, dirt or extraneous material was then removed from the bowl. Approximately 1 ml (depending on relative volume of eggs) of milt was added and then the egg / milt mixture was swirled gently to

ensure adequate mixing of gametes. The eggs and milt were allowed to sit undisturbed for ~1 minute. Then ~100 ml of local stream water (enough to just cover the eggs) was added to the bowl. The gametes were gently swirled for three minutes. Afterward, an additional 500 ml of stream water was added to water harden the fertilized eggs. The entire contents of the bowl were then poured into a labeled plastic bag and sealed. Each bag was labeled according to the female from which the eggs came, as well as the location. Prior to transport to the ENSR, the bag containing the fertilized gametes was partially inflated with oxygen, placed into a separate bag (double bagged), and returned to storage on ice (@ 4°C). The fertilized gametes were placed in a cooler for storage and transfer to the laboratory to protect them from sunlight and to keep them cold. A transponder that recorded temperature at 1 minute intervals was placed in each cooler prior to shipment to monitor the temperature during transport.

Adult fish were sacrificed for whole body selenium analysis. The adult fish carcasses were packaged in double plastic Ziploc® bags and stored on ice or frozen prior to shipment to ENSR along with the final egg batches. Because egg batches had to be delivered to ENSR within a narrow time window, and because ENSR had a large walk-in cooler/freezer, adult carcasses were initially shipped to ENSR. Once all carcasses were at ENSR and thoroughly frozen, a single shipment which included a subsample of eggs and all the adult fish carcasses for selenium tissue concentration analysis was sent on dry ice to CAS. Total selenium analysis and percent solids content were performed on all the submitted samples according to the methods described in the Work Plan (NewFields 2007a).

2.1.3 Hatchery Fish

Hatchery fish were used as method controls. Two sets of adult brown trout were obtained from the Saratoga National Fish Hatchery, Saratoga, WY (courtesy of Lee Bender) on October 23, 2007. Throughout this study, fish from this hatchery are identified as SC. The initial hatchery fish were obtained approximately three weeks prior to the first field-collected (wild) fish. Eggs from the initial set of hatchery fish were fertilized in the laboratory (transport of eggs and milt separately to the laboratory) according to the methods described above. Maternal fish were sacrificed to obtain whole body selenium tissue concentrations that corresponded to egg clutches from each female, consistent with the methods utilized for the wild fish.

A second set of hatchery fish were obtained in December 2007 from the Spring Creek Trout Hatchery in Lewistown, Montana as an additional method control. Eggs from these fish are referred to as SPC. These gametes were obtained in bulk as eyed eggs (i.e., already fertilized). Parents were unknown and mixed, thus whole body parental tissue concentrations were not obtained. Since the SPC eggs were a batch sample, they were submitted to the analytical laboratory for total selenium analysis as one sample.

The hatchery fish that were obtained served as method controls for the experimental process.. Hatchery fish and the resultant eggs are not subject to the same rigors as wild fish. The additional stresses that wild fish face beyond selenium exposure are not present for hatchery fish. As these non-selenium stressors for wild fish can also affect the test endpoints, including hatchery fish that do not experience these stressors in comparisons between these populations to study endpoints is inappropriate.

While hatchery fish were not used to quantitatively assess effects endpoints, the hatchery fish are important to illustrate the range of method variability that can and does occur in larval fish survival, growth, and deformities when no selenium exposure has occurred. The data for these hatchery fish and the measured response are included alongside the field-collected fish to illustrate that variability.

2.2 Reproduction Study and Laboratory Test Methods

The reproduction portions of this testing were carried out by ENSR's environmental toxicology laboratory in Fort Collins, Colorado. The methods are presented in detail in Appendix A. The study plan design was initially developed based on exposure areas and grouping of fish from the same collection areas. However, the study approach was modified to collect trout at several different locations (exposure areas) and raise eggs from each maternal fish as an independent unit (i.e., paired data). The collection of paired data for individual fish is expected to provide better insight on the relationships between tissue concentrations and reproductive success.

Adult trout carcasses and a subsample of eggs were sent to CAS for analysis of total selenium and percent solids. Tissue was analyzed for selenium using Gaseous Hydride Atomic Absorption Spectroscopy (GH-AAS) Method 7742. Percent solids were measured via freeze drying.

2.3 Deformity Assessment

Dr. Kevin Bestgen at Colorado State University's Larval Fish Laboratory was contracted to conduct the deformity assessment. He evaluated over 10,000 individuals and each individual fish was evaluated for up to four different deformities and four possible levels of deformity (Appendix C). Dr. Bestgen developed a process for assessing deformities which gave specific measurements to each ranking, thereby allowing for some measure of repeatability and accuracy. He received samples essentially as blinds because he did not know what the sample locations were or their locations relative to selenium concentrations.

The general criteria were adopted from Holm et al. (2003), and included assessments of craniofacial deformities (mostly of the head, eyes, and jaw), vertebral deformities, fin deformities, and edema. The original publication showed pictures of some deformities but

others, particularly the intermediate categories, were not illustrated or were poorly described. More specific definitions for each of the assessment categories were developed to give better repeatability and consistency across studies, and to aid others in learning the range of deformities possible.

Deformities in each of the categories described above were given a score from 0-3, with 0 being a normal condition and 3 being the most deformed. Some range finding was conducted over the first several samples to find background and severe levels of deformities in each category. Initial samples were rescored as necessary to bring them into compliance with the standards that were used throughout the assessment.

The protocol for assessing damage was to place several fish, head to the left, in a Petri dish and examine them under a dissecting microscope and 10X magnification. The lateral side was examined for spinal deformities (lordosis, kyphosis), appearance of the eye, head and snout shape, edema, and fin deformities. The fish was turned ventrally to look for mouth deformities and further spinal deformities (scoliosis), turned laterally again for the same criteria as the other side, and then dorsally for issues associated with eyes, head size, spinal deformities.

Craniofacial deformities included shortening of the jaw, snout, and missing or poorly developed eye or eyes, and head shape abnormalities. A slightly shortened lower jaw (≤ 1 lip width) received a 1, a shortened jaw = 2 lip widths or a slightly shortened and slightly disfigured jaw = 2, and a flat lower jaw or much disfigured (non-functional) jaw = 3. An assessment of fish independent of this study revealed that other brown trout of the same size and developmental state did not have the slight deformity that was assessed as CF = 1 for the jaw (J). Thus, the CF = 1 score where the J was concerned were deemed real. A slightly blunted snout (about 50 percent eye diameter, usually is $>$ than that) = 1, very blunt or flat = 2, deformed or bulbous = 3. Eye deformities were scored as one eye blind or poorly pigmented or poorly developed = 1, both poorly developed = 2, both blind = 3. Skulls that were slightly bulbous ($1/3 >$ normal) = 1, moderately bulbous ($2/3 >$ normal) = 2, and bulbous ($1x$ or $>$ than normal) = 3.

Skeletal deformities included any deformity of the vertebrae or spines. A slight bend of less than 45 degrees (but $>$ than body width off of straight) or a minor body constriction (e.g., a tight rubber band about the body effect) was given a score of 1, 2 slight bends or constrictions anywhere, or bend of $>$ 45-90 degrees was scored a 2, and multi-directional bends $>$ 90 degrees were given a 3.

Fin deformities included variation in fin or finfold morphology and a slightly smaller or missing fin (in thin fish, the adipose fin was often absent, indicating fat absorption, not uncommon and scored 1) or one with a bend or incomplete ray development (in older fish) was given a 1, 2 fins damaged or malformed = 2, and $>$ 2 fins malformed or if fins were missing (except adipose) was = 3.

Edema was detected by an obvious swelling and fluid buildup, usually abdominally, and ventrally, which often displaced the gut, and was usually clear fluid that was slightly soft when touched with a blunt probe. Slight edema = 1 was for a fish with up to 1X swelling of the normal body width or depth, up to 2x = 2, and > 2x = 3.

2.3.1 Data Reduction of Deformity Rankings

Individual files, representing scoring sheets, were received for each sample evaluated. All files were combined in Excel to form a master file. Data were summarized using the Pivot table function in Excel to produce counts and percentages of normal fish, deformed fish, and total number of fish evaluated. Similar to the method of Holm et al. (2005), a Graduated Severity Index (GSI) was derived based on the deformity rankings and counts for progeny from each parent. A total score was computed as follows:

$$[(\# \text{ fish for CF}=1) \times (1)] + [(\# \text{ fish for CF}=2) \times (2)] + [(\# \text{ fish for CF}=3) \times (3)].$$

This method differs slightly from Holm et al. (2005) as it weights each ranking with more weight given to more severe deformities. Fish scored as 0 (normal) observations did not enter into this calculation of total score. The final GSI score was computed as the sum score/total # fish including those ranked as "0". The total scores were summed and divided by the number of categories of deformities assessed (usually 4) to derive a mean GSI score.

Because the USEPA's Toxicity Relationship Analysis Program (TRAP) version 1.2 (Erickson 2008) logistic functions were designed to derive an inverse sigmoidal curve, commonly used to illustrate the dose-response curve of increasing exposure concentration and declining biological observation (e.g., survival, growth, etc.), deformities were evaluated as the sum fraction of normal fish (sum of normal fish/ total number of fish) for each deformity. This approach did not take into account severity of deformity, simply the frequency of deformities which is consistent with USEPA's (2004) approach to analysis of similar data.

2.4 Statistical Analysis

Multiple test-effects endpoints were measured at different times during the test including: fecundity, fertilization success, hatching success, deformities, length, weight, survival (different times during the study), and tissue concentrations (egg and whole body). These endpoints were consistent with those of Holm et al. (2005), Hardy (2005), and Kennedy et al. (2000). Feeding success was added as a test endpoint to evaluate the change from endogenous to exogenous feeding post swim up.

Scatter plots and best-fit ordinary least squares (OLS) regressions were used as an exploratory tool to evaluate the potential for meaningful relationships. Ordinary least squares regression

analysis was used as a preliminary method to assess if relationships existed between individual exposure assessment endpoints (i.e., parental selenium body burdens or egg selenium concentrations) and test-effects endpoints measured in the study. The dose-response relationships for exposure and effects endpoints were evaluated further. Logistic regression analysis was performed using USEPA regression-analysis software (TRAP version 1.2; Erickson 2008) for the effects endpoints showing the strongest relationships to the exposure endpoints. USEPA's TRAP software provides a number of statistical analysis tools, including logistic regression, to evaluate the presence of dose-response relationships. The logistic regression approach is consistent with the methods utilized by the USEPA in their assessment of dose-response data for the 2004 Draft Criterion. USEPA's TRAP software also allows for prediction of Effect Concentration (EC) values to estimate thresholds for potential effects for brown trout. Both EC₁₀ and EC₂₀ values were derived for each relevant relationship developed.

3.0 RESULTS

3.1 Surface Water Quality

Water quality data were collected in October during the initial effort to capture spawning trout. Tables 3, 4, and 5 present field measured parameters, conventional, and selenium concentrations in surface waters, respectively. At the upper Crow Creek locations (CC-75, 150, and 350), total selenium ranged from 0.00055 (CC-75) to 0.0015 mg/L (CC-150). In Sage Creek at LSV-2C, the selenium concentration in surface water was 0.0384 mg/L. At the lower Crow Creek locations downstream of Sage Creek, selenium concentrations were 0.0017 mg/L at CC-3A and 0.0028 mg/L at CC-1A. At both the Montpelier and Stump Creek locations, selenium was less than detectable (0.0002 mg/L) (Table 4). No additional water quality data were collected in November, as the time frame between the October and November sampling period was short. The expected exposure for fish from an exposure area is aggregated over a period of time, thus the exposure is best represented by the dietary and surface water selenium concentrations occurring prior to the time of sampling.

Water quality data for selenium have been collected at numerous locations within the study area and outside the study area over a fairly long period of time. Two additional sources of data are available to evaluate selenium concentrations both temporally and spatially. These datasets include:

- Fall 2006 to Spring 2008 surface water quality data collected for the SSSC study; and
- Spring and Fall annual effectiveness monitoring program data.

Information from these datasets is presented in Section 4 to help put selenium concentrations in surface water measured in October 2007 in context.

3.2 Wild Fish

During the October sampling period for spawning brown trout, approximately 300 adult brown trout were captured and 104 were retained as potential spawners. Of these, 36 were males and 68 were suspected females (Table 1). After a brief holding period all of these fish were released at the locations from where they were collected, because no eggs could be stripped from the female fish.

During the second sampling round in November, more than 300 adult brown trout were collected over a five day period. Equivalent numbers of prospective ripe males and females

were retained. Eggs were collected from 26 ripe females from three locations representing a range of ambient selenium concentrations in water, including two locations in the watershed upstream of Sage Creek and one location in Sage Creek. Consistent with the October sampling, the largest number of brown trout was collected in Sage Creek from upstream of South Fork Sage Creek to Hoopes Spring. During October and November, several brown trout were collected at the Meade Peak Ranch (CC-1A and CC-3A). However, no ripe females which would readily express eggs were captured. No ripe fish were collected from the Stump Creek reference area (Table 5).

3.2.1 Adult Size and Selenium Concentrations -Whole Body and Eggs

The initial target range for fish age and size was 3+ years (>230 mm). The trout collected from the field ranged from 265 mm to 391 mm with most of the females being 300 mm or larger. Ripe fish, particularly females, were generally closer to 300 mm in size and often larger than 300 mm. It is assumed that the size range of female fish collected represents the 3 and 4 year old age classes. A narrower range of adult sizes would have been preferred because fish size (i.e., age and maturity) can affect egg production. However, the number of ripe females collected made it necessary to include fish from a wider size range to meet study goals.

The whole-body selenium concentrations measured in the adult female carcasses whose eggs were used in this study are presented in Table 6. Whole-body selenium concentrations measured in field-collected brown trout ranged from 4.7 to 22.6 mg/kg dw. Figure 3 shows adult hatchery and wild fish lengths relative to their respective whole body selenium concentrations. The hatchery fish are larger and have a lower whole body selenium concentration, while the wild fish have considerable variability in whole body selenium concentrations reflecting the varying histories of selenium exposure. Figure 4 shows a similar relationship using adult fish weight (wet). The range of hatchery fish weights is larger than that observed for length, however, a similar trend is observed for wild adult fish, exhibiting relatively consistent weights despite the variation in whole body selenium concentration.

Figure 5 shows the size distribution (based on adult length) versus the whole body selenium concentration for the adult female brown trout collected as part of this reproduction study in November (red asterisks), the brown trout tissue concentrations collected as part of the field monitoring for the SSSC studies (Table 7), and any brown trout data collected since 2004 when tissue data were collected for the Smoky Canyon Mine Site Investigation. The range of tissue concentrations illustrated in Figure 5 (excluding the adults for the reproduction study) represents the brown trout data available at the time the adult study was being planned and are the basis for the number of samples derived (Section 2.1) as a target for field collection. It is important to note that previous monitoring and redd evaluations indicated very low numbers of adult fish of spawning size found in the Hoopes Spring channel (HS-3) or Hoopes Spring location (HS). As shown in Figure 5, data for fish tissue concentrations greater than 25 mg/kg dw were only found

in fish from Hoopes Spring. The range of tissue concentrations for female trout collected in November spans nearly the entire range of brown trout tissue concentrations collected from the previous field monitoring data.

The concentrations of selenium in egg batches from each female parent were also measured. Selenium concentration in fish eggs collected from CC-150 ranged from 6.2 to 12.8 mg/kg dw. Egg selenium concentrations from CC-350 ranged from 6.9 to 14.0 mg/kg dw, while those from LSV-2C ranged from 11.2 to 40.3 mg/kg dw (Table 6). Figure 6 shows the egg selenium concentrations ordered from lowest to highest from hatchery fish and wild collected fish. Appendix D includes the raw data for selenium concentrations in whole body and egg tissue.

3.3 Hatchery Fish

The Saratoga National Fish hatchery (SC), Saratoga, WY, was the first source of brown trout eggs for the laboratory controls in this study. Because these fish were spawned about 3 weeks ahead of the field-collected (wild) fish, due to spawning time in the field versus spawning time at the hatchery, it was not possible to select smaller spawning females from the hatchery. These eggs were fertilized at ENSR following collection at the fish hatchery. As discussed later, hatch rates were lower than expected and some fungal contamination occurred in the SC egg batches, therefore, a second set of hatchery eggs were obtained. Spawning was complete for SC hatchery fish, thus an alternative source of eggs was necessary.

The Spring Creek Trout Hatchery (SPC), Lewistown, Montana, was the second source of brown trout eggs for the laboratory controls in this study. This second set of eggs was obtained to ensure that the test system was suitable. Because it was late in the season these eggs were only available as eyed-up embryos.

3.3.1 Adult Size and Selenium Concentrations – Whole Body and Eggs

Eight adult trout from the Saratoga hatchery were used, ranging in length from 420 mm to 562 mm. Hatchery fish were larger than fish captured at field locations (Table 6). Maternal whole-body selenium concentrations measured in the hatchery trout ranged from 2.5 to 4.3 mg/kg dry wt. Bulk eyed-eggs were received from the SPC, thus no information of specific adult size and whole body maternal tissue concentrations was gathered.

The selenium concentration of the SC hatchery eggs ranged from 0.76 to 1.2 mg/kg dw, while the selenium concentration in the SPC hatchery eggs was 0.73 mg/kg dw (Table 6). As illustrated in Figure 6, egg selenium concentrations in the hatchery fish were considerably lower than egg selenium concentrations in the wild fish.

3.4 Reproduction Testing

Appendix A details all laboratory results of the reproduction study. The following endpoints were measured in the laboratory: fecundity, hatch, growth, survival/mortality, and feeding success (growth) post swim up. Additional measures included day to first hatch, day of swim up, and day of test termination.

3.4.1 Fecundity

Fecundity, measured as the number of eggs/female, was measured for each parent. Section 3.1 of Appendix A details egg quantity per female, number of eggs used in the study, and selenium tissue concentrations. Twenty-six egg samples from wild-collected brown trout were submitted to ENSR for the reproduction study. Of these, one set of eggs was dead upon arrival (LSV-2C-006). The number of eggs from individual field-collected fish ranged from 161 (LSV-2C-010) to 1,658 (CC-150-016). The number of eggs collected from the individual SC hatchery fish ranged from 1,248 to 5,448. Figure 7 illustrates the number of eggs relative to adult size (length). As mentioned, the hatchery fish were larger than the field-collected (wild) fish, and on average had almost two times or more eggs.

Figure 8 shows the relationship of the total number of eggs produced from each female versus the respective concentration of selenium in the eggs. Figure 9 shows the relationship of the total number of eggs produced from each female versus the respective concentration of selenium in the adult female fish.

3.4.2 Egg Mortality

The goal of the test was to begin each test chamber with 600 fertilized eggs. Not all wild fish produced sufficient egg numbers to do this, so a proportion of eggs was used to begin the test that allowed for adequate biomass of eggs to be left over for selenium concentration analysis. The minimum number of eggs included in a test chamber was 100 (LSV-2C-010). For sample LSV-2C-006, the entire egg batch was declared dead upon delivery to the laboratory and was not included in the test. For sample LSV-2C-007, eggs were initially included in the test, but it was determined, due to high numbers of eggs turning opaque, that these eggs were not fertilized, thus eggs from this sample were also not included in the test. Maternal whole body tissue and an egg subsample were collected for these two samples. Table 3-1 in Appendix A summarizes the number of eggs placed in the study for each sample. Egg mortality was measured based on the number of fish that hatched subtracted from the number of eggs at test initiation for each sample.

Egg mortality for the SPC parents was low (<3 percent), while egg mortality for SC parents ranged from 6.8 percent to 88 percent. It is not clear if higher egg mortality in SC derived eggs

was due to incomplete fertilization or due to the fungal problem that arose with these eggs. Egg mortality from parents collected at the background areas ranged from as low as about 3 percent to as high as 85 percent. From the LSV-2C exposure area, egg mortality ranged from 1 percent to 49 percent across each of the conditions: hatchery, background (CC-150), low (CC-350), or high (LSV-2C) exposure, egg mortality was highly variable (Figure 10).

3.4.3 Hatch to Swim Up

Appendix A, Table 3-3 summarizes hatch data for the different samples, including day of first hatch, percent hatched, day of swim up, and percent swim up. Day of first hatch was simply the day when hatching was first observed for eggs in a sample. Percent hatch was the number of eggs that successfully hatched divided by the original number of eggs for that sample times 100. The day of swim up was when greater than 80 percent of the alevins for a specific clutch of eggs reached swim up (i.e., absorbed their yolk and were actively feeding). Percent swim up was the number of alevins that had absorbed their yolk sac and were actively feeding divided by the total number of eggs used to begin the test times 100.

The percent hatch for the SPC hatchery fish was 97.5 to 100 percent. The holding period was significantly shorter for SPC eggs. This difference may contribute to the higher hatch success. The percent hatch for the field collected eggs was 11 to 93 percent. Eggs samples collected from CC-150 ranged from 14.8 to 97.3 percent, with an average of 71.9 percent. Average hatch out for the eggs collected from fish at CC-350 was 56.4 percent. Average hatch out from LSV-2C was 83.9 percent (not including LSV-2C-007 whose eggs were not fertilized). The lowest percent hatch from the LSV-2C treatments was 50.6 percent, while the highest was 99.0 percent. The field collected fish eggs (which were fertilized in the field) indicated that the change in fertilization technique (field fertilization vs. laboratory fertilization) resulted in a higher hatch success rate compared with the SC hatchery fish (which were fertilized in the laboratory). Figure 11 illustrates percent hatch for each sample ranked from lowest to highest egg selenium concentration. Figure 12 shows the relationship of percent hatch to swim up. There is for most fish, a 1:1 relationship of hatch to swim up. However, for several LSV-2C samples, while eggs hatched (>50 percent), yolk fry never swam up, despite the extension of the study 15 days past the time of swim up to assess feeding transition. The range of egg selenium concentrations for those alevins that did not swim up was 26.8 to 40.3 mg/kg dw. Except those five samples shown for LSV-2C, eggs that hatched resulted in young that were able to reach swim up. As shown in Figure 13, the eggs from those five samples had the highest selenium concentrations (i.e., greater than 25 mg/kg dw). Figure 13 shows the number of days to first hatch and swim up versus egg selenium concentrations. While hatch rates were similar across the board, the number of days to swim up increased with egg selenium concentrations greater than 25 mg/kg dw.

3.4.4 Mortality

Mortality was assessed at several different periods during the test, including:

- **at swim up** ($[(\text{number of eggs used to begin the test} - \text{number of fish surviving to swim up}) / \text{total number of eggs at beginning of test}] * 100$);
- **from hatch to test termination** ($[\text{percent survival at hatch} - \text{percent survival at test end}]$);
- **at test termination as overall mortality** ($[(\text{number of eggs used to begin the test} - \text{total number of fish at the end of the test}) / \text{total number of eggs at beginning of test}] * 100$); and
- **at the end of the 15-day post swim up feeding success trial** ($[(\text{number of fish used to begin the post swim up feeding trial, usually } n = 100 - \text{number of fish at the end of the feeding trial at 15 days}) / \text{total number of fish used to begin the test}] * 100$).

Mortality at swim up ranged from 0 to 6 percent in SPC fish, 7 to 90 percent in SC fish, and about 4 to 92 percent in wild fish (excluding the samples where no eggs were fertilized or eggs died before test initiation). Figure 14 shows mortality at swim up and overall mortality at the end of the test by location ranked by egg selenium concentration. It shows that overall mortality closely tracks mortality at swim up for most samples, except those from LSV-2C where overall mortality is greater than swim-up mortality. No real trends are observable from these data.

Figure 15 shows mortality in a similar fashion for the period, hatch to test termination. Mortality from hatch to the end shows a clear trend of low mortality in most all samples (i.e., <10 percent) and much higher mortality in a number of samples from Sage Creek below Hoopes Spring.

Percent mortality for fish from hatch to test end was substantially greater in the same five LSV-2C samples as those that did not achieve swim up (Section 3.4.3) and that had the highest egg selenium concentrations (greater than 25 mg/kg dw). Mortality during this period drops substantially in samples where egg selenium was about 20 mg/kg or less as shown by the 10 percent or less mortality for all remaining samples.

Finally, mortality as a function of the 15-day post swim up feeding trials was evaluated and is shown on Figure 16. The duration during which mortality was measured for this phase of the test was short, extending from swim up to 15 days past swim up to evaluate if fish could transition to active feeding. Similar to mortality from hatch to test end, mortality during the feeding trial shows highest mortality only occurred in the LSV-2C samples. Isolation of post-hatch mortality shows that for fish from most samples, mortality is low and only increases at locations where egg selenium concentrations were highest. Furthermore, the greatest percentage of mortality occurred in the last 15 days of test as shown in the difference between mortality rates in Figures 15 and 16. This is consistent with the prior observations that several

of the samples from LSV-2C with the highest egg selenium concentrations did not swim up, therefore, their ability to transition to active exogenous feeding was impaired.

3.4.5 Growth

Growth endpoints measured include length and dry weight. The results of length and dry weight analysis for the target of 20 organisms per chamber at the end of the 15-day post swim up phase of the study are provided in Appendix A, Table 3-5. Average length of fish per sample location is shown in Figure 17. There were no apparent differences in length noted between hatchery fish and wild fish, or among wild fish, following the feeding trial. Growth as measured by dry weight is shown in Figure 18. Similar to the length data, these are presented in a rank order from lowest to highest egg selenium concentrations. There does appear to be an inverse relationship of increasing egg selenium and reduced dry weight. However this difference is not as apparent when the hatchery fish are excluded.

3.4.6 Deformities

Appendix B provides a summary of the counts of deformity rankings for each sample and a series of graphics illustrating sample deformities as a percentage of each sample. Observations made during scoring that resulted in defining a level of severity for a fish being examined are reported below:

- **Cranio-Facial Deformities** - Usually factors occurred together so a combination of two "1" conditions = 2, three "1" conditions = 3, or a 1 and a 2 = 3, and so on. For example, a deformed jaw and a blind eye = 2, two blind eyes = 2, but a badly deformed jaw (= 2 alone) plus a blind eye (= 1 alone), = 3.
- **Skeletal Deformities** - Bends caused by skeletal deformities were usually detectable from normal bending of the body during preservation (these fish were usually well preserved, very straight) by presence of a slight or greater bump below the surface of the epidermis on the outside of the bend. However, some fish with SD = 1 had just a very slight bend in the range the deformity described but could be due to preservation or the poor condition of the fish. This was sometimes especially true in larger fish, which may be more muscular and undergo stronger contraction during preservation and thus, bend slightly. A score "CF = 1" was a slight deformity, if at all. The scores of SD = 1 involving kyphosis or lordosis were deemed real because that is an unusual preservation deformity. Also, samples BKD 015 SU (i.e., extra fry from CC-150-015 at swim up), LOW 008 SU (i.e., extra fry from CC-350-008 at swim up), and SC 003 SU (i.e., extra fry from SC-003 at swim up) were re-examined; most fish were very straight so some samples with higher SD scores (e.g., post swim-up samples) were determined accurate. Thin fish were difficult to score, as they often looked emaciated.

- **Fin and Finfold Deformities** - Often fins were malformed associated with vertebral deformities that did not permit proper development. Folded finfolds as a result of preservation were not counted.
- **Edema** - Edema was not originally scheduled for assessment because it was thought sometimes not a teratogenic effect and may be transitory as fish develop. However, it was assessed because it was common in one early sample and not others, and because it was thought a condition that could affect emergence, mobility, and other factors that may limit survival of fish in the wild. The yolk, which was present in some quantity in some study specimens, also created some swelling but was typically yellowish, opaque, and small, and hard to the touch in preservation.

A sample of 50 fish and a sample of 30 fish were scored twice, the same fish for each batch but not necessarily the same order. This sample was characterized by a low incidence of fin deformities (slow development) and a high incidence of jaw deformities and blindness (SC 003 SU). Those cranio-facial traits are difficult to score because they are additive, and subjective as to severity. Thus, the results may be a conservative view of what score replicability should be like for other traits in other samples that are easier to score.

Replicability of frequency of cranio-facial abnormalities was high among assessments at 50 and 52 percent in the first sample of 50 fish, and identical frequencies of 46.7 percent in each assessment for the sample of 30 fish. The cumulative sums of the scores were also quite close, but reflecting variability in scoring for all three categories of severity in each sample. Replicability of fin ray development assessments for both frequency and the sum of the scores was identical in both samples.

The results of the deformity assessment are visually displayed in Figures 19 through Figure 22. The results of the cranio-facial (CF), skeletal (SK), finfold (FD), and edematous tissue (ED) deformity frequency are depicted separately. Each of these figures is similar for the remaining deformity assessments and present the data on a percent basis for each of the rankings used in the assessment.

Cranio-Facial Deformities

For CF deformities (Figure 19 and Appendix B Figure 1), hatchery fish, except young from two parents, showed that greater than 80 percent of the young in each sample were ranked as normal. When deformities were present in the hatchery fish, they were ranked as slight. For the two samples that had higher numbers of non-normal fish (SC-003 and 004), craniofacial deformities ranged from slight to severe. For wild fish, from background and low areas (CC-150 and CC-350) from about 65 to 75 percent of all fish were ranked as normal for cranio-facial deformities. CF deformities ranked as slight were observed more often in fish from CC-150. At

CC-150, the percentage of normal fish ranged from 52 to 96 percent, whereas at CC-350 it ranged from 59 to 93 percent. For the CC-150 and CC-350 samples, fish not ranked as normal were predominantly ranked as slightly deformed for the CF characteristic. For samples from the LSV-2C area, more than 70 percent of the fish examined ranked as normal. Of the 12 samples submitted from the LSV-2C area, five (LSV-2C 003, 004, 005, 021, and 010) had a higher incidence of slight, moderate, and severe CF deformities in fry examined. For these five samples, fish were more often ranked as moderately deformed than those ranked as slightly deformed for this characteristic. In the remaining 7 samples, from 77 to 97 percent of fish in each sample ranked as normal, similar to the hatchery or background samples (Figure 19).

Skeletal Deformities

For SK deformities, the range of normal fish found in each sample was 60 to 91 percent and collectively, for both sets of hatchery fish was greater than 80 percent for all samples. In fish samples from CC150 and 350, the range of normal fish was 51 to 92 percent. For all sample from CC-150, 80 percent were ranked as normal fish, while at CC-350, only 66 percent of fish were ranked as normal. For fish from the LSV-2C area, the range of normal fish was 0 to 93 percent (Figure 20 and Appendix C Figure 2). Overall, 65 percent of the fish were ranked as normal. For this deformity category, there was a higher frequency of fish in most samples ranked as slight. The frequency of SK deformities in the moderate to severe range for hatchery fish or fish from CC-150 was low. Moderate to severe SK deformities were observed more frequently in fish from CC-350. Fish from LSV-2C showed a similar trend for SK deformities to those observed for CF deformities. The same five samples had higher percentages of the sample with slight, moderate, and severe SK deformities. For some of the fish in these five samples, larvae had deteriorated and could not accurately be ranked (shown as the blank category Figure 20). As noted in Appendix A, some larvae were clearly dying, but not dead, thus began deteriorating prior to them being preserved.

Fin or Finfold Deformities

For fin or FD deformities, the range of hatchery fish ranked as normal was 81 to 100 percent while overall for both sets of hatchery fish, greater than 90 percent of the fish were ranked as normal. For fish from CC-150, the range was 92 to 100 percent of fish ranked as normal in individual samples, while overall, 98 percent of the CC-150 fish were ranked as normal. The range of fish ranked as normal for CC-350 samples was 73 to 89 percent while overall, 85 percent of the fish from all samples were ranked as normal. Fish ranked as slight moderate, and severe for finfold deformities were observed more often in samples from CC-350 but the percentages were 6 percent or less for each of these categories. Normal fish from the LSV-2C area ranged from 0 to 100 percent. Across all samples, 80 percent of the fish were ranked as normal. Excluding samples LSV-2C 003 and 010, whose fish were too deteriorated to accurately assess fin deformities, the range of normal fish was 16 to 100 percent normal (Figure

21 and Appendix C Figure 3). The same five samples from LSV-2C either could not be evaluated for this characteristic, due to the condition of the fish in that sample, or had higher frequencies of moderate and severe fin deformities. As stated previously, several samples from LSV-2C experienced mortality before the swim up stage or did not swim up at all. Some of these fish were deteriorated and an accurate assessment of the severity of the deformity for this condition could not be made.

Edema Deformities

ED was almost non-existent in hatchery fish and fish from CC-150 and CC-350 (Figure 22 and Appendix C Figure 4). Similar to the other three categories of deformities evaluated, edema was present at a higher and more severe level in the same five samples as identified previously for LSV-2C samples.

Graduated Severity Index (GSI)

To capture the severity of each of these deformities into a single unit, a GSI was derived for fry originating from each egg batch as described in Section 2.3.1. The mean GSI score for all treatments ranked from left to right by the lowest to highest egg selenium concentrations is shown in Figure 23. Because a fish can have more than one deformity score, rankings higher than 1 are possible. Mean GSI scores show that for most samples GSI scores are 0.2 or less and a small number of samples ranged from 0.2 to 0.4. Mean GSI scores of 0.4 or less span the range of treatments/sites (hatchery and field). Only a small number of samples, the same five as identified in previous sections, from LSV-2C had mean GSI scores that were substantially greater than 0.4, ranging from 0.87 to 1.7 indicating that these samples had a higher frequency and severity in the level of deformities found. Figure 23 also shows the number of fish assessed from each sample.

4.0 ANALYSIS OF RESULTS

The following section presents preliminary analysis of the brown trout study data. A focus of these analyses is the relationship between maternal tissue concentrations and various reproductive effects identified in literature. Additional analyses will ultimately be conducted in conjunction with the full range of Site-specific information on exposure conditions and the biological community structure. These findings will also be contrasted with the work of others.

4.1 Surface Water Quality

Table 4 shows selenium concentrations from October 2007 surface waters, as well as selenium concentration data collected from pre- and post-October 2007 monitoring events. Both total and dissolved selenium analyses were conducted, and dissolved selenium is dominant in all surface water samples. At all of the upstream background locations on Crow Creek, the October 2007 total selenium concentrations are within the range of total selenium concentrations measured from 2006 to 2008 during seasonal monitoring. Total selenium concentrations at the Crow Creek locations upstream of Sage Creek (CC-75, CC-150, CC-350) are always below Idaho's Surface Water Quality selenium standard (0.005 mg/L).

The total selenium concentration at LSV-2C in October 2007 was high (0.0384 mg/L) compared to previous sampling events at this site. Figure 24 shows longer-term monitoring at the Hoopes Spring location immediately upstream of the LSV-2C location and shows that surface water selenium concentrations were increasing in 2007 and 2008.

Longer-term water quality monitoring in Sage Creek near Crow Creek road (LSV-4) provides information on selenium concentrations in Sage Creek before discharge to Crow Creek. Figure 24 shows longer-term selenium concentration data from this location and a gradual increase in selenium concentrations during 2007 and 2008 consistent with increases observed at Hoopes Spring and South Fork Sage Creek Springs. Concentrations during 2007 and 2008 ranged from 0.008 to 0.011 mg/L total selenium at LSV-4.

Selenium concentrations measured in Crow Creek samples collected downstream of Sage Creek at CC-1A and CC-3A in October 2007 were lower than Idaho's Surface Water Quality Standard. This is consistent with measurement data before and after this time period for these locations (Table 5). Selenium in Crow Creek downstream of Sage Creek is typically lower than the State Standard as illustrated by data in Figure 24 and Table 4.

Overall, selenium concentrations are highest in Hoopes Spring and Sage Creek immediately below Hoopes Spring (LSV-2C), and decrease in surface water with distance downstream of

Hoopes Spring (Figure 24). In Sage Creek upstream of its discharge to Crow Creek, selenium concentrations exceeded the State Standard of 0.005 mg/L, recently by about 2 times. In Crow Creek downstream of Sage Creek, infrequent exceedences of the State Standard have been observed. Regular monitoring data suggested selenium in Crow Creek downstream of Sage Creek is on average about 0.5 times the State Standard (0.005 mg/L). These surface water data set the stage for selenium exposure to fish and their food resources. Exposure is primarily dietary for fish but does include an aqueous component. Whole body maternal tissue concentrations of selenium described below integrate the water and diet exposures to provide a good indicator of exposure and bioaccumulation.

4.2 Relationship of Maternal Whole Body to Egg Selenium Concentrations

While more recent studies focus on selenium concentrations in eggs as the best predictor of reproductive effects there is also a substantial amount of information on whole body tissue concentrations in the literature. This initial regression analysis was performed to better understand the relationship between maternal whole body and egg selenium concentrations, and the results of the preliminary analysis confirm a strong relationship. The remainder of the analyses in this study focuses on the effects endpoint relationships to egg selenium concentrations. The relationship between egg selenium and whole body selenium allows for consideration of both measures as the study progresses.

Both selenium concentrations in whole body maternal fish and their eggs were measured. Figure 25 shows the relationship of egg selenium to whole body tissue selenium for wild brown trout collected for this study. This relationship is strong ($R^2 = 0.80$) with 80 percent of the egg selenium variability in wild fish explained by the selenium content of maternal tissue. It is important that this relationship be strong in order to confidently predict whole body tissue concentrations from egg concentrations. This is consistent with USEPA (2004) methods that developed translators for various tissue types. In this case, effects are related to egg selenium concentration, which, based on the current literature, is the best tissue for measuring a response for effects due to selenium.

4.3 Relationship of Egg Selenium Concentrations to Specific Effects Endpoints

Data for selenium in brown trout egg tissue are evaluated for effects on various endpoints for reproduction. Scatter plots and best-fit ordinary least squares regressions are used as an exploratory tool to further evaluate potential relationships. These regression relationships are not used as a final analysis tool or to determine biological significance of the regression because the distribution of the various effects endpoints (e.g., dependent variable) is typically skewed. Tests for normality were conducted and these data are not normally distributed nor are they log-normally distributed, which is one of the underlying assumptions of a least squares

regression analysis. Use of logistic regression approach as an analysis tool is presented in Section 4.4.

Figure 7 shows total number of eggs per female versus the individual female length. As expected, the smaller, wild fish produced fewer numbers of eggs relative to the larger, hatchery fish. Size may not be the only controlling factor. Overall health of the fish due to environmental and or chemical stressors may also play a role, as discussed previously. As shown in Figures 8 and 9, selenium concentrations in eggs or whole body adults, while varied, suggest relatively similar egg production, although when just examining wild fish egg production, there does appear to be a relationship of decreasing egg abundance with higher parental or egg selenium concentrations. Figure 26 shows wild brown trout egg abundance relative to selenium concentrations in egg tissue. For fish collected as part of this study, the relationship is poor ($R^2 = 0.2$); only a small percentage of the variability in observed egg production for wild fish is explained by egg selenium concentrations. Because fecundity can be related to the size of the female (i.e., heavier, longer fish, are indicative of older fish potentially in a more robust condition), these data were also normalized to 320 mm by dividing the total number of eggs by the length of the female parent. This value was then multiplied times 320. The red diamonds on Figure 26 illustrate that length-normalized fecundity does not reduce variability nor improve this weak relationship. Based on these data, it is difficult to suggest that fecundity is an important endpoint in assessing selenium effects in trout. The literature and field data will be used to further assess this endpoint through examination of literature-reported fecundity effects and observed population data from the Crow and Sage Creeks.

Figure 27 illustrates the percent hatch of brown trout fry versus egg selenium concentrations for hatchery and wild fish. No apparent relationship was observed. Figure 28 illustrates both hatchery and wild fish egg selenium concentration versus the percent of fry that achieved swim up. The regression analysis indicates a possible relationship ($R^2 = 0.49$) between the percent of fry that swim up and egg selenium concentration.

Larval fish growth, measured as the dry weight of the organism following the 15-day post swim up feeding trial, was also investigated as a potentially important endpoint. Figure 29 shows dry weights of larval fish versus egg selenium concentrations and two relationships, one including the hatchery fish and one including only the wild fish. Both relationships show decreased dry weight with increasing egg selenium concentrations. However, the strength of the relationship is moderate at best (i.e., its predictive ability) when including the hatchery fish ($R^2 = 0.66$) data and poor when using the wild fish data ($R^2 = 0.26$). The growth data provides a strong indication about how inclusion of hatchery fish in the analysis may provide misleading results. Note the difference in the R^2 values for the two relationships shown in Figure 29. Clearly, hatchery larvae are larger as a group when compared to wild fish. Clearly, egg selenium concentrations in hatchery fish are lower than any wild fish collected. However, many other factors including

differences in nutrition, relative physical stresses such as energy needed to forage, and size of wild versus hatchery fish adults may have effects that are independent of selenium exposures.

4.3.1 Survival/Mortality and Deformities

As discussed in Section 3, survival or mortality of eggs, alevins, and swim ups was measured throughout the study. For the purpose of the following analyses and those later presented for the logistic regression analysis, survival will be the endpoint used. The logistic regression analysis software requires that the effects variables be declining in their relationship to the exposure variables, and thus, will only correctly analyze, for example, survival data, not mortality data. To put these data in the same context for those analyses that precede the logistic regression analyses, survival data will be presented.

Survival was evaluated at several key points during the life stage: egg, hatch, swim up, at test termination following a 15-day feeding trial, and as overall survival. Of these, overall survival was initially evaluated as one of the endpoints more obviously correlated to egg selenium concentration. Figure 30 shows a scatter plot of this relationship, including wild and hatchery fish. Both hatchery fish and wild fish survival rates are variable on the lower end of the egg selenium range; however, it is clear from the data presented on Figure 30 that survival drops consistently to 30 percent or less at egg selenium concentrations above about 25 mg/kg dw. A third order polynomial regression provides a best fit model for these data ($R^2 = 0.543$) with egg selenium explaining about 54 percent of the variability in the overall survival endpoint when only wild fish are considered. The correlation coefficient is lower when hatchery fish are included.

Given the variability of the overall survival/mortality endpoint, as shown in Figures 14 and 15, and the influence of egg hatch on overall mortality/survival, the endpoint of survival/mortality from time of hatch to test end was investigated. Figure 31 illustrates this relationship including wild and hatchery fish. Review of this figure indicates a substantial decrease in survival between 20 and 25 mg/kg dw selenium in eggs. When both wild and hatchery fish are included in the 3rd order polynomial regression, the relationship is strong ($R^2 = 0.90$) with egg selenium concentrations explaining 90 percent of the variability in survival from hatch to test end. When considering only wild fish, the correlation coefficient R^2 remains essentially the same (0.89).

Figure 32 shows percent survival during the 15 day post swim up feeding trial versus egg selenium concentrations. When both wild and hatchery fish are included in the 3rd order polynomial regression, the relationship is strong ($R^2 = 0.96$) with egg selenium concentrations explaining 96 percent of the variability in survival during the post swim up feeding trial. When considering only wild fish, the correlation coefficient R^2 remains essentially the same (0.96). Of the survival terms, survival measured during the post swim up feeding trial presents the strongest relationship to egg selenium concentrations. As discussed later in Section 4.5, the

strong relationship of post-hatch alevins and fish survival to selenium concentrations in egg or ovary tissue is consistent with the findings of others.

Frequency and severity of deformities, or lack thereof, was also evaluated relative to egg selenium concentrations. Figure 33 shows the sum fraction of non-normal fish larvae relative to egg selenium. The 3rd order polynomial function provided the best fit line ($R^2 = 0.85$). The increase in non normal fish is evident between 20 to 25 mg/kg egg selenium. Recall that a similar break point was also observed for survival (Figures 31 and 32) which indicates an important break point in two related but different effects endpoints.

Figure 34 shows the sum fraction normal fish versus egg selenium concentration. The 3rd order polynomial function provided the best fit line ($R^2 = 0.87$) and a slightly stronger relationship than the fraction of non-normal fish. This may result due to the higher numbers of fish that were ranked as normal for each sample as opposed to having some level of deformity. Again, as egg selenium concentration increases, there is a distinct break in the data for the fraction of normal fish, which in this case, decreases substantially at egg selenium concentrations greater than 25 mg/kg dw.

Figure 35 shows the mean GSI score versus egg selenium concentration and again, a 3rd order polynomial provides the best fit line ($R^2 = 0.84$). The mean GSI score, fraction non-normal fish, and fraction normal fish provide very similar endpoints, albeit based on the same data, the derivation for each is quite different. Fraction non-normal and normal fish are simply frequency values, while the GSI score is frequency and severity of the deformity.

Figure 33 through 35 similarly illustrate an inflection point, similar to the survival endpoints where increased effects are occurring relative to egg selenium concentrations. Due to the strong relationship of the deformity frequency and GSI effects endpoints to egg selenium, additional analyses of the individual frequency for the different deformity categories will also be assessed in the subsequent section.

4.4 Logistic Analyses

Based on the results of the preliminary regression analyses described above, a subset of effects endpoints was selected for logistic-regression analysis² to determine dose-response

² Unlike traditional linear regression models, which assume equality of variance and normal distributions, the logistic regression model does not require nor have the same assumptions which can lead to Type I and Type II errors. Logistic regression has many analogies to OLS regression: logit coefficients correspond to b coefficients in the logistic regression equation, the standardized logit coefficients correspond to beta weights, and a pseudo R^2 statistic is available to summarize the strength of the relationship. Logistic regression does not assume linearity of relationship between the independent variables and the dependent, does not require normally distributed variables, does not assume homoscedasticity, and in general has less stringent requirements. Logistic regression finds the equation that best predicts the value of the Y variable for each value of the X variable. The Y variable is not directly measured; it is instead the probability of obtaining a particular value of a nominal variable.

relationships with egg selenium concentrations. Log-transformed egg selenium tissue concentrations (mg/kg dw) were related to growth (based on larval fish weight), survival (various stages), and deformities (various measures) (Table 8). USEPA's TRAP software was used to derive a best fit logistic regression model for each effect endpoint distribution. Summary statistics for each regression run, a graphic of the curve plotting the actual data and predicted curve, and EC_x for egg selenium residues based on the endpoint effect distribution are included in Appendix C for each of the models run.

4.4.1 Growth

Growth was measured in the post swim up feeding trial fish at the end of the 15 day period. These fish were carried through the test to the swim up stage. Twenty fish (or fewer if 20 were not available), for each sample, were fed for another 15 days to examine if there might be differences in the ability of swim ups to transition from endogenous to exogenous feeding. Morphological or physiological impairments could arise in young fish exposed to elevated selenium that may limit successful growth. Average growth of post feeding swim ups, as measured by dry weight, was related to egg selenium levels. Figure 37 shows this relationship for wild fish. Except for samples LSV-2C-003 and LSV-2C-010, 20 fish were included in this analysis for every location (Appendix A). The R^2 for this model is 0.208 and, as illustrated in Figure 36, the model fit does not resemble a dose response curve. TRAP software indicates a large standard error for the steepness of the slope. Table 8 shows the EC_{10} and EC_{20} values for growth along with the predicted confidence intervals. Slope steepness, or lack thereof combined with wide confidence intervals in the predictive ability of the model suggests a poor relationship of growth to egg selenium concentrations. The lack of fit may be due to several factors.

4.4.2 Survival

Survival was also measured as part of the post swim up study. Figure 37 shows the logistic regression curve fitted to survival data in the post swim up feeding trial versus log egg selenium concentrations. The R^2 for this model is high (0.96) and the fit of the predicted data to the observed data is good. Confidence intervals are also narrow for the predicted EC_{10} and EC_{20} values (Table 8). Recall that percent survival measured as part of the 15 day post swim up feeding trial had the best fit polynomial regression to egg selenium concentration as well. While the time frame of this endpoint is narrow, it is based on eggs resulting from maternal transfer of selenium, successful hatch, successful swim up, and survival of the larvae from the alevin endogenous feeding stage to exogenous feeding stage. The dose response curve reflected by this model illustrates a similar breakpoint in effects as previously mentioned.

Total survival through the duration of the test related to log egg selenium concentration is shown in Figure 38. The R^2 for this relationship is 0.31 and the error report of the logistic regression

model output from TRAP indicates a large standard error for the slope steepness. As illustrated, the fit of the data between observed and predicted values is low and the width of the confidence intervals about the EC_{10} or EC_{20} values is large, encompassing a large range of the curve. While the endpoint is relevant, the variability of the overall survival endpoint is not well suited for the logistic function, and thus its predictive ability for EC_x values is low.

Figure 39 shows the logistic regression for log egg selenium concentrations versus percent survival (hatch to test end). As illustrated, the predicted line fits the data well resulting in an R^2 of 0.89. The confidence intervals for the predicted EC_{10} and EC_{20} values are narrow and no errors (standard error was small and convergence was met) were reported in the TRAP software output (Table 8). Reduced variability of the survival term post-hatch likely strengthens this relationship and removes factors such as incomplete fertilization or egg viability, which could be affected by egg selenium concentrations as well as other factors. The strength of this model suggests it is a good predictor of survival effects post hatch due to selenium concentrations.

Based on the growth and survival models evaluated, the survival endpoints provide a much stronger measure of effects relative to egg selenium concentrations than does the growth endpoint. EC_x values for the three survival endpoints are very similar, although each endpoint represents different stages of development of young fish. Percent survival in the post swim up feeding study and percent survival from hatch to test end both appear to provide data that are strongly related to log egg selenium concentrations in terms of a dose response. Both predict similar EC_x values and narrow confidence intervals about the EC_x . Both provide biologically meaningful and relevant measures of effects, although survival during the 15 day post swim up feeding trial is more refined, as the variability of survival pre-swim up is eliminated. Survival from hatch to test end is an inclusive endpoint and encompasses the 15 day post swim up survival rate. The similarity of the two endpoints and the effects predicted based on their relationship to egg selenium concentration suggests that either endpoint may provide a suitable measure from which to gauge effects.

4.4.3 Deformities

The four primary deformity categories examined were: cranio-facial; skeletal; fin fold; and edema. Initial analyses were conducted to derive fractions or percentages of deformed fish relative to the total number of fish evaluated for an egg clutch. However, the TRAP software is sensitive to a declining effects response versus the exposure variable. For the purposes of fitting within the model framework, these data were structured in terms of the fraction of normal fish (number of normal fish/the total number of fish evaluated for an egg clutch). The GSI data are not structured for use in the TRAP model as GSI scores increase with increasing egg selenium. Summary data and graphics for deformity counts are presented in the earlier results section of this document and in Appendix C.

Figure 40 shows the logistic function for log egg selenium versus the fraction normal for cranio-facial deformity assessment. Scatter of the observed values relative to the predicted values reduced the fit of this model as well as the R^2 (0.70). The TRAP software error report of the logistic regression model output indicates a large standard error for the slope steepness. The confidence intervals for the predicted EC_{10} and EC_{20} values are slightly larger than that found for the total fraction normal endpoint.

Figure 41 shows the logistic function for log egg selenium versus the fraction normal for skeletal deformity assessment. Similar to the cranio-facial plot, the observed data do not fit the predicted model, although the R^2 value is higher than that of the craniofacial endpoint ($R^2 = 0.81$). The TRAP software error report of the logistic regression model output indicates a large standard error for the slope steepness, convergence was not reached at the maximum number of model iterations, and the steepness was at a maximum or minimum limit.

Figure 42 shows the logistic function for log egg selenium versus the fraction normal for finfold deformity assessment. The R^2 for this function is low (0.28) probably due to the lack of adequate data at the high end of the egg selenium concentration. Skeletal deformities for some samples could not be accurately assessed. The TRAP software error report of the logistic regression model output indicates a large standard error for the slope steepness, and convergence was not reached at the maximum number of model iterations. The errors associated with this model and poor fit reduce the utility of predicted EC_{10} and EC_{20} values.

Figure 43 shows the logistic function for log egg selenium versus the fraction normal for edema deformity assessment. The R^2 for this function is high (0.96) and the observed data fit the predicted model well. No errors were reported for this model from the TRAP software output. The predicted EC_{10} and EC_{20} values and their confidence intervals intersect the predicted dose response curve at the top end of the curve, with no inclusion of higher effects levels at the lower end of the curve.

Figure 44 illustrates the logistic regression for log egg selenium versus total fraction normal. This endpoint is a summed value proportion of the total number of normal fish per egg clutch to the total number of fish examined for that egg clutch. Because an individual fish could have more than one type of deformity and because it is a summation of fractions, it can be greater than 1 and in fact could be as high as 4. These data show a good fit to the predicted function and the confidence limits for the predicted ECs are narrow. Residual error is small (Appendix E) and the R^2 is high (0.88). A plot of the EC_{10} and EC_{20} and their confidence intervals over the predicted curve shows the EC values intersect the predicted line, bisecting the observed data where a clear break in effects has been previously discussed for other endpoints. The confidence limits are tight about the predicted EC_{10} and EC_{20} values suggesting not only a good fit, but a low variability as well. Figure 45 shows essentially the same relationship, only the

mean fraction normal was used as the dependent variable. The R^2 is the same as for sum fraction normal and the EC_{10} and EC_{20} values are nearly identical.

4.5 Predicted Effects Concentration for Brown Trout – Consistency with Literature

USEPA (2004) opted to use logistic regression analysis to define the dose-response relationship to derive its Draft chronic tissue-based value. The EC_{20} was used and defined as a reduction of 20 percent in the response observed at control. As presented above, both EC_{20} and EC_{10} values were derived using the TRAP software. In its 2004 Draft criterion document, USEPA provides the rationale for selection of the EC_{20} as the chronic value. USEPA states that the EC_{20} represents a low level of effect that is generally significantly different from the control (U.S. EPA 1999). Smaller reductions in growth, survival, or other endpoints only rarely can be detected statistically. Effect concentrations associated with such small reductions have wide uncertainty bands, making them unreliable for criteria derivation (USEPA 2004). In his work to develop a screening benchmark, Suter (1996) indicates that “the 20 percent figure was chosen because it is a little lower than the mean level of effect on individual response parameters observed at CVs, and it is a minimum detectable difference in population characteristics in the field.” In its revision of the 2004 Draft Selenium Criterion, USEPA is contemplating the use of EC_{10} s for long-term exposure criteria for tissue. Results of the analyses presented as part of this Site-specific laboratory study include both the EC_{10} and EC_{20} values.

For this study, an analysis consistent with the approach utilized by EPA for the 2004 draft criterion was utilized. For these analyses, “controls” are the response of fish from background locations. Based on the preliminary analysis presented above, the best-suited end points to calculate EC_x for brown trout egg selenium concentrations (dw) would be the sum fraction normal fish (causing less than a some percent reduction in the sum fraction of normal fish/total fish), and survival (causing less than a some percent reduction in survival post hatch). In total, logistic regression models were run for nine different endpoints including growth, survival, and deformities. For almost all (8 out of 9) of the models run, predicted EC_{10} or EC_{20} values were within 1 to 3 mg/kg dw egg selenium concentration of each other, respectively. The two models that appear to have a good overall fit with tight confidence intervals about the EC_{10} or EC_{20} predictions, and are biologically relevant, are the survival (hatch to test end) and total fraction normal endpoints. EC_{10} values for these endpoints are 17.67 and 19.33 mg/kg dw, respectively, while the EC_{20} values for these two endpoints are 21.63 and 21.7 mg/kg dw, respectively.

The endpoint for survival, based on hatch to test end, is consistent with the findings of Rudolph et al. (2008), who found a significant relationship of alevin mortality to egg selenium concentration. It has been suggested that selenium does not exert its toxic effects until a developing fish absorbs its yolk and accumulated selenium (Lemly 1997 and Holm et al. 2005 as cited in Rudolph et al. 2008). Hatchability of eggs is not affected by elevated selenium even

though there may be a high incidence of deformities in resultant larvae and fry, and many may fail to survive (Gillespie and Baumann 1986; Coyle et al. 1983).

Data for brown trout presented in this study showed a highly variable mortality rate prior to hatch, which may have been due several factors, including incomplete fertilization, disease, or reduced egg viability due to elevated selenium concentrations. However, the latter is not consistent with the review by Holm et al. (2005) who reports that although egg selenium is present in the yolk throughout development, it may affect larval development rather than egg development because it is mobilized to a greater degree after hatch.

Deformity frequency, as measured in this study based on fraction normal fish relative to the total number of fish assessed for deformities provides an endpoint that is consistent with the studies of Holm et al. (2005), Kennedy et al. (2000), Hardy (2005), Rudolph et al. (2008), Muscatello et al. (2006), and de Rosemond et al. (2005) in terms of cited developmental effects due to increased egg selenium concentrations. Table 9 shows the range of effects concentrations for the varying endpoints evaluated.

Hodson and Hilton (1983) and Lemly (1997) both suggest that developmental malformations are reliable indicators of chronic selenium toxicity to fish. Lemly (1997) described the sequence of selenium toxicity to larval fish: parental exposure, maternal deposition of selenium into eggs during vitellogenesis, and subsequent exposure during yolk resorption in developing larvae. Both the literature and the results of this study indicate that survival and developmental malformations of larval fish are clear and supportable endpoints for developing effects concentrations for fish. For brown trout, this study indicates that EC_{20} values would range from 21.6 to 21.7 mg/kg egg selenium for alevin survival (measured as hatch to test end) and larval deformities (measured as the sum fraction of normal fish), respectively.

4.6 Extrapolating Selenium Concentrations in Egg Tissue to Whole Body Tissue

In Section 4.2, the relationship of maternal whole body selenium concentrations to egg selenium concentrations was derived using wild brown trout collected for this study. The regression relationship has the form:

$$\text{Log}_{10}(y) = 1.1926(\text{Log}_{10}x) - 0.0071$$

The R^2 for whole body maternal selenium concentration relationship to selenium concentration in their respective eggs is 0.8. As stated previously, this relationship becomes important in this preliminary analysis in order to relate EC_x values derived for selenium concentrations in egg tissue back to selenium concentrations in whole body fish. Outside of this specific reproduction study, the larger body of data available for this site is for whole body tissue. Furthermore, whole body tissue concentration is a more practical endpoint to measure throughout the year than is

egg tissue. Based on an effects threshold value derived for selenium concentration in eggs and the site-specific relationship developed for whole body to eggs, a whole body tissue concentration of 13.35 mg/kg dw was derived based on the EC₂₀ for survival and a whole body tissue concentration of 11.27 mg/kg dw was derived based on the EC₁₀ for survival. Muscatello et al. (2006) reported whole body selenium EC₂₀ of 15.56 mg/kg dw for northern pike derived from the egg EC₂₀. Hardy (2005) reported a whole body NOEC for cutthroat trout of 11.37 mg/kg dw. Conversion of the Holm (2002) and Holm et al. (2003) ovary tissue selenium concentrations presented as chronic values from ovary to whole body using USEPA (2004) equations yields the following values: 19.96 mg/kg dw (rainbow trout), 16.06 mg/kg dw (rainbow trout), and 12.24 mg/kg dw (brook trout). Currently, the Draft National criterion recommends a value of 7.91 mg/kg dw.

These preliminary values of 11.27 or 13.25 mg/kg dw were derived for comparison purposes only to evaluate if the relationships derived for brown trout are consistent with values in the literature from other studies of maternal transfer. Based on the literature reviewed, the brown trout whole body value falls within the range of whole body tissue concentrations reported for other cold water species.

4.7 Data Adequacy

The critical question to be addressed for this study is whether or not the data adequately address the range of tissue concentrations in maternal parents which ultimately affects the offspring produced. Four key points address the adequacy of the data utilized for this study:

1. The goal was to capture adult brown trout with tissue selenium concentrations greater than 20 mg/kg dw, which represented the upper 90th percentile of the tissue selenium data for brown trout available when this study commenced. That goal was met as shown in the data presented earlier in this document.
2. Studies carried out with the brown trout and eggs collected yielded results that spanned a range of effects, including no or low effects and high and adverse levels of effects. In any toxicity study, being able to define the upper thresholds of effects is a critical component of the study. Effects were well defined for two important endpoints out of several that were evaluated, including survival and deformities for a sensitive life stage of brown trout, which are consistent endpoints defined in the literature for other trout species.
3. The distribution of effects and exposure data indicates that there are no large gaps in the data and that relationships between effects and egg selenium concentrations can be defined with confidence.

4. Collection of more fish with tissue concentrations higher than those collected for this study will not improve the study because effects are defined near the middle of the egg selenium concentrations which correspond to whole body selenium tissue concentrations that are lower than the upper whole body tissue concentrations (i.e., greater than 20 mg/kg dw).

4.8 Dose Response Analysis Update

Throughout this report, reference has been made to two particular effects thresholds, the EC₁₀ and EC₂₀. The brown trout data presented in the Draft Final Brown Trout Laboratory Reproduction Studies Conducted in Support of Development of a Site-Specific Selenium Criterion (NewFields 2009) were submitted to USEPA for use in their derivation of the National Criterion. The first draft of the Interpretive Report (August 2010) proposed an EC₂₀ as the SSC for this project based on the brown trout data. USEPA provided formal comments (December 21, 2010) on the Draft Interpretive Report and the criterion proposed and suggested that some alternative evaluations may be practical. USEPA's review of the brown trout data indicates agreement with the selection of the endpoint for survival (hatch to test end). Further, the USEPA has made it clear that they intend to propose an EC₁₀ in their Draft National Criterion, which includes the brown trout data developed as part of this study. In their comments, EPA stated that, for this project, the EC₁₀ is a more appropriate endpoint than the proposed EC₂₀ in developing a proposed SSC for the Smoky Canyon Site. Their primary rationale is that as a bioaccumulative pollutant that accumulates in fish tissue, concentrations in fish tissue are more stable over time than aqueous selenium concentrations. This stability may lead to concentrations that are just below the criterion for extended periods of time.

The TRAP software used to derive the logistic regression conducted throughout this brown trout study also includes two additional non-linear models, threshold sigmoidal and piecewise linear models. USEPA's comment letter illustrated an investigation of each of these models relative to the logistic model used as part of the brown trout studies presented above, and found that the projected EC_x values are likely conservative. As part of the USEPA's evaluation, another alternative examined exclusion of data points that exceeded 30 mg/kg dw in eggs, due to the fact that effects were already occurring between 15 and 30 mg/kg dw. This approach was investigated as a means of optimizing the model output. By eliminating the three highest data points, the logistic model is able to focus on the region of interest (i.e., between 15 and 30 mg/kg dw egg selenium). Using this approach, the logistic model run using log-transformed exposure data (egg selenium concentrations) versus survival (hatch to test end) results in a model with a R² = 0.99 (Figure 46). This improved model results in an EC₁₀ equal to 20.8 (95 percent LCL – 19.89, 95 percent UCL – 21.83) mg/kg dw egg selenium and an EC₂₀ equal to 23.1 (95 percent LCL – 22.37, 95 percent UCL – 23.77) mg/kg dw egg selenium.

Confidence intervals derived for the estimated EC_x values are also tight about the estimates and the standard error of the model is low.

The effects concentration for fraction normal fish was also re-evaluated to assess whether the previously derived EC_{10} and EC_{20} values were similarly affected. Using the same approach as presented above, the revised EC_{10} and EC_{20} values for fraction normal fish are 22 and 23.4 mg/kg dw egg selenium, respectively.

Using the brown trout regression model presented in Section 4.6 as a translator between egg selenium and whole body selenium concentrations, the resulting whole body values corresponding to the egg EC_{10} and EC_{20} values are 12.91 and 14.1 mg/kg dw, respectively.

5.0 SUMMARY AND CONCLUSIONS

The effects of maternal selenium transfer in wild brown trout were evaluated as part of this study. Eggs from wild female brown trout collected from different locations with varying selenium exposure levels were used to assess a number of reproductive endpoints as part of this study. Initially, the data were plotted and reviewed for any obvious relationships and patterns. In the initial review, a consistent breakpoint was identified where egg selenium concentrations were contrasted with reproduction test endpoints. These observed relationships are consistent with expected dose-response relationships.

Moving forward from these initially-defined relationships, adult whole body and egg selenium concentrations were considered the independent variables in a regression-based analysis approach. The focus of the analysis was narrowed to focus on egg selenium concentration versus growth, survival, and deformity endpoints. Logistic regression was used to develop dose-response relationships and predict egg selenium concentrations related to different effects (EC_x). Post-hatch survival and total deformity frequency (fraction normal) were found to be the most biologically relevant endpoints exhibiting dose response relationships and concurrence of observed data to predicted values. The predicted post-hatch survival EC_{20} was 21.63 (95 percent LCL – 17.77, 95 percent UCL – 26.32) mg/kg dw egg selenium, while the EC_{10} for this endpoint was 17.68 (95 percent LCL – 13.44, 95 percent UCL – 23.25) mg/kg dw egg selenium. For deformities, the sum fraction normal endpoint, the EC_{20} was 21.7 (95 percent LCL – 18.09, 95 percent UCL – 26.02) mg/kg dw egg selenium, while the EC_{10} for this endpoint was 19.33 (95 percent LCL – 15.07, 95 percent UCL – 24.79) mg/kg dw egg selenium.

Consistent with comments received from the USEPA relative to derivation of the effect concentrations previously presented, a revised dose-response model was utilized that included using the same logistic regression model, and eliminated egg selenium concentrations above the already defined effects threshold (i.e., >30 mg/kg dw). Using this approach allowed the model to be more focused on the region where effects occur to brown trout. This improved model results in an EC_{20} equal to 23.1 (95 percent LCL – 22.37, 95 percent UCL – 23.77) mg/kg dw egg selenium and an EC_{10} equal to 20.8 (95 percent LCL – 19.89, 95 percent UCL – 21.83) mg/kg dw.

Egg tissue concentrations may not be a practical endpoint for routine monitoring because it requires collection of samples during a narrow window of opportunity during autumn of each year. Collection of adult tissue samples is more practical since it can be done with lower effort and with less stringent constraints for the schedule under which sampling must be done. Therefore, the high correlation between selenium in eggs and adult tissue was used to identify an adult tissue concentration that corresponds to the revised EC_{20} and EC_{10} estimates based on

egg selenium. The resulting adult (whole body) tissue concentration for this Site based on the revised EC₂₀ is 14.1 mg/kg dw and based on the revised EC₁₀ it is 12.9 mg/kg dw.

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TABLES

**Table 1
Monitoring Locations, Coordinates, and Counts for October 2007 Sampling in Support of the Brown Trout Laboratory Toxicity Studies**

Location	Reach	Reach Boundary	Easting	Northing	Date	Fishing Time (Sec)	# caught and released		# retained		Total Brown Trout
							>230<300	>300	males	females	
Reference											
MPC-1	Montepelier Creek	Downstream	485847	4690589	10/23/2007	3750	3	0	0	0	3
		Upstream	485591	4693679							
STUMP-3	Stump Creek	Downstream	493711	4738229	10/26/2007	2378	7	0	7	9	23
		Upstream	488968	4743637							
Upstream of Sage Creek											
CC-75	Crow Creek upstream of Wells Canyon	Downstream	486291	4710439	10/22/2007	1008	6	1	7	5	19
		Upstream	486334	4710260							
CC-150-Redd	Crow Creek upstream of Deer Creek	Downstream	487299	4713431	10/24/2007	1422	10	6	1	8	25
		Upstream	487204	4713372							
CC-350	Crow Creek downstream of Deer Creek	Downstream	489434	4715598	10/22/2008	4122	26	7	5	8	44
		Upstream	489460	4715334	10/24/2008						
Hoopes Spring and Sage Creek											
LSV-2c	Lower Sage Creek downstream of Hoopes Spring	Downstream	491398	4720245	10/25/2007	2865	56	44	10	32	142
		Upstream	491293	4720575							
Downstream of Sage Creek											
CC-1A	Crow Creek downstream of Sage Creek	Downstream	493433	4719137	10/22/2007	2034	2	0	3	4	12
		Upstream	493280	4719057							
CC-3A	Crow Creek downstream of Sage Creek and CC-1A	Downstream	494938	4720410	10/24/2007	3848	18	0	3	2	33
		Upstream	494648	4720085							
Totals							128	58	36	68	301

Table 2
Summary of Field-Measured Water Quality Parameters Collected in October 2007 for Sampling in Support of the Brown Trout Laboratory Toxicity Studies

Stream	Location	Date	pH (SU)	Specific Conductance (umhos/cm)	Temp. (°C)	Dissolved Oxygen (mg/L)	ORP (mV)
Reference							
Montpelier Creek	MC-1	10/23/2007	7.70	437	2.1	12.36	152.1
Stump Creek	SC-3	10/26/2007	8.19	610	9.16	11.98	127.2
Upstream of Sage Creek							
Crow Creek	CC-75	9/2/2006	7.93	426	7.11	8.22	NM
		5/8/2007	8.29	532	7.01	11.59	176.5
		8/23/2007	8.04	551	10.32	10.65	131.8
		10/22/2007	8.21	572	5.45	11.37	82.5
		5/12/2008	8.05	397	6.31	10.55	161.2
	CC-150	9/3/2006	7.58	399	7.94	9.26	NM
		5/9/2007	8.34	444	9.42	9.32	204.3
		8/24/2007	8.35	437	8.12	11.84	194.3
	CC-350	5/12/2008	8.53	361	10.44	10.00	164.3
		8/31/2006	8.89	458	14.8	10.57	NM
		5/8/2007	8.47	572	14.27	11.23	104.6
		8/23/2007	8.61	643	17.1	11.16	90.1
		10/22/2007	8.79	573	6.24	13.06	55.6
		5/13/2008	8.44	431	15.04	9.05	202.9
Hoopes Spring and Sage Creek							
Hoopes Spring	HS	9/8/2006	7.46	461	11.77	5.46	NM
		5/14/2007	7.60	503	11.84	5.21	51.9
		8/24/2007	7.49	473	11.89	6.32	78.2
		5/17/2008	7.33	302	12.02	6.08	201.1
	HS-3	9/6/2006	7.43	489	10.37	7.33	NM
		5/12/2007	8.46	484	16.57	7.22	89.5
		8/28/2007	8.25	460	17.1	9.41	85.4
		5/17/2008	8.38	289	17.05	9.01	152.7
Sage Creek	LSV-2C	9/6/2006	8.56	478	18.62	6.72	NM
		5/12/2007	8.35	498	11.09	8.90	19.7
		8/28/2007	8.37	458	14.27	10.68	-37.1
		10/25/2007	8.31	462	8.17	11.36	124.4
		5/17/2008	8.40	283	18.72	8.92	188.6
	LSV-4	9/5/2006	7.81	454	9.49	7.95	NM
		5/9/2007	8.50	402	16.54	7.60	125.4
Downstream of Sage Creek							
Crow Creek	CC-1A	9/1/2006	8.37	590	13.4	9.08	NM
		5/10/2007	8.44	591	10.39	9.14	156.5
		8/25/2007	8.43	577	10.61	12.62	29.3
		10/22/2007	8.31	572	3.06	12.73	129.2
		5/14/2008	8.09	358	7.39	9.80	161.2
	CC-3A	9/4/2006	7.91	561	11.32	8.94	NM
		5/11/2007	8.47	601	9.48	9.28	161.2
		8/26/2007	8.20	583	11.26	10.12	19.8
		10/24/2007	8.08	616	4.02	11.55	183.7
		5/15/2008	8.42	370	13.42	10.49	219.6

Note: All field monitoring data collected through Spring 2008 is presented on this table. Green shading highlights October 2007 monitoring data.

Table 3
Water Quality Parameters Measured During the October 2007 Sampling
in Support of the Brown Trout Laboratory Toxicity Studies

Stream	Location	Date	Alkalinity (mg/L)	Hardness (mg/L as CaCO ₃)	Sulfate, SO ₄ (mg/L)
Reference					
Montpelier Creek	MPC-1	10/23/2007	95.9J	138J	13.6J
Stump Creek	SC-3	10/26/2007	195J	222J	39.2J
Upstream of Sage Creek					
Crow Creek	CC-75	9/2/2006	215J	199	24.6
		5/8/2007	230J	175	23.5
		8/23/2007	214J	152J	25.5
		8/23/2007-dup	127J	212J	23.4
		10/22/2007	208J	233J	32.6J
		5/12/2008	196	208	24.7
	5/12/2008-dup	195	205	24.7	
	CC-150	9/3/2006	175J	153	11.8
		5/9/2007	209	152	14.4
		8/24/2007	179J	204J	12.6
		5/12/2008	192	199	17.8
	CC-150 REDD	10/24/2007	190J	202J	13.1J
	CC-350	8/31/2006	200J	187	16.1
		5/8/2007	203J	149	18.4
		8/23/2007	134J	197J	17.8
		10/22/2007	124J	224J	18.5J
5/13/2008	197	211	23.7		
Hoopes Spring and Sage Creek					
Hoopes Spring	HS	9/8/2006	194	209	47.2
		9/8/2006-dup	194	199	46.8
		5/14/2007	197	149	48.7
		8/24/2007	154J	231J	46.5
		5/17/2008	196	232	49.8
	HS-3	9/6/2006	205J	220	39.1
		5/12/2007	196	150	44.3
		8/28/2007	202J	222J	42.0
5/17/2008	197	223	43.2		
Sage Creek	LSV-2C	9/6/2006	201J	206	38
		5/12/2007	209	156	41.0
		8/28/2007	205J	223J	40.3
		10/25/2007	198J	237J	42.8J
	5/17/2008	199	218	38.7	
	LSV-4	9/5/2006	203J	187	29
5/9/2007	186	129	31.2		
Downstream of Sage Creek					
Crow Creek	CC-1A	9/1/2006	162J	160	24.7
		5/10/2007	140	151	22.4
		5/10/2007-dup	187	155	28.5
		8/25/2007	202J	213J	35.0
		10/22/2007	199J	229J	34.9J
	5/14/2008	202	221	31.4	
	CC-3A	9/4/2006	211J	195	17.5
		9/4/2006-dup	206J	191	19.3
		5/11/2007	206	152	29.9
		8/26/2007	195J	212J	35.8
10/24/2007		195J	228J	36.5J	
5/15/2008	197	216	32.1		

Note: All field monitoring data collected to date is presented on this table. Green shading highlights October 2007 monitoring data.

Table 4
Summary of Total Selenium Concentrations Measured in Surface Waters in Support of the Brown Trout Laboratory Toxicity Studies

Stream	Location	Date	Selenium Concentrations	
			Total Selenium (mg/L)	Dissolved Selenium (mg/L)
Reference				
Montpelier Creek	MPC-1	10/23/2007	0.0002UJ	0.0002UJ
		10/23/2007-dup	0.0002UJ	0.0002UJ
Stump Creek	SC-3	10/26/2007	0.0002UJ	0.0002R
Upstream of Sage Creek				
Crow Creek	CC-75	9/2/2006	0.00053	0.00057
		5/8/2007	0.00047J	0.00046J
		8/23/2007	0.00033J	0.00033J-
		8/23/2007-dup	0.00079J	0.0004J-
		10/22/2007	0.00055J-	0.0002UJ
		5/12/2008	0.0012	0.0012
	CC-150	9/3/2006	0.00062	0.00067
		5/9/2007	0.00083J	0.00092J
		8/24/2007	0.00059J	0.00068J-
		5/12/2008	0.0018	0.0014
		5/12/2008-dup	NM	NM
	CC-150 REDD	10/24/2007	0.0015J-	0.0011J-
	CC-350	9/1/2006	0.00083	0.00082
		5/8/2007	0.00084J	0.0011J
		8/23/2007	0.0002UJ	0.00026J-
		10/22/2007	0.0003J-	0.0002UJ
		5/13/2008	0.001	0.00089
Hoopes Spring and Sage Creek				
Hoopes Spring	HS	9/8/2006	0.0174	0.0174
		9/8/2006-dup	0.0174	0.0168
		5/14/2007	0.0301J	0.0205J
		8/24/2007	0.0242J	0.0214J-
	5/17/2008	0.0296	0.0273	
HS-3	9/6/2006	0.0108	0.0092	
	5/12/2007	0.0198J	0.018J	
	8/28/2007	0.0158J	0.0161J-	
Sage Creek	LSV-2C	5/17/2008	0.0223	0.026
		9/6/2006	0.0095	0.0093
		5/12/2007	0.0135J	0.0135J
		8/28/2007	0.0144J	0.0143J-
		10/25/2007	0.0384J-	0.0279J-
5/17/2008	0.0145	0.0141		
Downstream of Sage Creek				
Crow Creek	CC-1A	9/1/2006	0.0029	0.0027
		5/10/2007	0.0016J	0.0012J
		5/10/2007-dup	0.0025J	0.002J
		8/25/2007	0.0014J	0.0022J-
		10/22/2007	0.0028	0.0013J-
	5/14/2008	0.0032	0.0029	
	CC-3A	9/4/2006	0.003	0.0029
		9/4/2006-dup	0.0029	0.0027
		5/11/2007	0.0013J	0.0014J
		5/11/2007-dup	NM	NM
		8/26/2007	0.0011J	0.0018J-
		8/26/2007-dup	NM	NM
		10/24/2007	0.0017J-	0.001J-
5/15/2008		0.0036	0.0026	

Note: All field monitoring data collected through Spring 2008 is presented on this table. Green shading highlights October 2007 monitoring data. J - Estimated, NM-Not measured, Bold concentrations are those currently exceeding the State standard for total selenium (0.005 mg/L).

Table 5
Monitoring Locations, Coordinates, and Counts for November 2007 Sampling in Support of the Brown Trout Laboratory Toxicity Studies

Reach	Location	Reach Boundary	Easting	Northing	Date	Fishing Time (Sec)	# Caught		Retained Ripe		Total Brown Trout	Total # Egg Samples Submitted	Notes
							>230<300 mm	>300 mm	males	females			
Reference													
Stump-1 and 3	Stump Creek	Downstream	493804	4737886	11/16/2007	2,594	NC	26	0	2	28	0	captured, checked for ripeness, released >300 mm, 2 females appeared ripe, no eggs expressed
		Upstream	493711	4738229									
	Stump Creek	Downstream	490825	4742505									
		Upstream	490605	4742633									
Upstream of Sage Creek													
CC-150	Nate property to CC-150 and upstream of CC-150	Downstream	487299	4713431	11/12/2007	8,347	43	18	7	13	81	9	9 out of 13 females expressed eggs
		Upstream	487178	4712722	11/15/2007								
CC-350	Crow Creek downstream of Deer Creek	Downstream	489569	4715684	11/15/2007	3,441	25	10	5	3	43	3	
		Upstream	489491	4715286									
Hoopes Spring and Sage Creek													
LSV-2c	Lower Sage Creek downstream of Hoopes Spring	Downstream	491298	4719723	11/14/2007	7,090	42	41	17	21	121	14	Stopped counting at ~40, too many fish. Estimate >300 fish >230mm caught
		Upstream	491283	4720601									
Downstream of Sage Creek													
CC-1A	Crow Creek downstream of Sage Creek	Downstream	493433	4719137	11/13/2007	7,129	32	37	2	0	69	0	
		Upstream	493136	471904									
Between CC-3A and CC-1A, upstream of CC-1A	Crow Creek downstream of Sage Creek	Downstream	494676	4720149	11/13/2007	7,129	32	37	2	0	69	0	
		Upstream	494036	4719245									
		Downstream	493433	4719137									
		Upstream	493136	471904									
CC-3A and downstream	Crow Creek downstream of Sage Creek and CC-1A	Downstream	495176	4720411	11/16/2007	3,184	0	28	3	0	31	0	
		Upstream	494874	4720281									
Totals							142	160	34	39	373		

Note: Note all trout collected or retained were spawned. Some females which appeared initially to be ripe, based on external features or even an initial expulsion of eggs did not provide eggs when stripping was conducted.

Table 6
Adult Female Brown Trout Length and Whole-Body Selenium (mg/kg dw) Data

Location	Sample ID	Total length (mm)	Wet weight (g)	Whole-body Selenium Concentration (mg/kg dw)	Egg Selenium Concentration (mg/kg dw)
Hatchery Fish	Saratoga National Fish Hatchery (SC-001)	498	1,855 ^a	3.6	0.76
	Saratoga National Fish Hatchery (SC-002)	420	1,089 ^a	4.1	0.94
	Saratoga National Fish Hatchery (SC-003)	520	2,072 ^a	3.7	0.83
	Saratoga National Fish Hatchery (SC-004)	562	3,350 ^a	4.3	0.92
	Saratoga National Fish Hatchery (SC-005)	558	2,927 ^a	3	1.2
	Saratoga National Fish Hatchery (SC-006)	439	1,111 ^a	3.1	1.2
	Saratoga National Fish Hatchery (SC-007)	449	1,561 ^a	2.7	1
	Saratoga National Fish Hatchery (SC-008)	494	1,927 ^a	2.5	0.96
Hatchery Fish (Second Set)	Spring Creek Fish Hatchery (SPC-001)				0.73
	Spring Creek Fish Hatchery (SPC-002)				
	Spring Creek Fish Hatchery (SPC-003)				
	Spring Creek Fish Hatchery (SPC-004)				
	Spring Creek Fish Hatchery (SPC-005)				
	Spring Creek Fish Hatchery (SPC-006)				
Wild Fish	Crow Creek (CC-150-009)	324	315	8.4	12.8
	Crow Creek (CC-150-011)	342	351	5.6	8.4
	Crow Creek (CC-150-012)	317	269	6.7	8.5
	Crow Creek (CC-150-013)	332	376	5.9	8.4
	Crow Creek (CC-150-015)	313	281	6	9.1
	Crow Creek (CC-150-016)	391	621	7	7.5
	Crow Creek (CC-150-017)	265	178	5.6	6.6
	Crow Creek (CC-150-018)	308	279	4.7	6.9
	Crow Creek (CC-150-020)	310	318	7.2	6.2
	Crow Creek (CC-350-006)	370	475	9.2	14
	Crow Creek (CC-350-007)	350	416	5.5	6.9
	Crow Creek (CC-350-008)	335	341	8.5	9.5
	Lower Sage Creek (LSV2C-002)	304	280	8.9	12.8
	Lower Sage Creek (LSV2C-003)	300	260	13.8	40.3
	Lower Sage Creek (LSV2C-004)	290	260	17.9	36
	Lower Sage Creek (LSV2C-005)	294	250	13.6	26.8
	Lower Sage Creek (LSV2C-006)	346	420	17.2	26.9
	Lower Sage Creek (LSV2C-007)	315	290	6.7	18.6
	Lower Sage Creek (LSV2C-008)	296	230	9.6	17.7
	Lower Sage Creek (LSV2C-010)	311	314	22.6	38.8
	Lower Sage Creek (LSV2C-012)	360	434a	7.2	13.2
	Lower Sage Creek (LSV2C-016)	300	260	9.2	13.4
	Lower Sage Creek (LSV2C-017)	341	310	13.2	20.5
Lower Sage Creek (LSV2C-019)	330	364	8.6	12.5	
Lower Sage Creek (LSV2C-020)	280	241	11.3	11.2	
Lower Sage Creek (LSV2C-021)	307	317	20	28.1	

^aData not found or measured. Weights were estimated from the relationship between the wet weights of field collected fish and wet weights measured by CAS ($r^2 = 0.963$).

Table 7
Summary of Brown Trout Tissue Selenium Data Collected in Support of the
Brown Trout Laboratory Toxicity Studies

Stream	Location	Date	Brown Trout (mg/kg dw)			
			Count	Mean	Min	Max
Upstream of Sage Creek						
Crow Creek	CC-150	9/3/06	4	5.83	4.75	7.71
		5/9/07	3	8.67	8.00	10.00
		8/24/07	2	5.20	4.30	6.10
		10/30/07	1	5.40	5.40	5.40
		11/15/07	9	6.34	4.70	8.40
	5/13/08	3	9.82	8.61	11.80	
	CC-350	8/31/06	1	7.40	7.40	7.40
		5/8/07	0	na	na	na
		8/23/07	3	5.43	4.60	6.00
		11/15/07	3	7.73	5.50	9.20
5/13/08		0	na	na	na	
Sage Creek						
Sage Creek	LSV-2C	9/6/06	6	19.45	16.00	22.82
		5/12/07	4	12.78	8.50	22.20
		8/28/07	9	22.67	10.80	33.30
		11/14/07	20	14.03	6.20	23.80
		5/16/08	6	20.25	11.40	29.60
Downstream of Sage Creek						
Crow Creek	CC-1A	9/1/06	3	9.76	8.15	11.86
		5/10/07	2	9.05	7.40	10.70
		8/25/07	11	9.95	6.30	14.80
		10/30/07	1	8.70	8.70	8.70
		5/14/08	5	17.54	16.40	18.30
	CC-3A	9/4/06	3	11.15	9.14	14.34
		5/11/07	4	9.20	7.50	12.70
		8/27/07	13	11.25	7.80	15.60
		10/24/07	4	6.50	5.30	8.00
		5/14/08	4	15.38	15.00	15.80

Note: All field monitoring data collected through Spring 2008 is presented on this table.
Green shading highlights October/November 2007 monitoring data.

na-Not Applicable.

Table 8
Effect Concentration (EC_x) Values for Egg Selenium Tissue Residues Versus Different
Biological Endpoints for Brown Trout

Biological Endpoints	Effect Concentration (EC _x)			R ²
	50	20	10	
Growth and Survival				
Growth	46.23	33.79	28.13	0.21
95% LCL	27.05	22.84	13.09	
95% UCL	79.01	50.00	60.44	
15-Day Post Survival	34.73	24.52	20.00	0.96
95% LCL	33.11	22.26	17.37	
95% UCL	36.42	26.99	23.02	
Total Survival	24.83	21.43	19.66	0.31
95% LCL	19.27	13.60	10.75	
95% UCL	32.00	33.77	35.98	
Survival Hatch -Test End	30.52	21.63	17.68	0.89
95% LCL	27.58	17.77	13.44	
95% UCL	33.78	26.32	23.25	
Deformities				
Fraction normal	26.43	21.70	19.33	0.88
95% LCL	23.94	18.09	15.07	
95% UCL	29.19	26.02	24.79	
CF fraction normal	26.04	22.31	20.37	0.68
95% LCL	22.06	15.91	12.79	
95% UCL	30.75	31.27	32.47	
SD fraction normal	25.13	23.30	22.29	0.81
95% LCL	19.89	15.01	12.68	
95% UCL	31.76	36.18	39.20	
FD fraction normal	27.65	23.22	20.96	0.28
95% LCL	24.27	17.85	14.30	
95% UCL	31.49	30.19	30.73	
ED fraction normal	26.98	21.23	18.45	0.96
95% LCL	25.31	18.96	15.82	
95% UCL	28.76	23.77	21.52	

CF-Cranio-facial deformity

SD-Skeletal deformity

FD-Fin deformity

ED-Edematous Tissue deformity

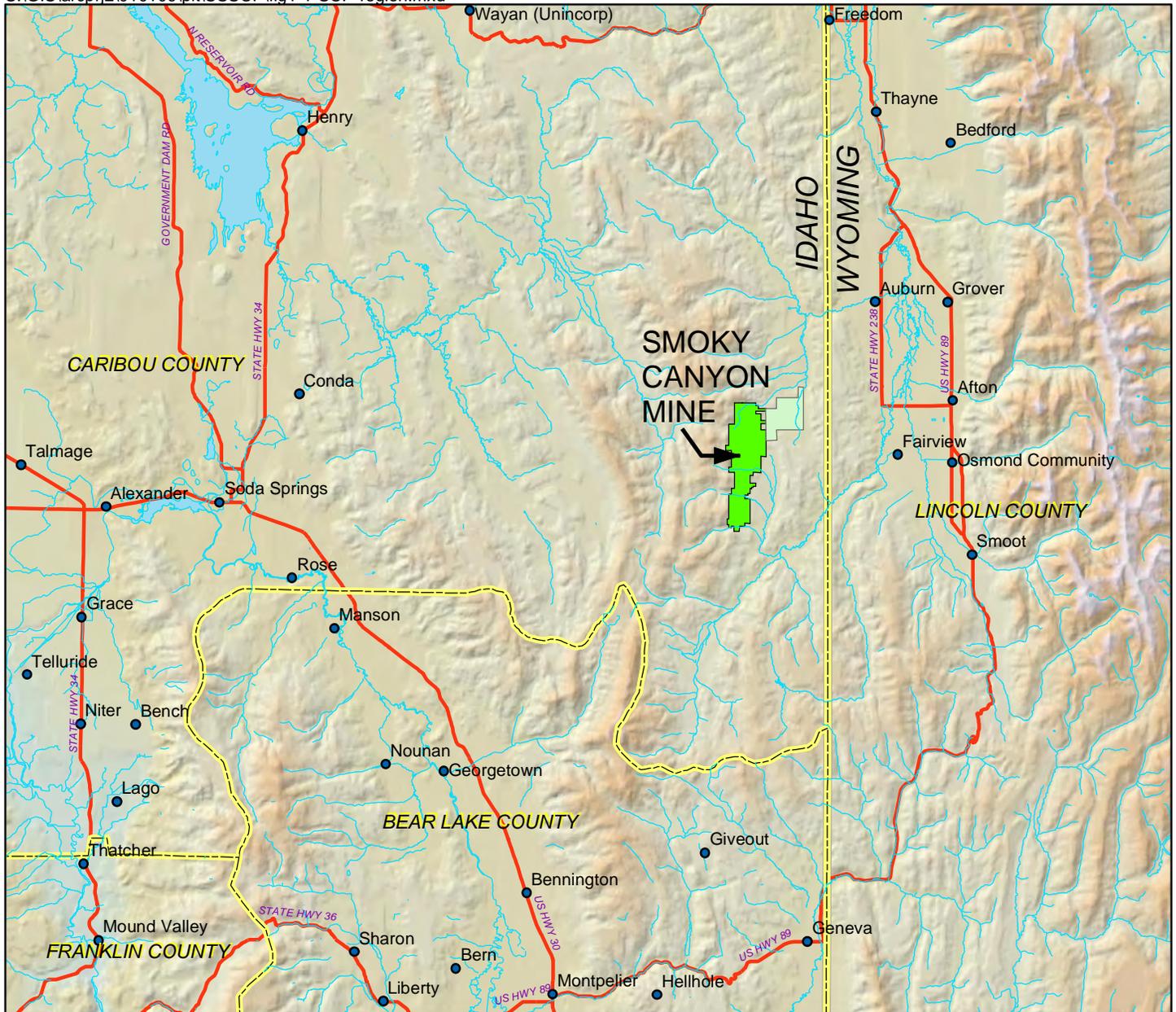
Table 9
Summary of Toxicity Studies that Evaluated Selenium Toxicity to Embryo/Larvae Resulting from Maternal Transfer

Species	Reference	Adult Exposure	Endpoint	Tissue	Selenium Concentration (µg/g dry weight)			
					NOEC	LOEC	EC ₁₀	EC ₂₀
Bluegill	Bryson et al. 1984	Field	Larval mortality	Ovary	--	<49	--	--
	Bryson et al. 1985a	Field	Hatchability/swim-up	Ovary	>9.1	--	--	--
		Field	Hatchability/swim-up	Ovary	-	<30	--	--
	Bryson et al. 1985b	Field	Hatchability/swim-up	Ovary	>14.8	--	--	--
		Field	Hatchability/swim-up	Ovary	>9.2	--	--	--
	Gillespie and Baumann 1986	Field	Larval edema	Ovary	--	<38.6	--	--
	Doroshov et al. 1992	Lab	Larval edema	Ovary	3.94	21.1	15	17
		Lab	Larval edema	Egg	8.55	25.81	21	23
	Coyle et al. 1993	Lab	Larval mortality	WB	7	16	8	8.5
		Lab	Larval mortality	Ovary	20	35	24	27
Lab		Larval mortality	Egg	22.5	41.3	22	26	
Hermanutz et al. 1996	Mesocosm	Larval edema	WB	4.4	21.8	--	--	
	Mesocosm	Larval edema	Ovary	17.3	69	--	--	
Fathead minnow	Ogle and Knight 1989	Lab	Reproduction	WB	>7.5	--	--	--
		Lab	Reproduction	Ovary	>10.92	--	--	--
	Schultz and Hermanutz 1990	Mesocosm	Larval edema/lordosis	Ovary	-	<39.3	-	-
	GEI Consultants	Field	Larval deformities/edema	WB	--	--	33	--
Field		Larval deformities/edema	Ovary	--	--	45 ^a	--	
Brook trout	Holm et al. 2005	Field	Larval deformities	Egg	>20	-	20 (EC06)	-
Cutthroat trout	Kennedy et al. 2000	Field	Larval deformities/ mortality	Egg	>21	-	-	-
	Hardy 2005	Lab	Larval deformities/ mortality	WB	>11.37	-	-	-
		Lab	Larval deformities/ mortality	Egg	>16.04	-	-	-
	Rudolph et al. 2008	Field	Larval deformities	Egg	20.6	46.8	-	-
Field		Alevin mortality	Egg	-	-	17	23	
Rainbow trout	Holm et al. 2005	Field	Larval deformities	Egg	17	25	26	29
Northern pike	Muscatello et al. 2006	Field	Larval deformities	Egg	3.8	31.28	20.4	33.55
White sucker	de Rosemond et al. 2005	Field	Larval deformities	Egg	-	-	26 (EC13)	-

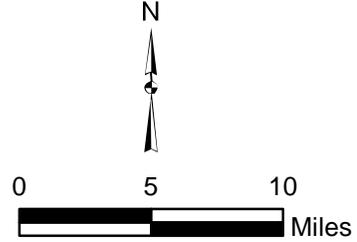
Original table Source: Selenium Tissue thresholds - Tissue Selection Criteria, Threshold Development Endpoints, and Potential to Predict Population or Community Effects in the Field (NAMC 2009).

^a An ovary-based EC₁₀ of 44.6502 µg/g was estimated from a whole-body EC₁₀ of 33.07 µg/g based on the whole-body-ovary selenium relationship for fathead minnows (FHM) presented in GEI Consultants (2008): FHM [Se] dw WB = 0.75826*(FHM ovary [Se] dw) - 0.78645.

FIGURES



Smoky Canyon Mine



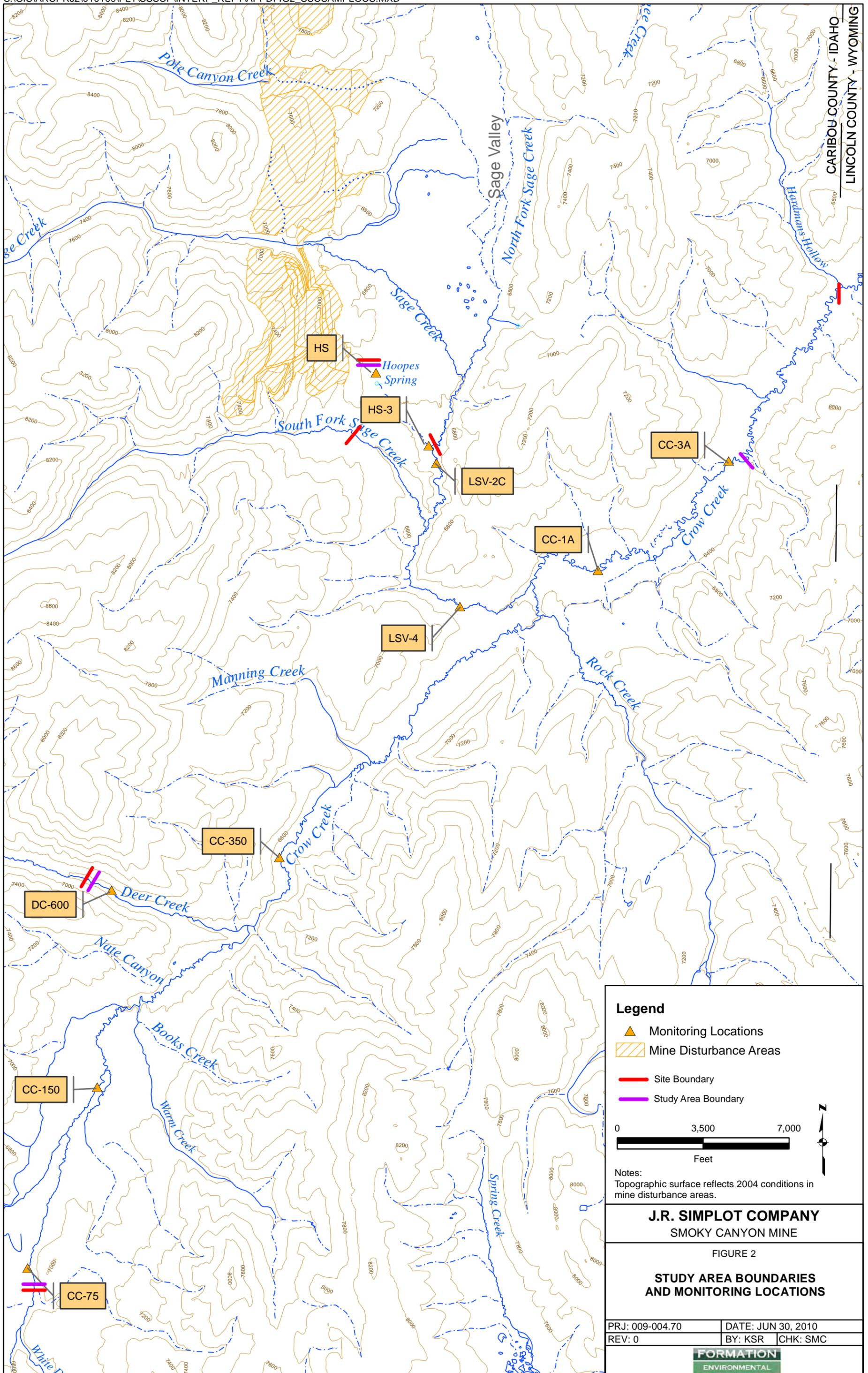
J.R. SIMPLOT COMPANY
SMOKY CANYON MINE

FIGURE 1

**LOCATION OF THE
SMOKY CANYON MINE**

PRJ: 0442-004-900.70	DATE: MAY. 05, 2011
REV: 0	BY: RCR CHK: SMC





CARIBOU COUNTY - IDAHO
LINCOLN COUNTY - WYOMING

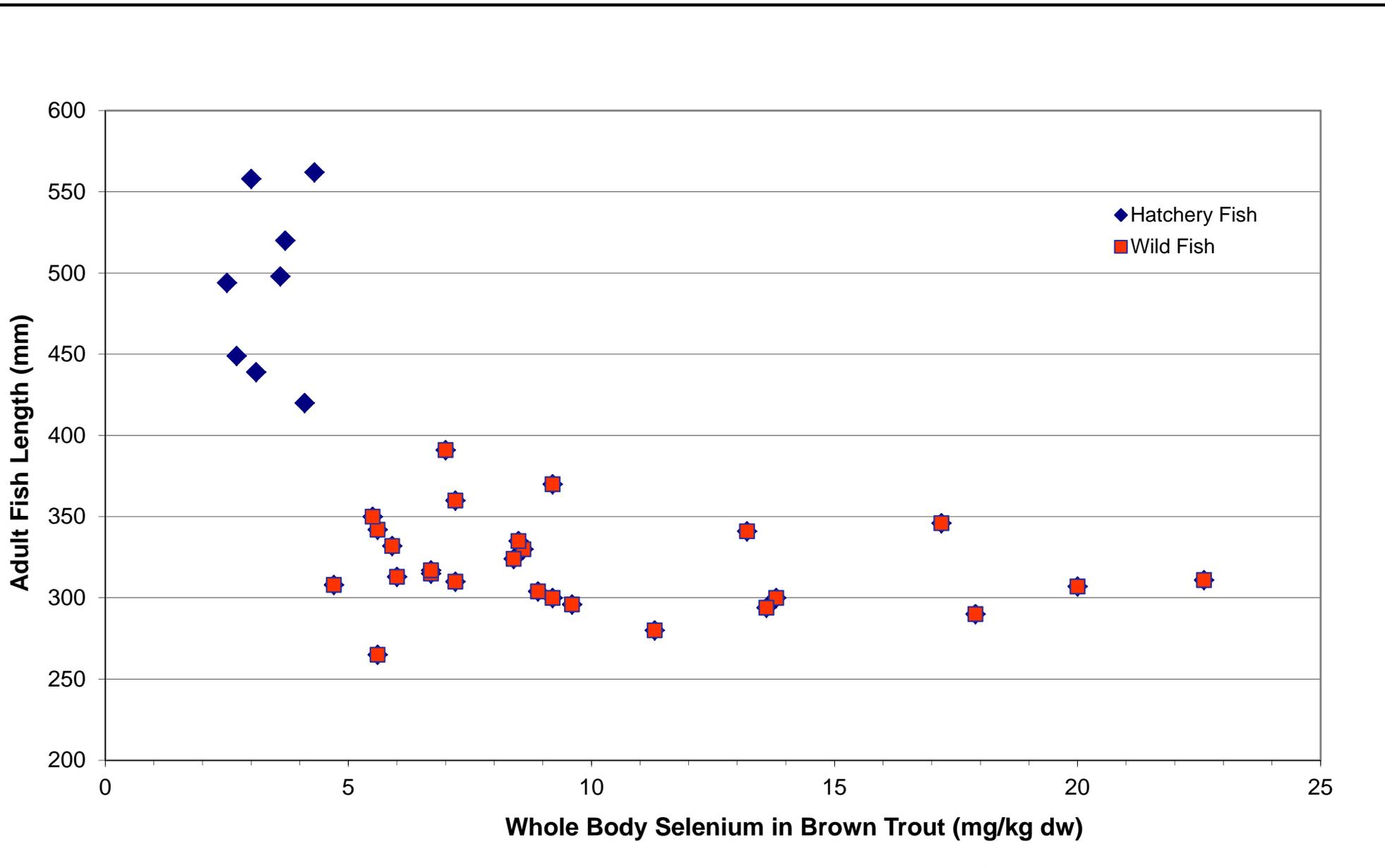


Figure 3
Whole Body Selenium Concentration Versus Fish Length

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC



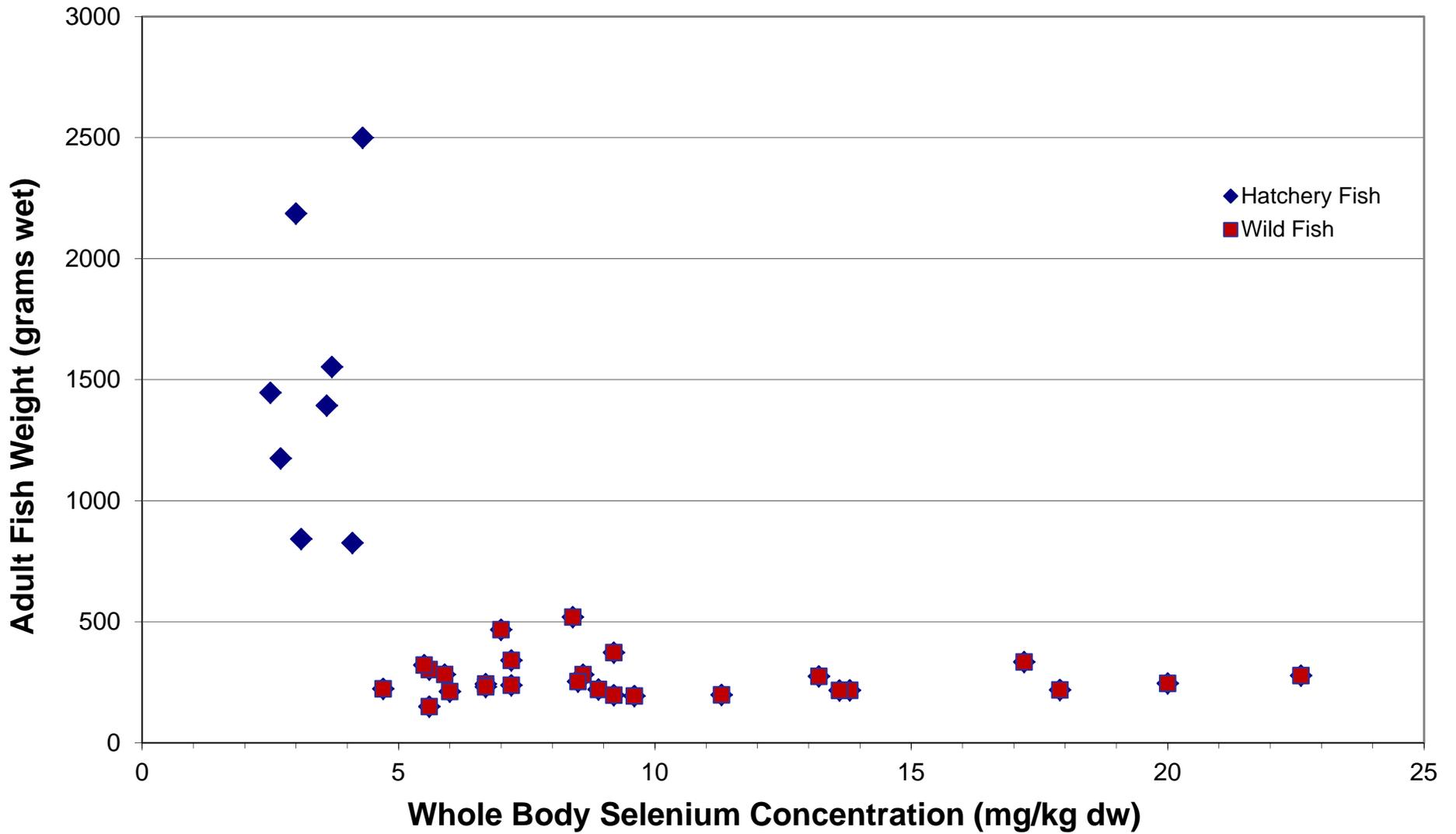


Figure 4
Whole Body Selenium Concentration Versus Adult Fish Weight

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

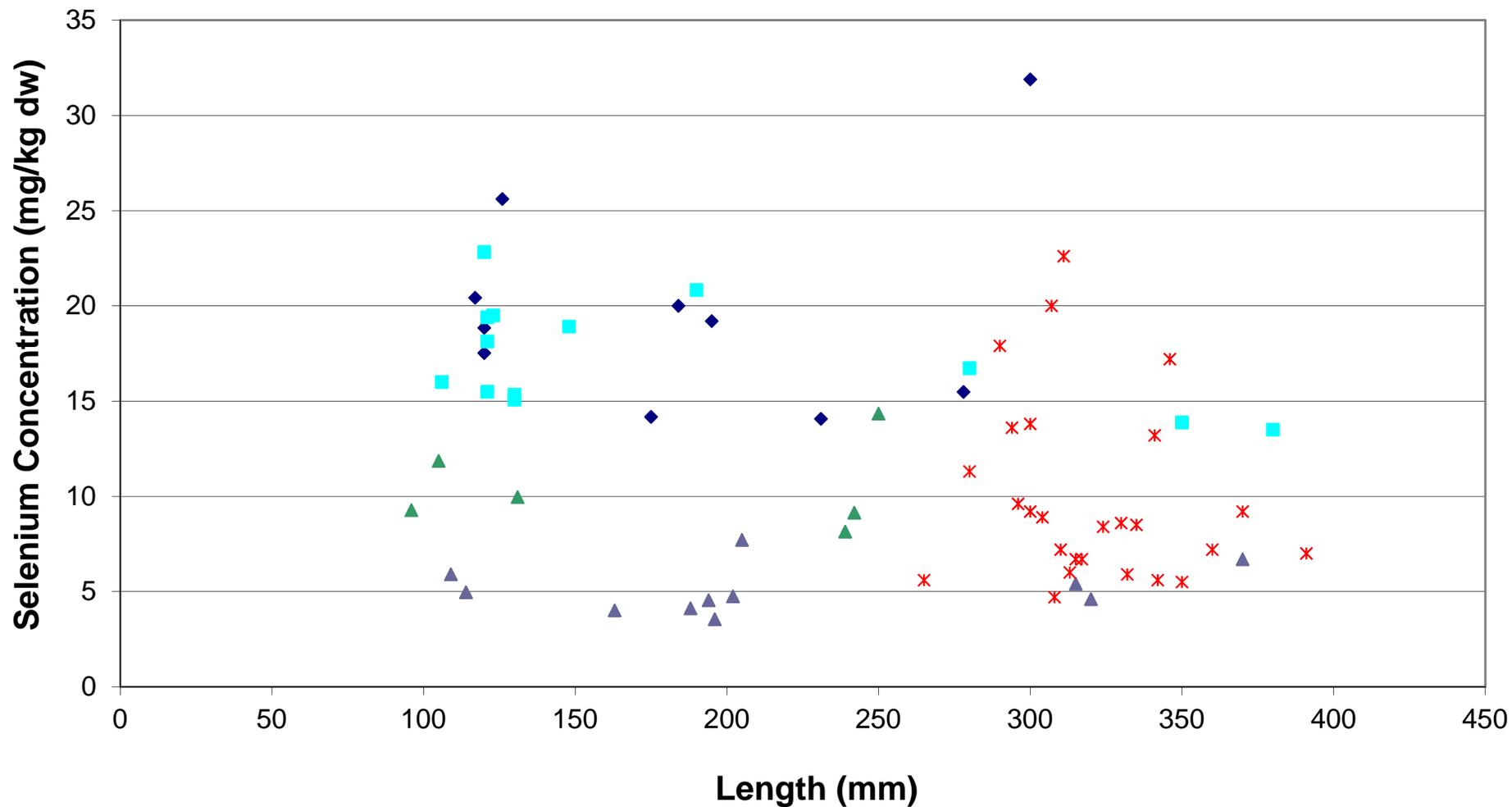
DATE: June 2011

REV: 1

BY: SMC

CHK: SMC





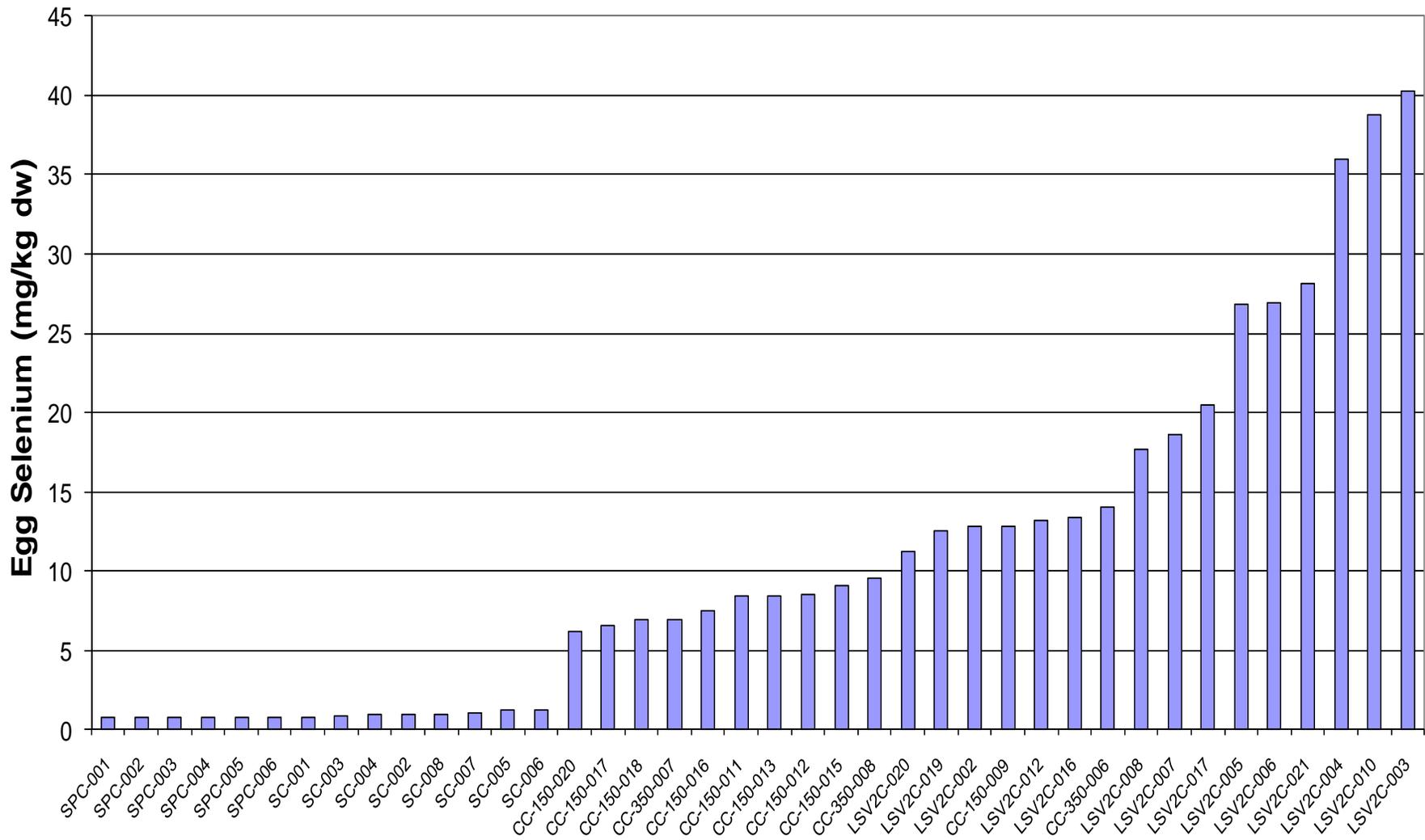
◆ Hoopes ■ Sage Ck ▲ Crow Ck d/s ▲ Background × Adult BT

Figure 5
Brown Trout Length Versus Whole Body Selenium Concentration

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC

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Note: The order of locations, as represented on the X axis, is in order of egg selenium concentration.

Figure 6
Brown Trout Egg Selenium Concentration by Location

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC



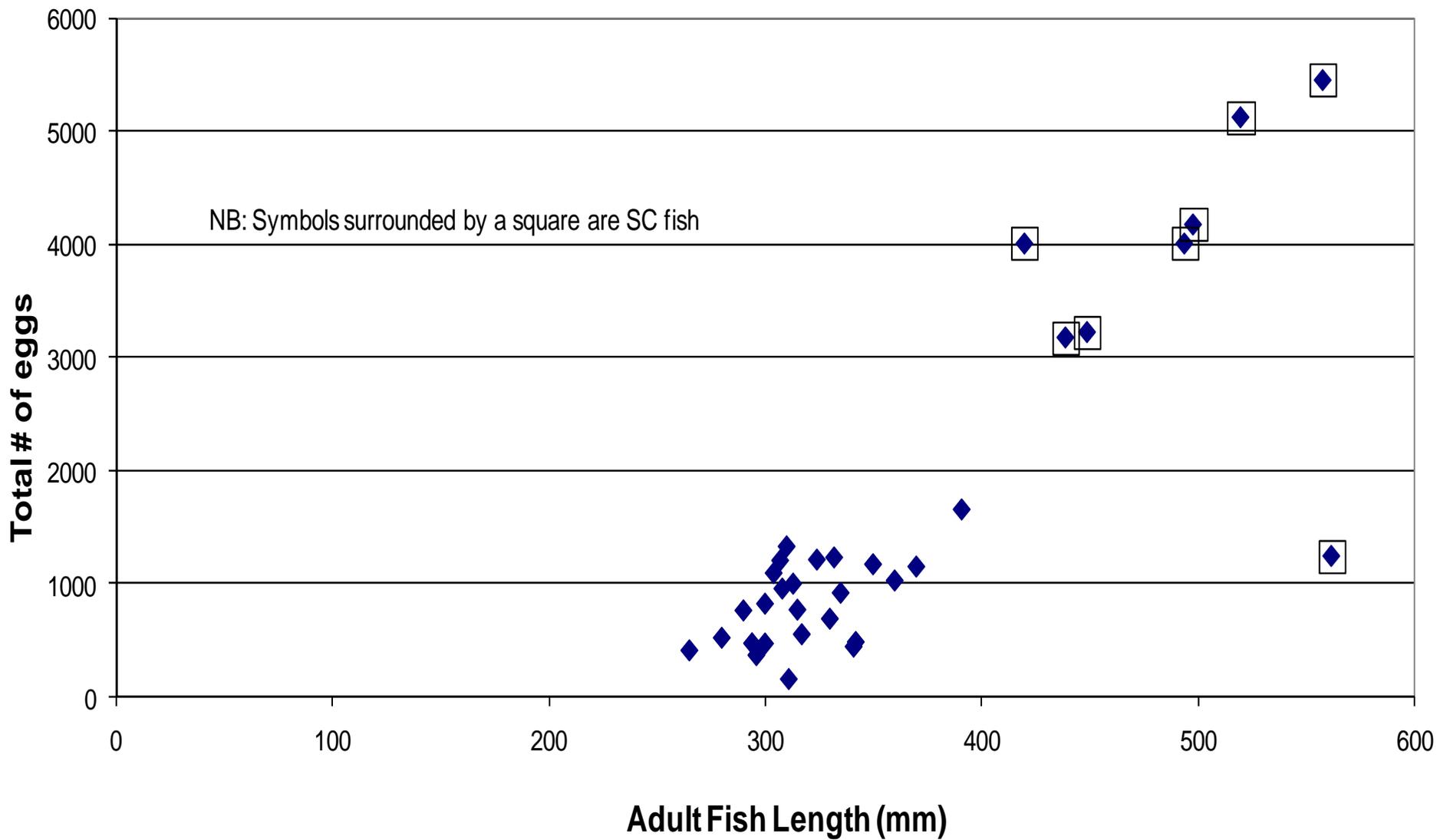


Figure 7
Comparison of Adult Fish Size (Length) and the Total Number of Eggs Produced

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC



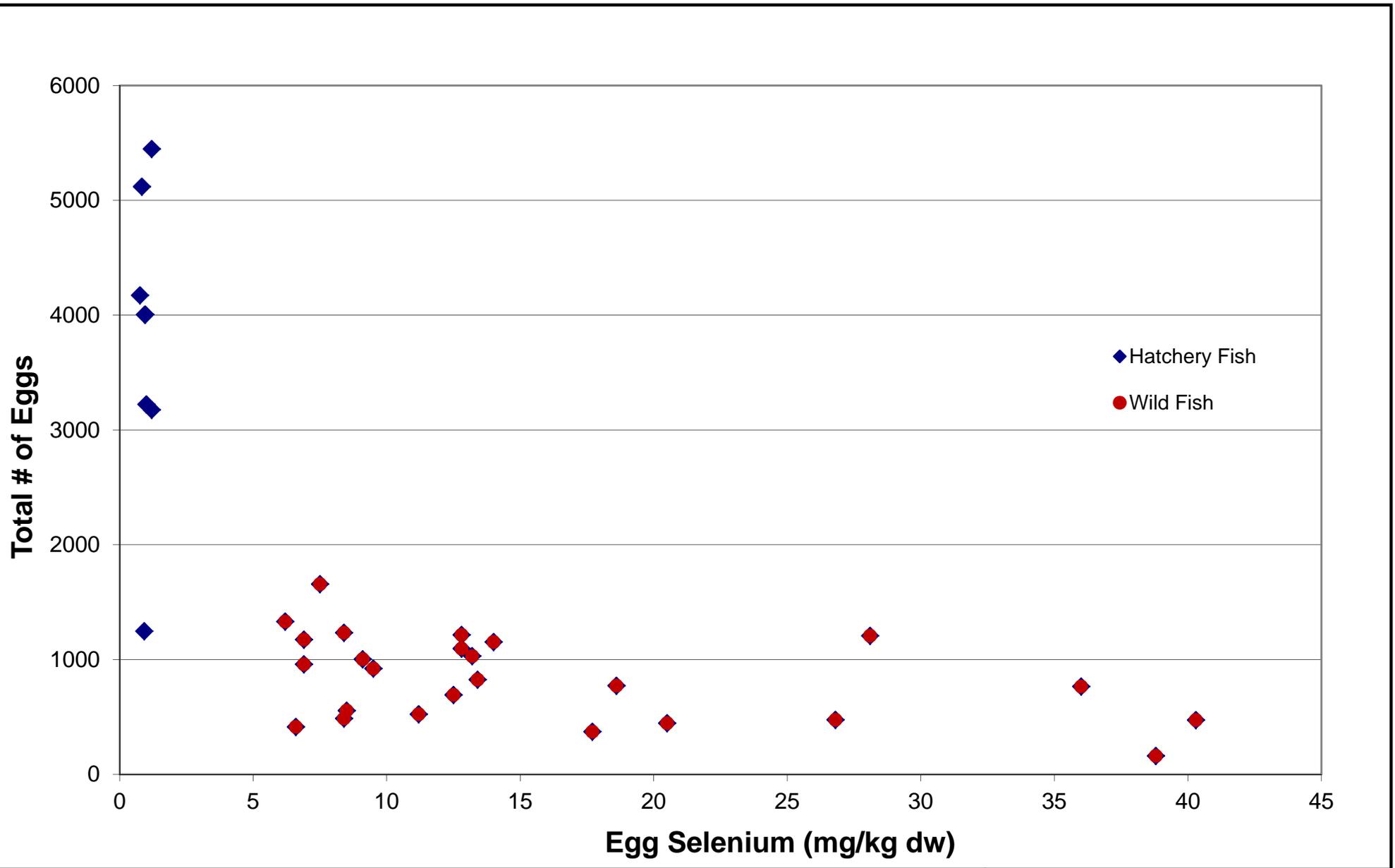


Figure 8
Egg Selenium Concentration Versus Total Egg Abundance

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC



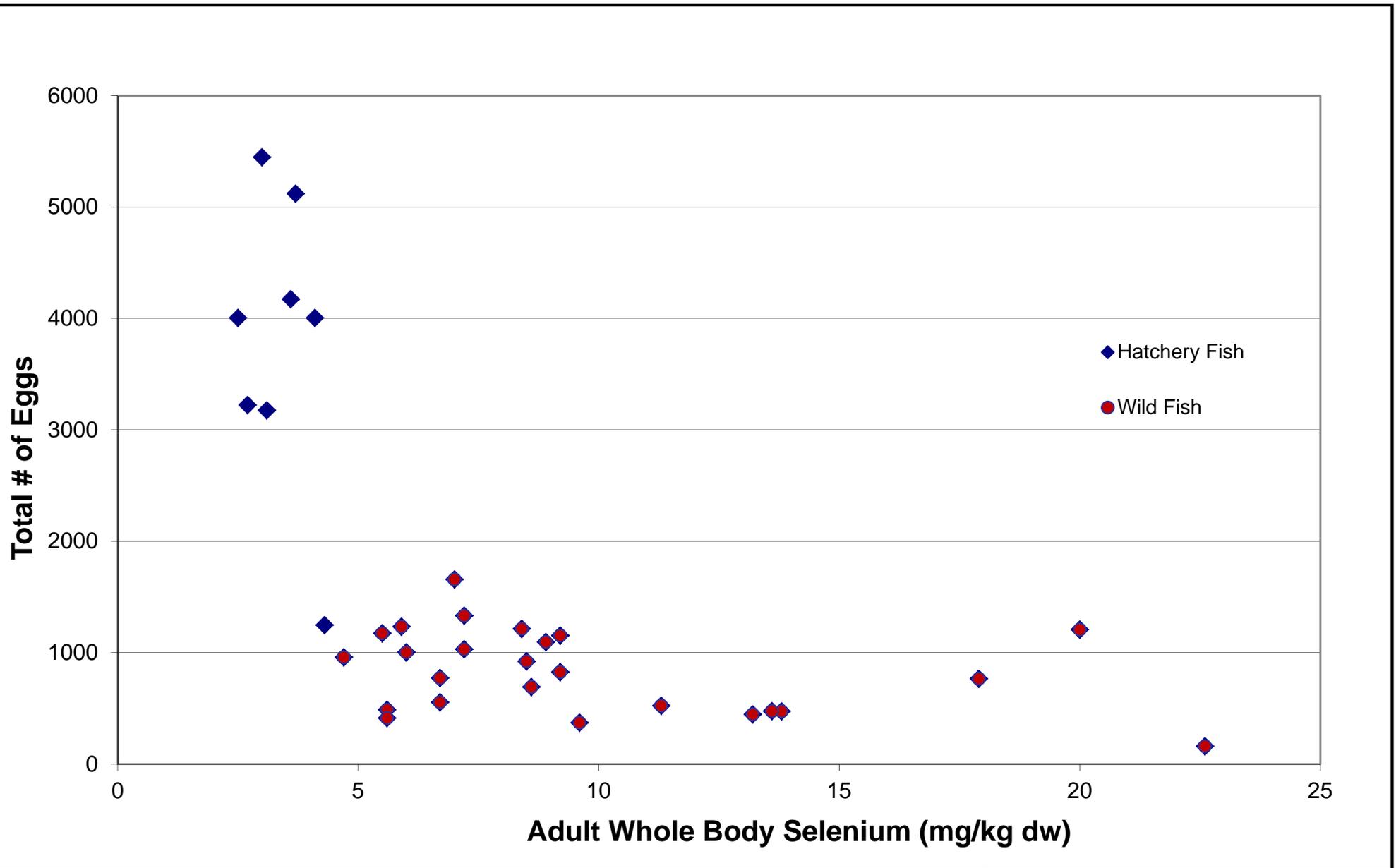


Figure 9
Adult Whole Body Selenium Concentration Versus Total Egg Abundance

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

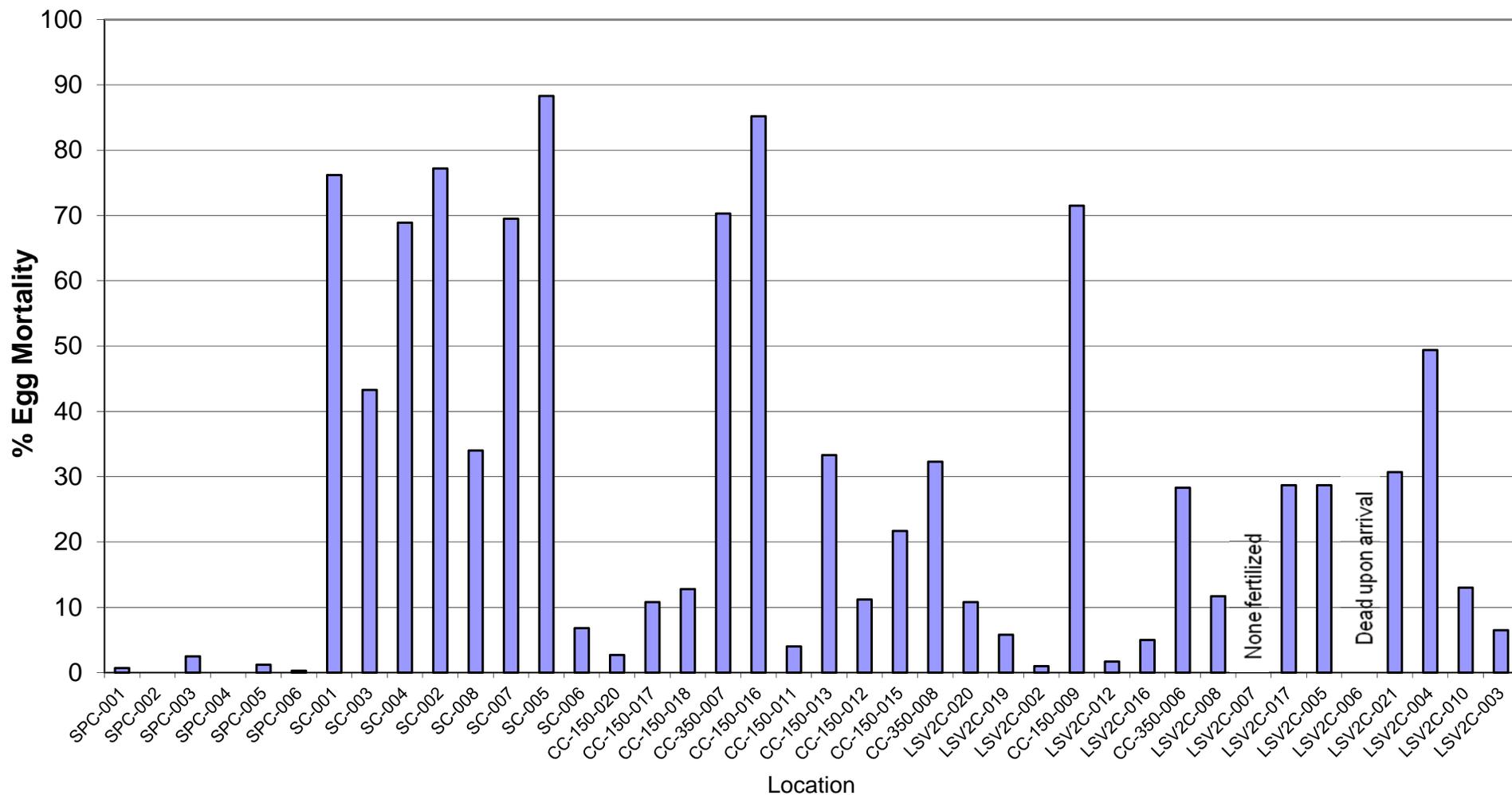
DATE: June 2011

REV: 1

BY: SMC

CHK: SMC





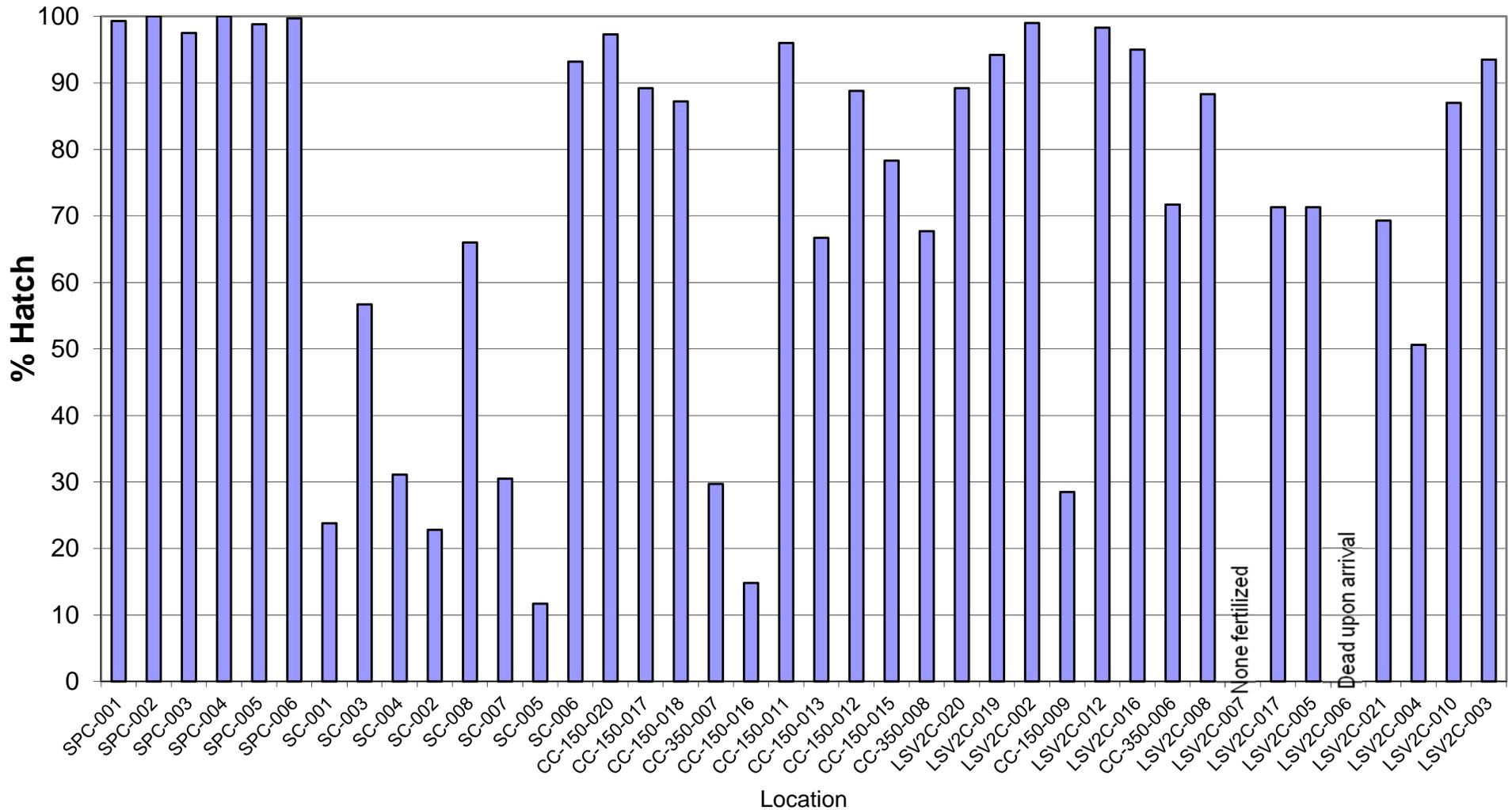
Note: The order of locations, as represented on the X axis, is in order of egg selenium concentration.

Figure 10
Percent Egg Mortality by Location

J.R. Simplot Company
Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC





Note: The order of locations, as represented on the X axis, is in order of egg selenium concentration.

Figure 11
Percent Hatch (or Percent Survival at Hatch) of Alevins

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011
REV: 1	BY: SMC CHK: SMC



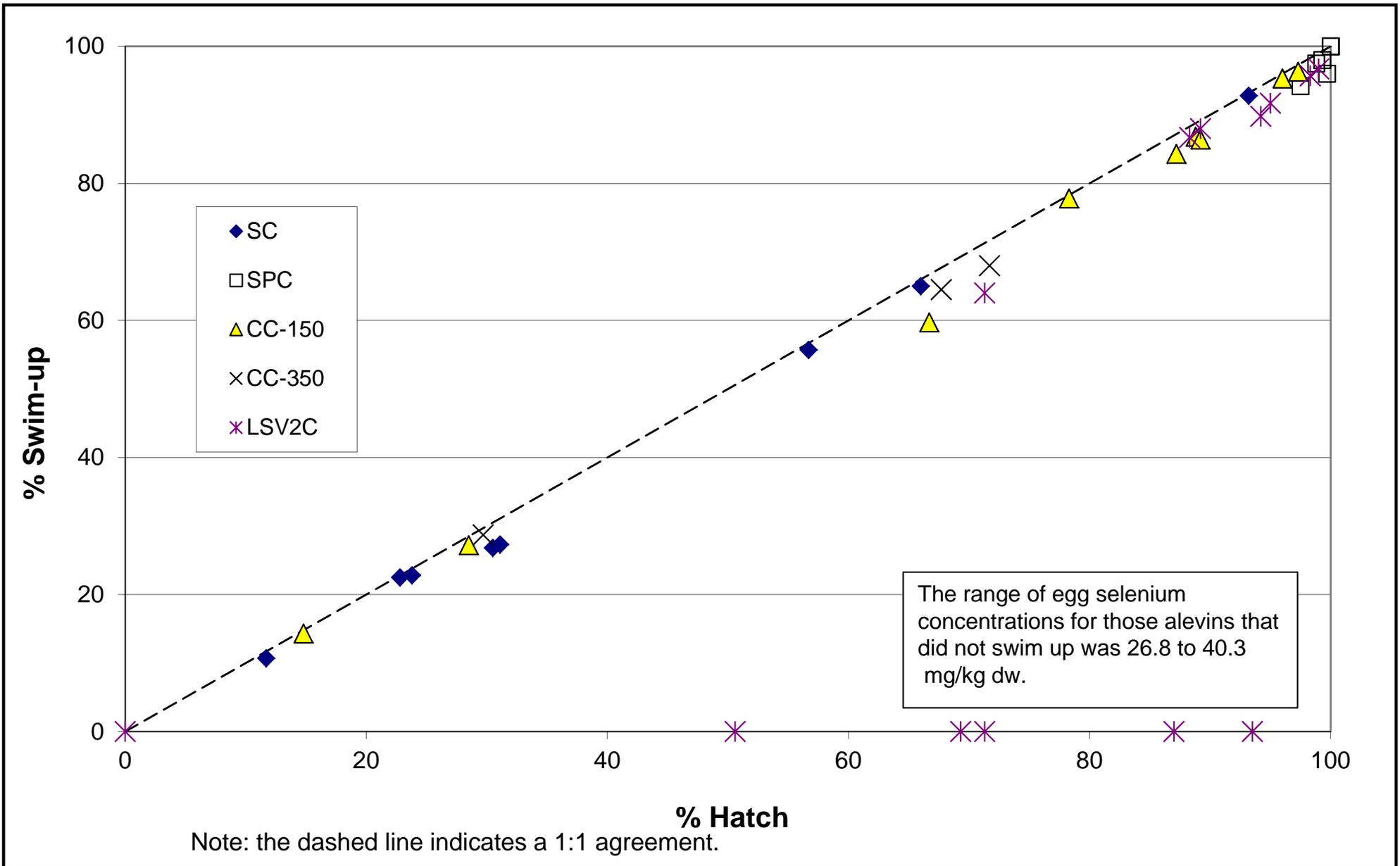


Figure 12
Relationship Between the Percentage of Eggs that Hatched and the Percentage of Fish that Reached Swim-Up

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011
REV: 1	BY: SMC CHK: SMC



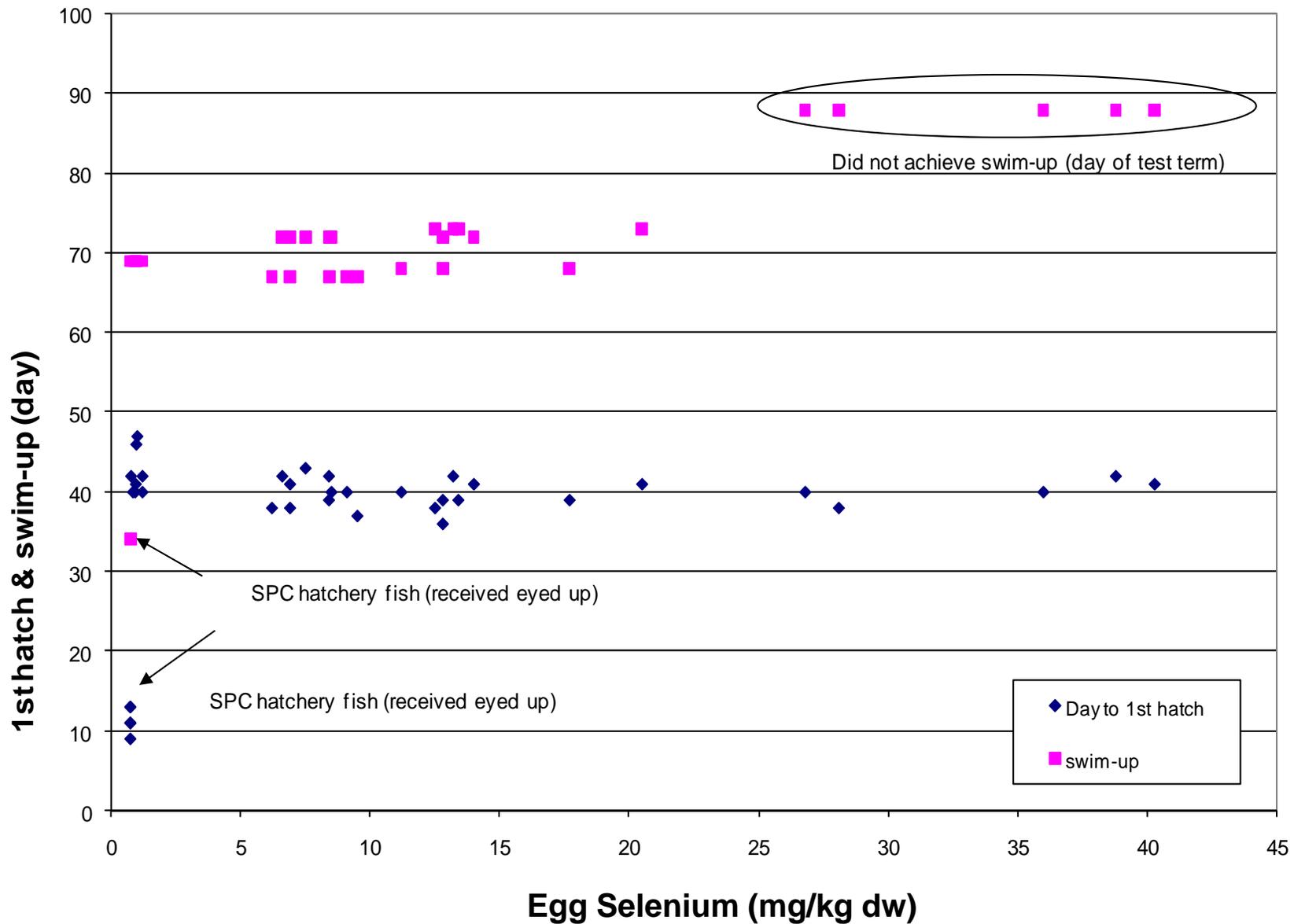
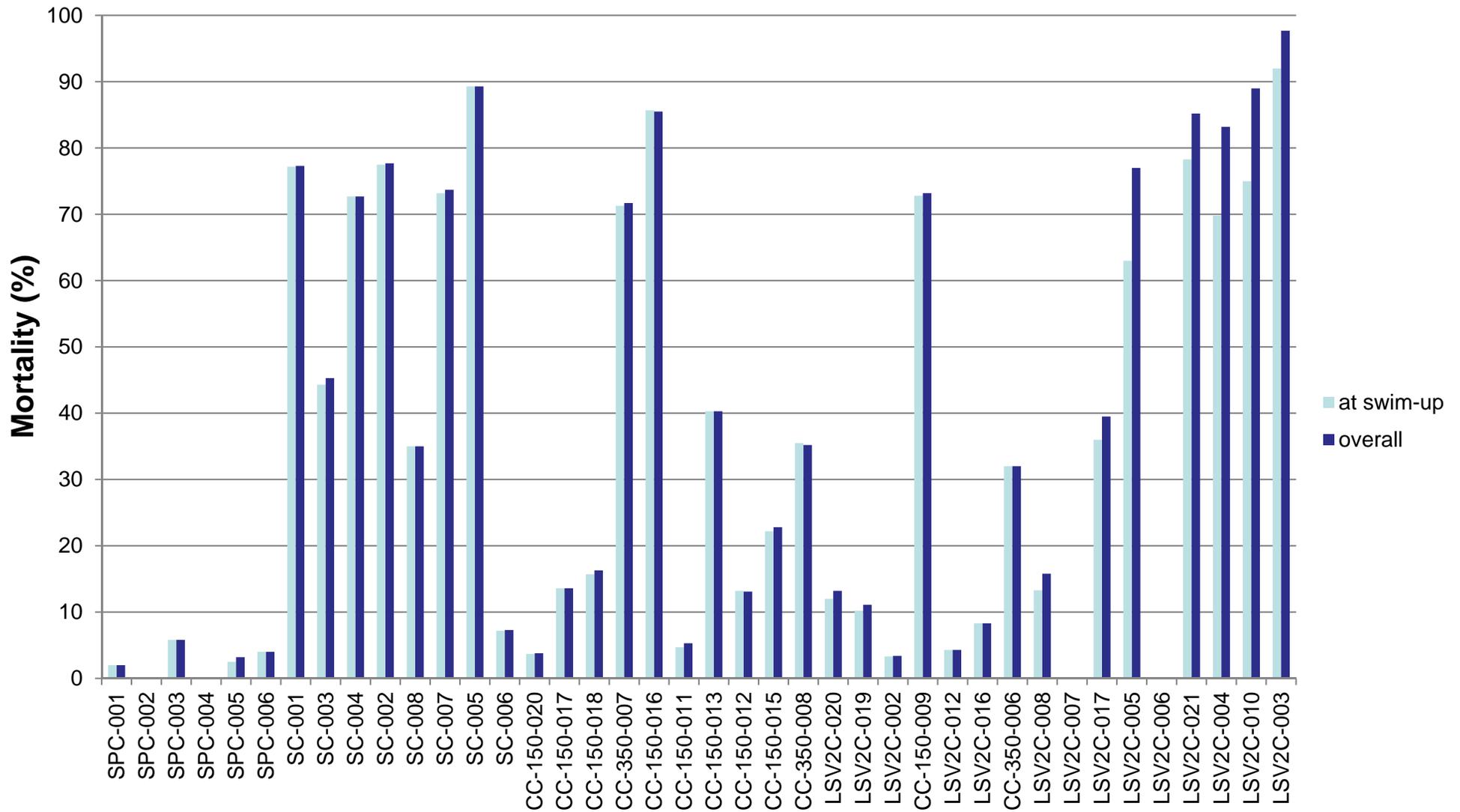


Figure 13
Egg Selenium Concentration Versus
Day to First Hatch and Day to Swim-Up

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC





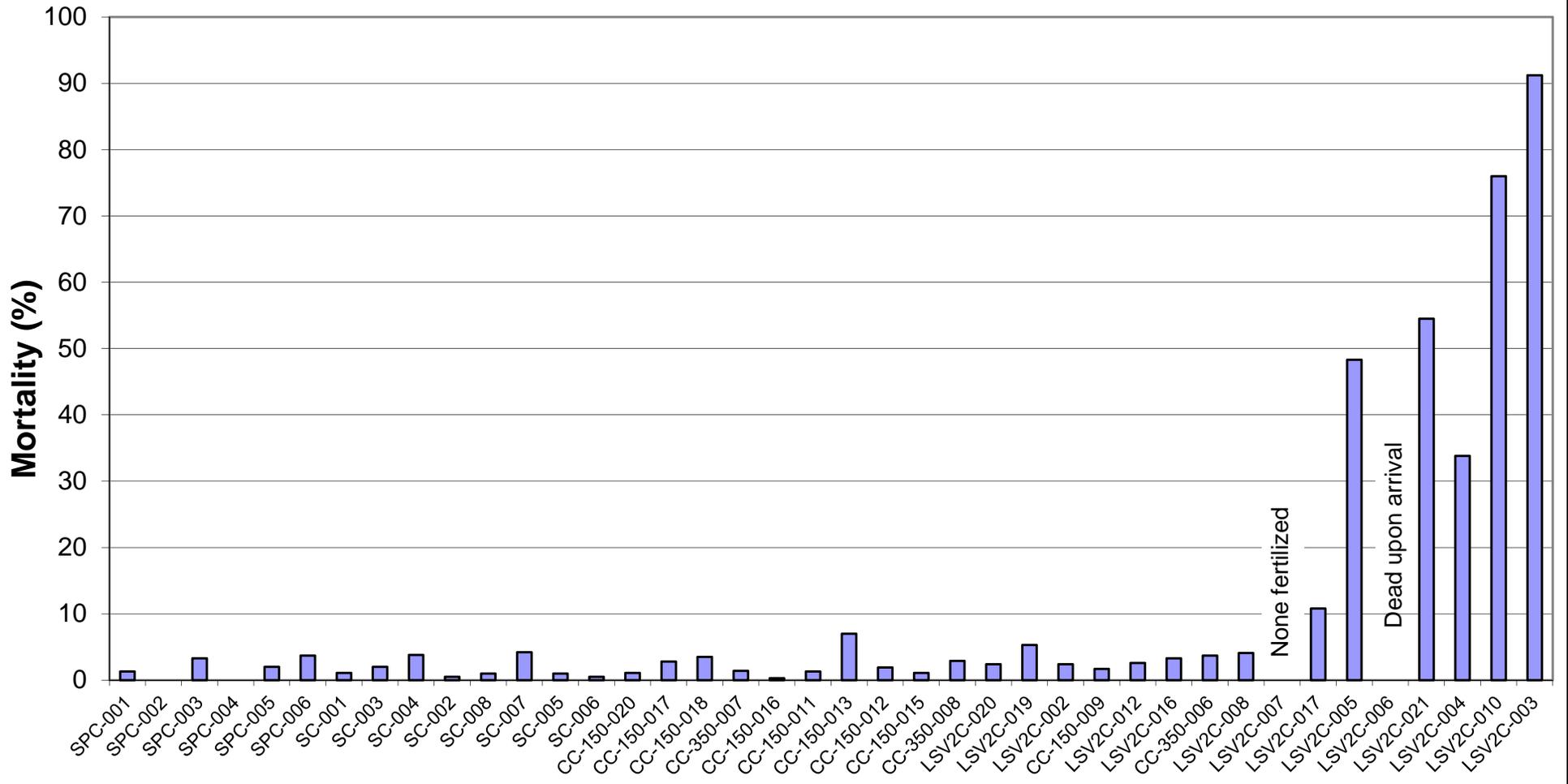
Note: The order of locations, as represented on the X axis, is in order of egg selenium concentration.

Figure 14
Fry Mortality at Swim-Up and Overall Mortality by Location

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC





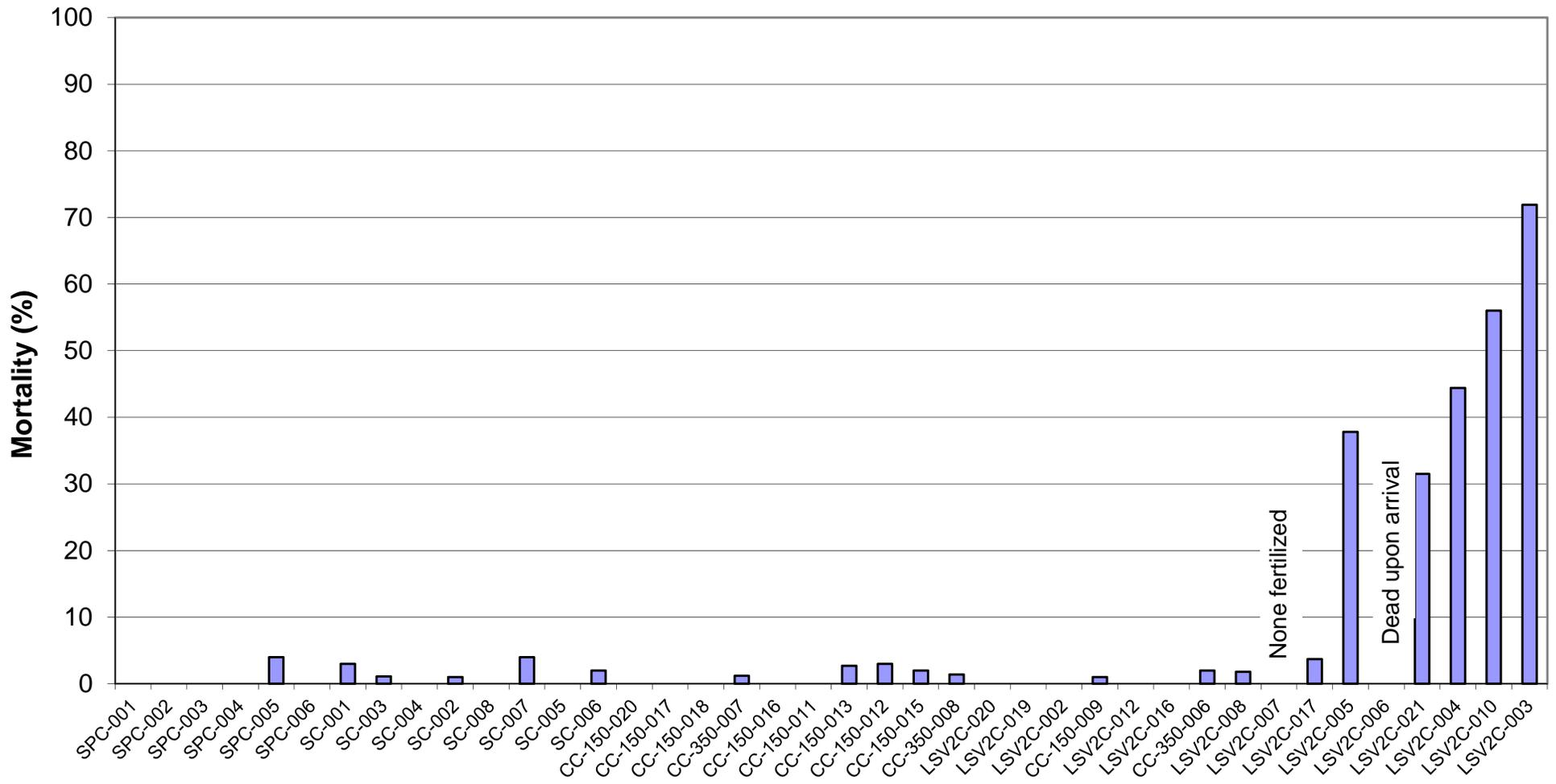
Note: The order of locations, as represented on the X axis, is in order of egg selenium concentration.

Figure 15
Percent Mortality from Hatch to Test Termination by Location

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC





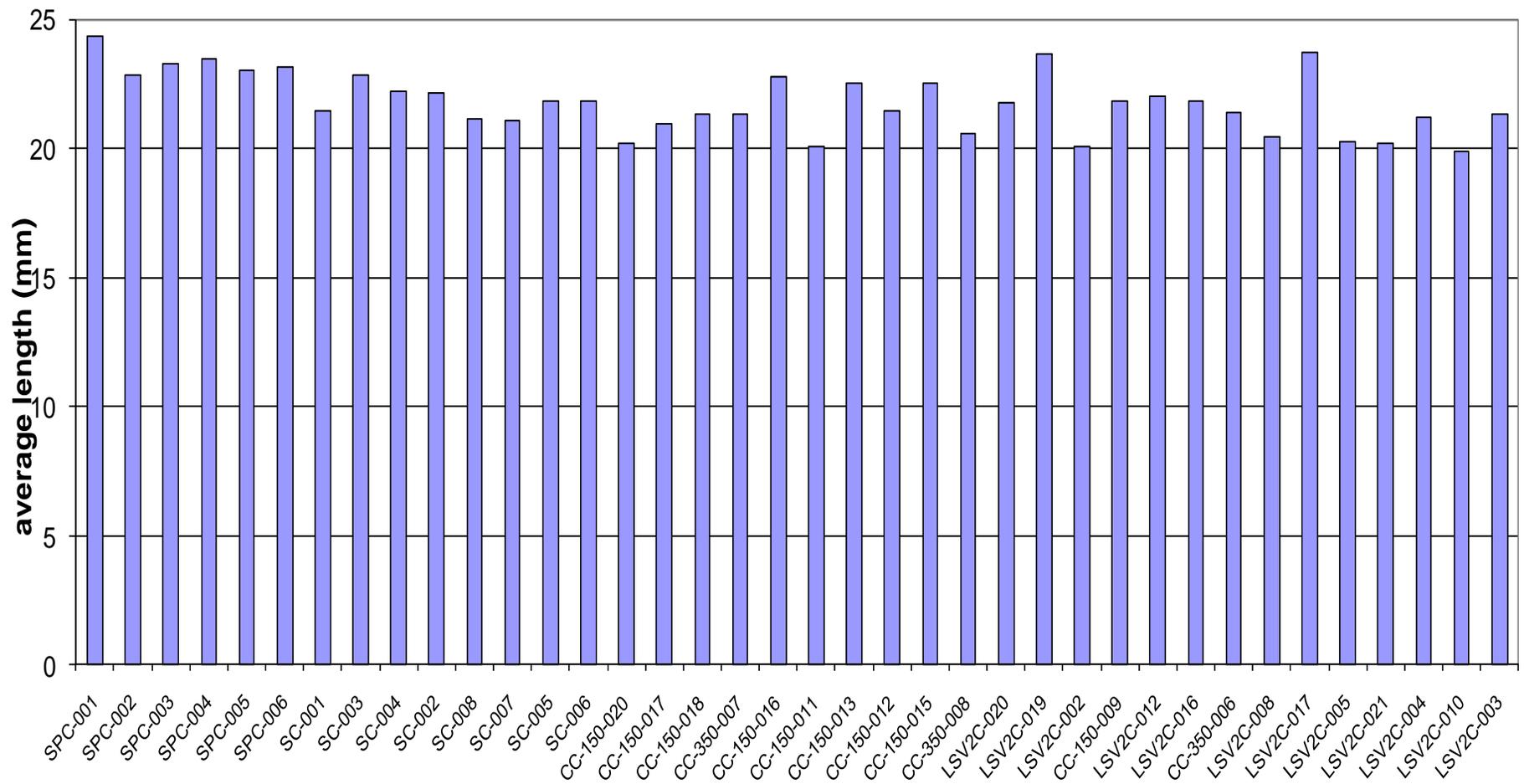
Note: The order of locations, as represented on the X axis, is in order of egg selenium concentration.

Figure 16
Percent Mortality 15-Day Post Swim-Up Feeding Trial

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC





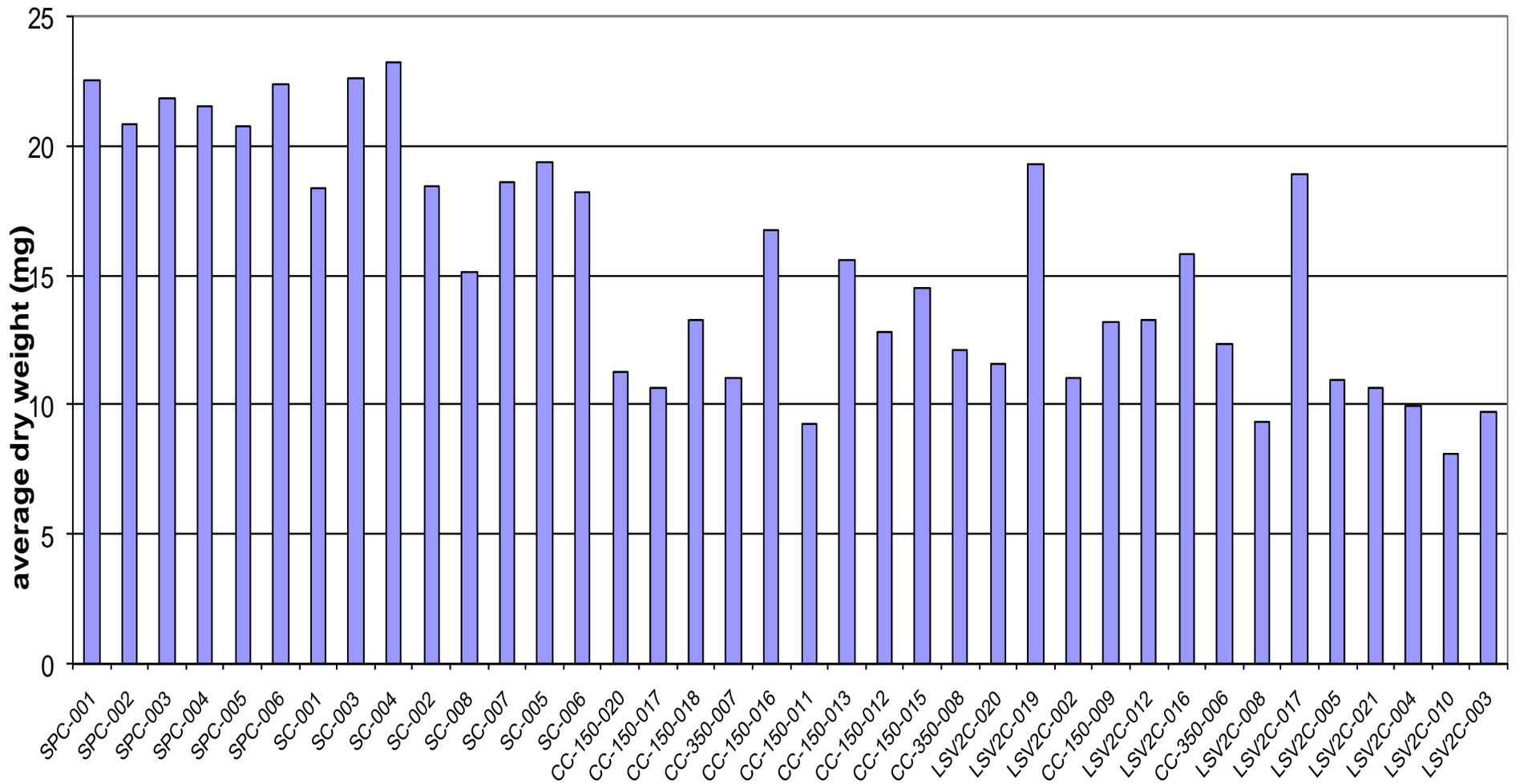
Note: The order of locations, as represented on the X axis, is in order of egg selenium concentration.

Figure 17
Brown Trout Growth based on Larval Fish Length

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC





Note: The order of locations, as represented on the X axis, is in order of egg selenium concentration.

Figure 18
Brown Trout Growth based on Larval Fish Dry Weight

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC



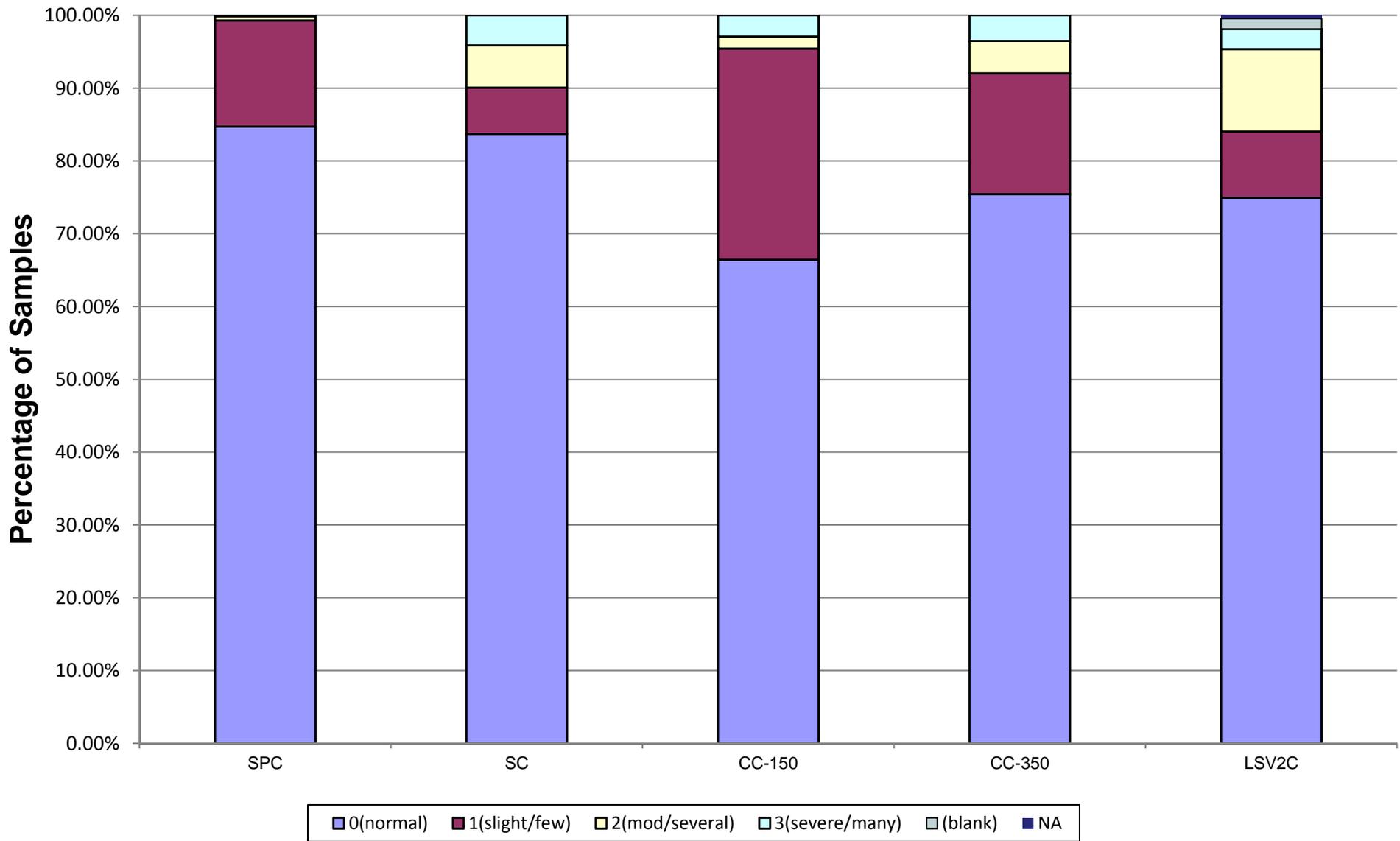


Figure 19
Percent Cranio-Facial Deformities for Larval Brown Trout

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC



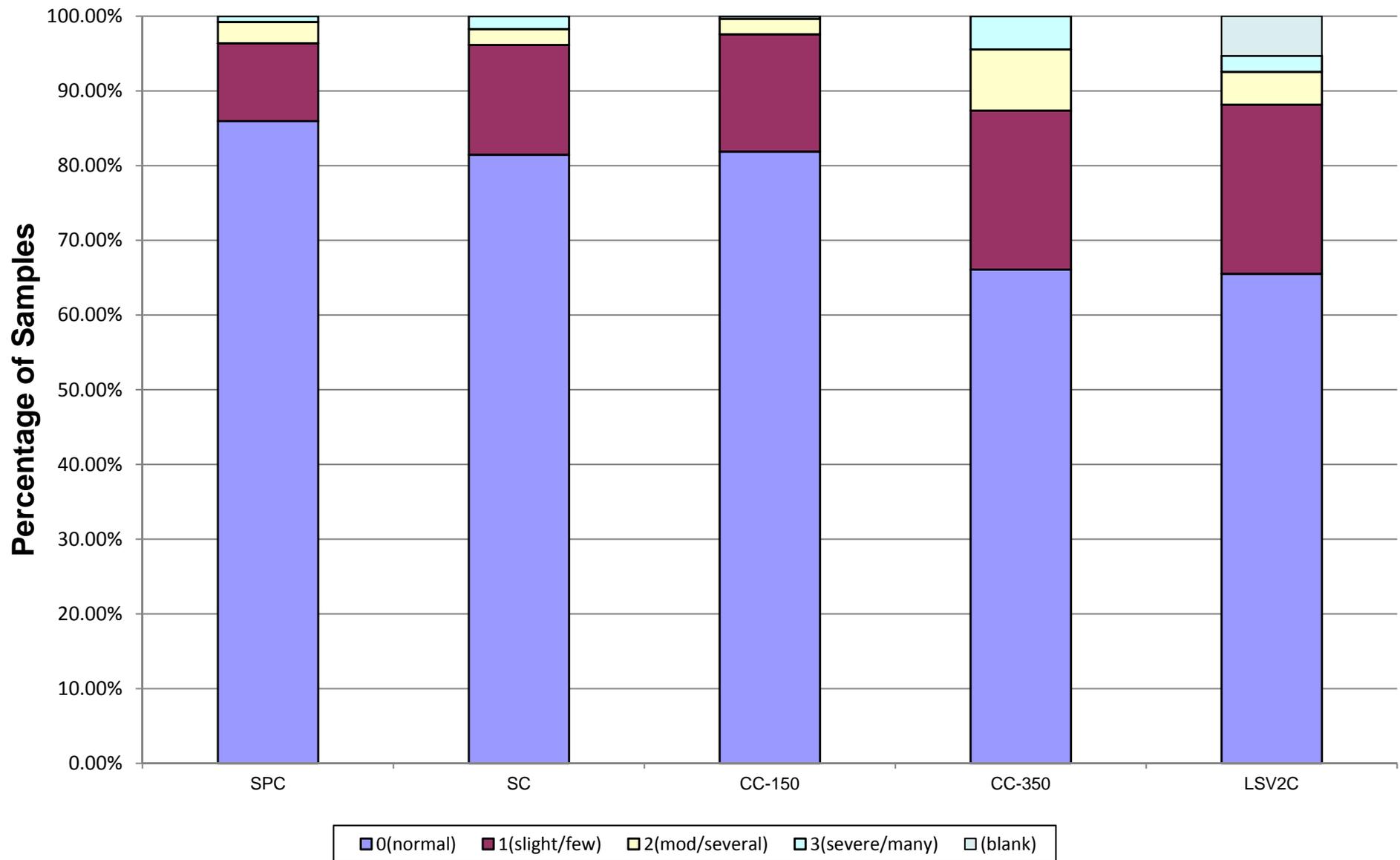


Figure 20
Percent Skeletal Deformities for Larval Brown Trout

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC



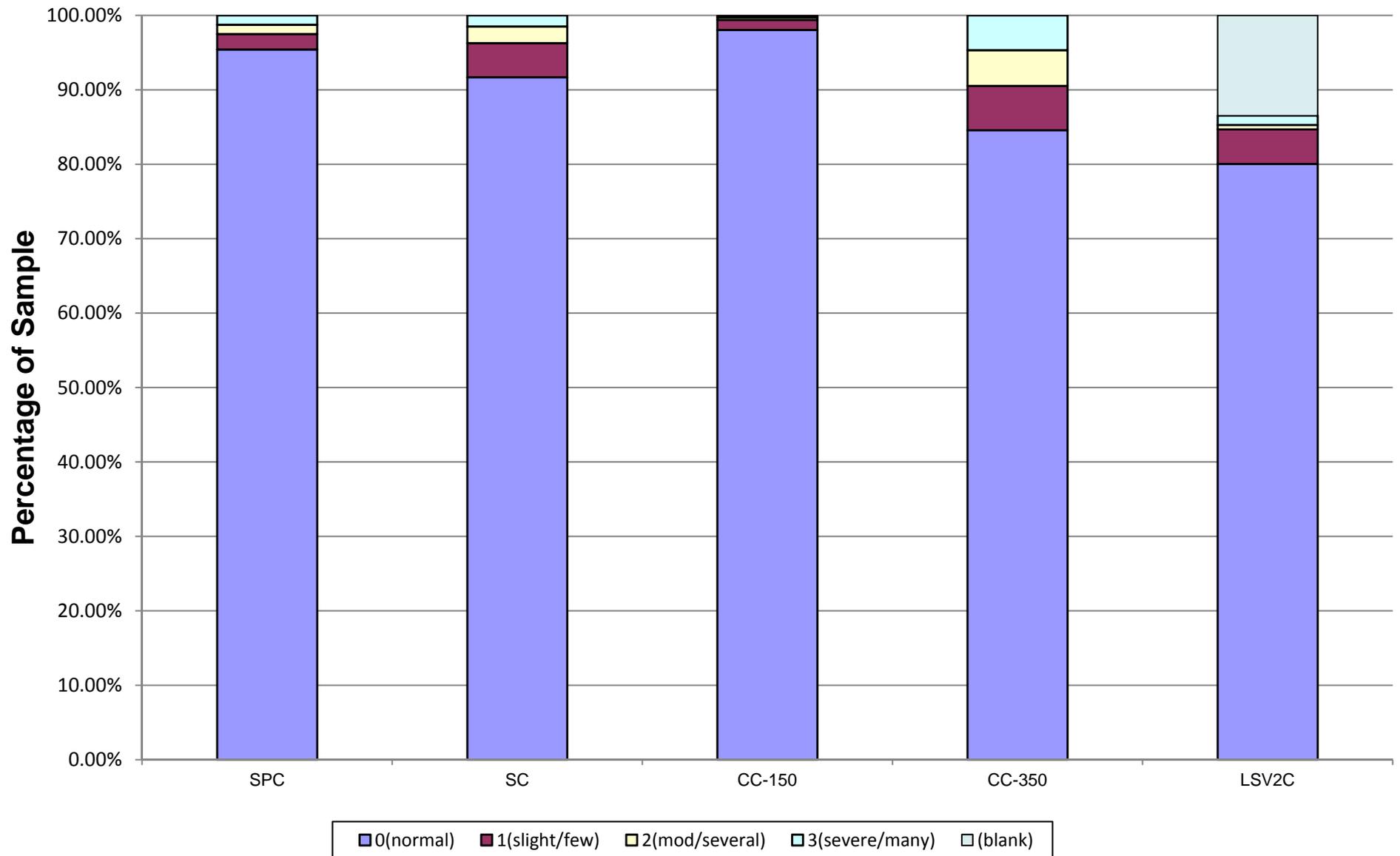


Figure 21
Percent Fin of Finfold Deformities for Larval Brown Trout

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC



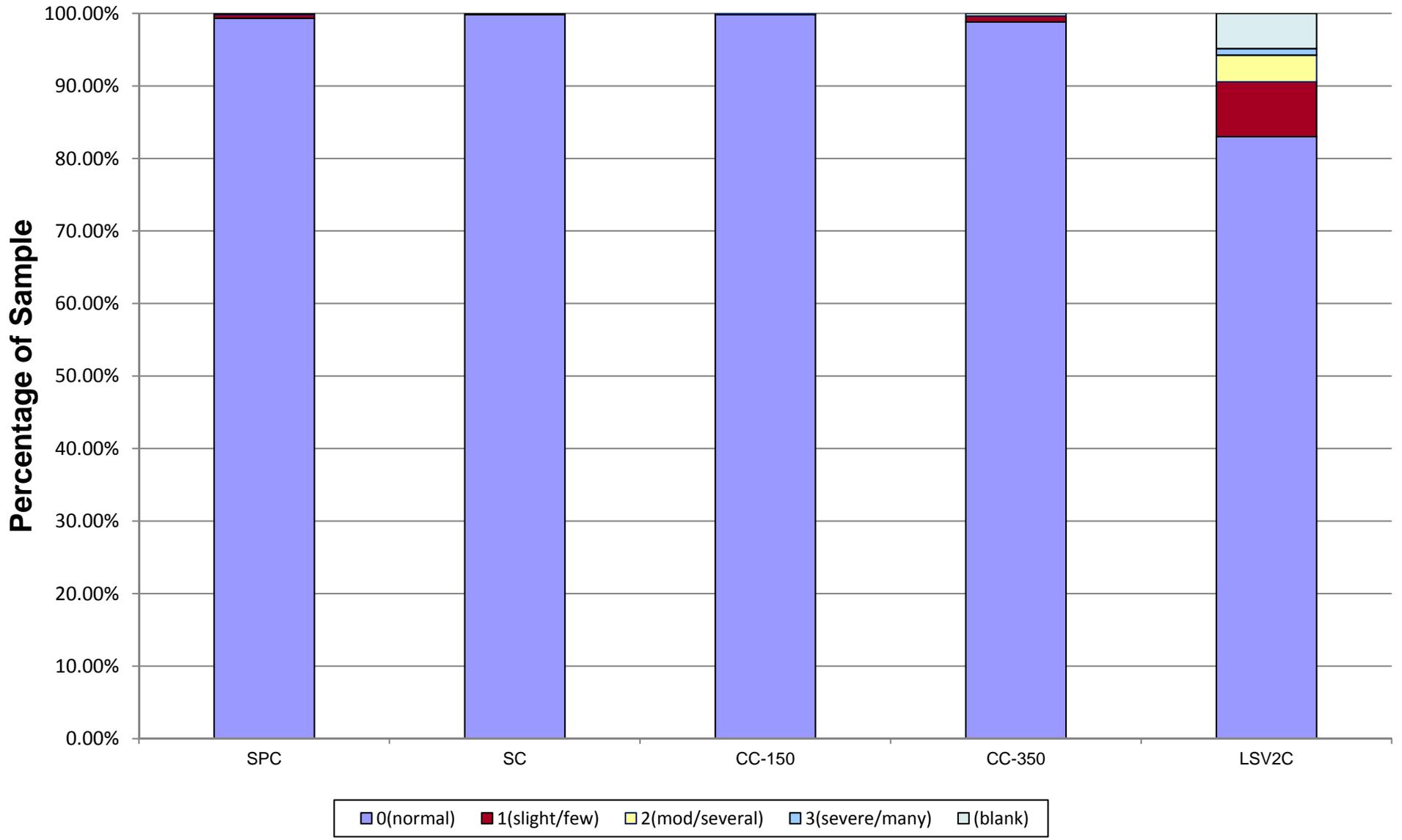
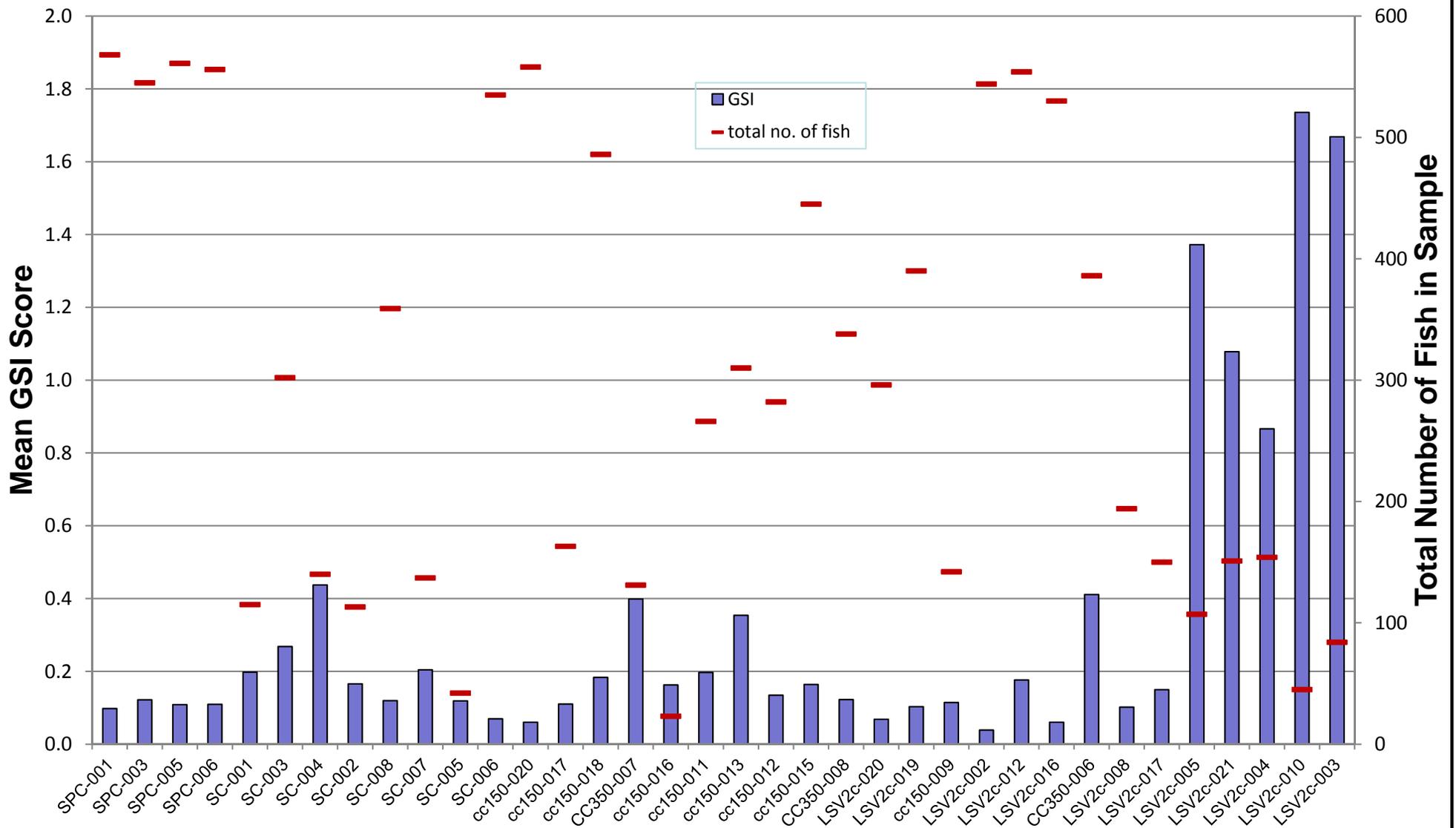


Figure 22
Percent Edematous Tissue Frequency for Larval Brown Trout

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011
REV: 1	BY: SMC CHK: SMC

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Note: The order of locations, as represented on the X axis, is in order of egg selenium concentration.

Figure 23
Mean GSI Score and Total Number of Fish Evaluated by Location and Sample ID
Ranked by Egg selenium Concentration

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011
REV: 1	BY: SMC CHK: SMC



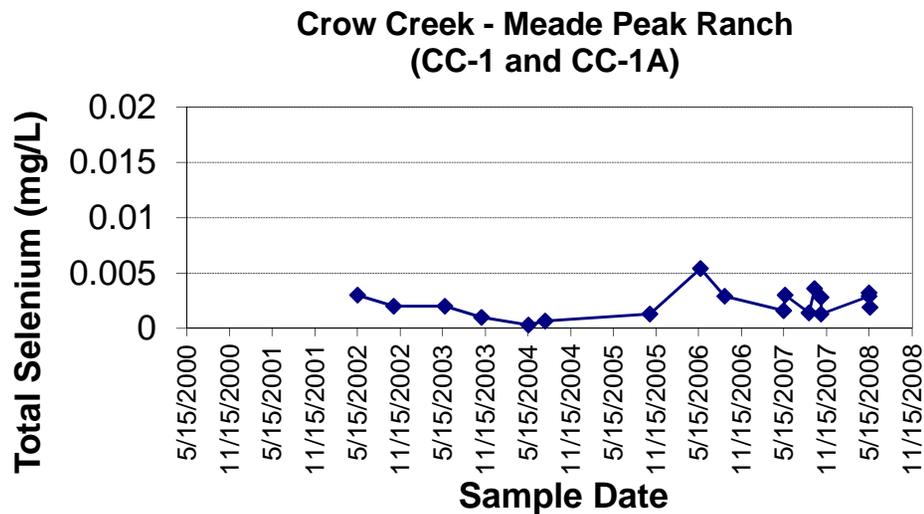
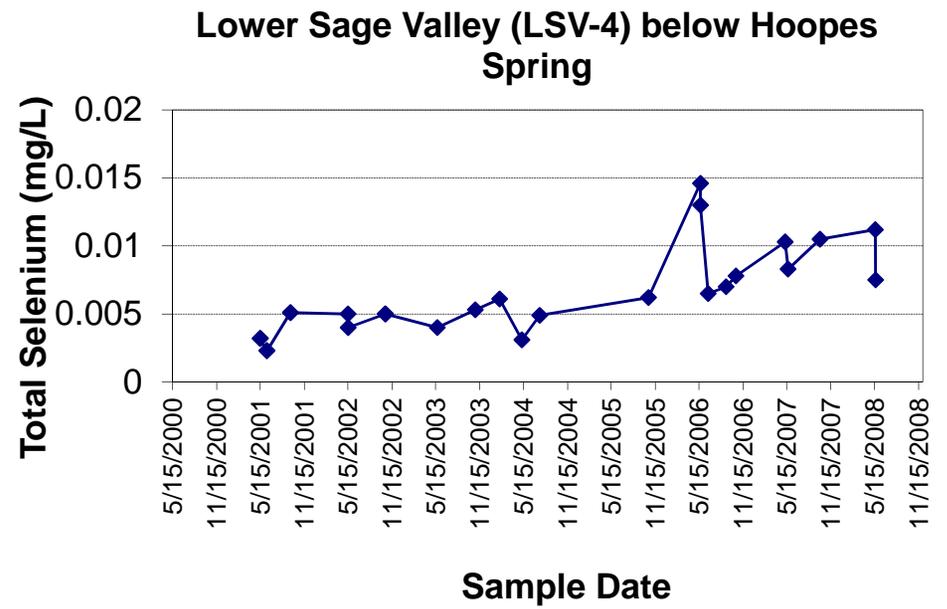
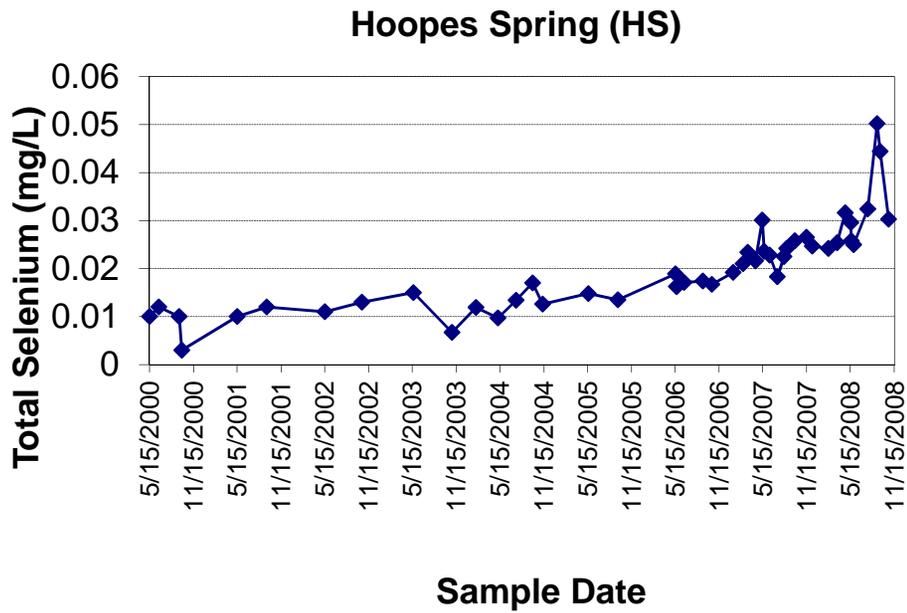


Figure 24
Surface Water Selenium Concentrations

J.R. Simplot Company
Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011
REV: 1	BY: SMC CHK: SMC



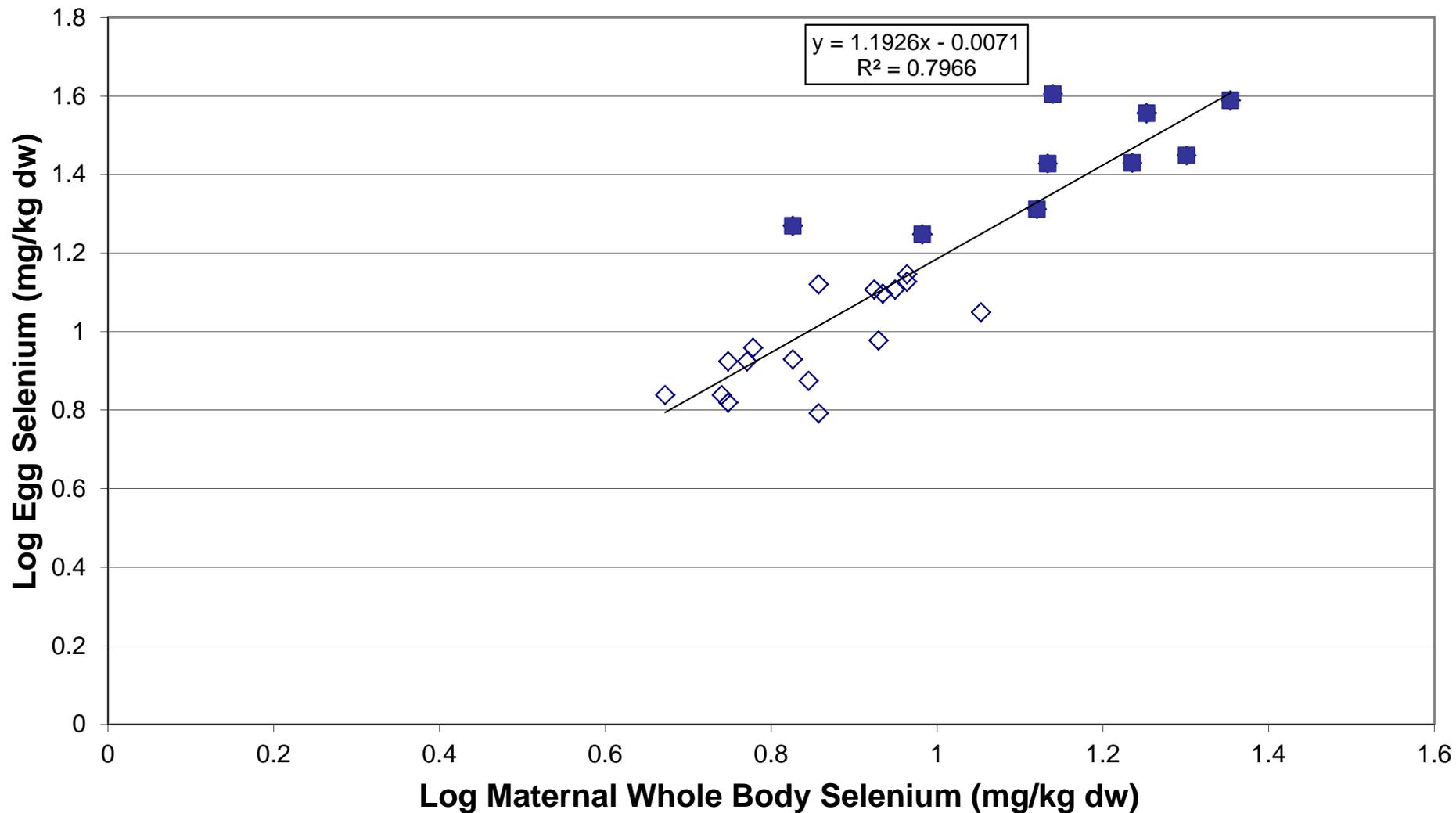


Figure 25
Log Maternal Whole Body Selenium Concentration (Wild Fish) Versus
Log Egg Selenium Concentrations

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011
REV: 1	BY: SMC CHK: SMC



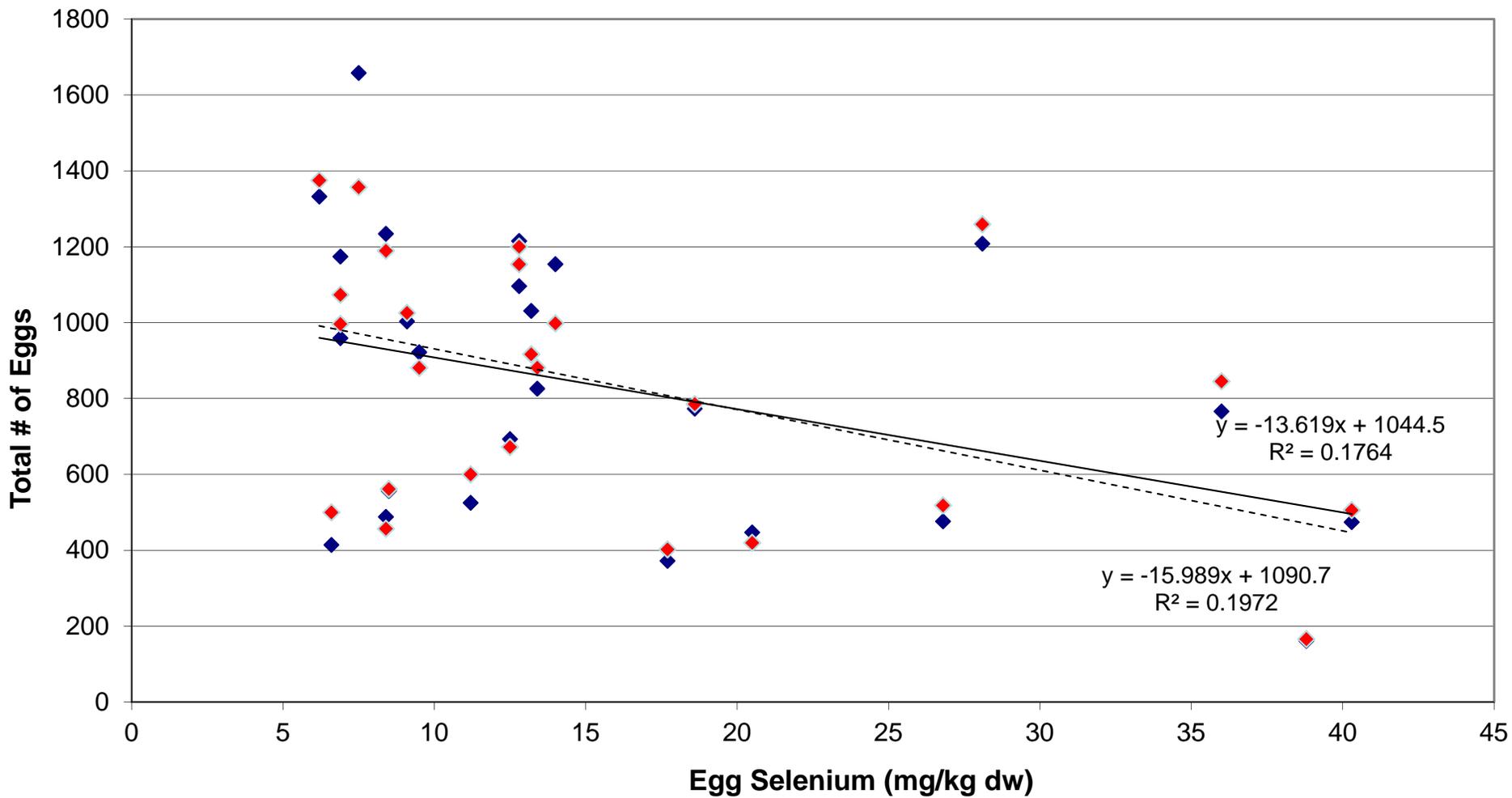


Figure 26
Egg Selenium Concentration Versus
Total Egg Abundance from Wild Female Fish

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011
REV: 1	BY: SMC CHK: SMC



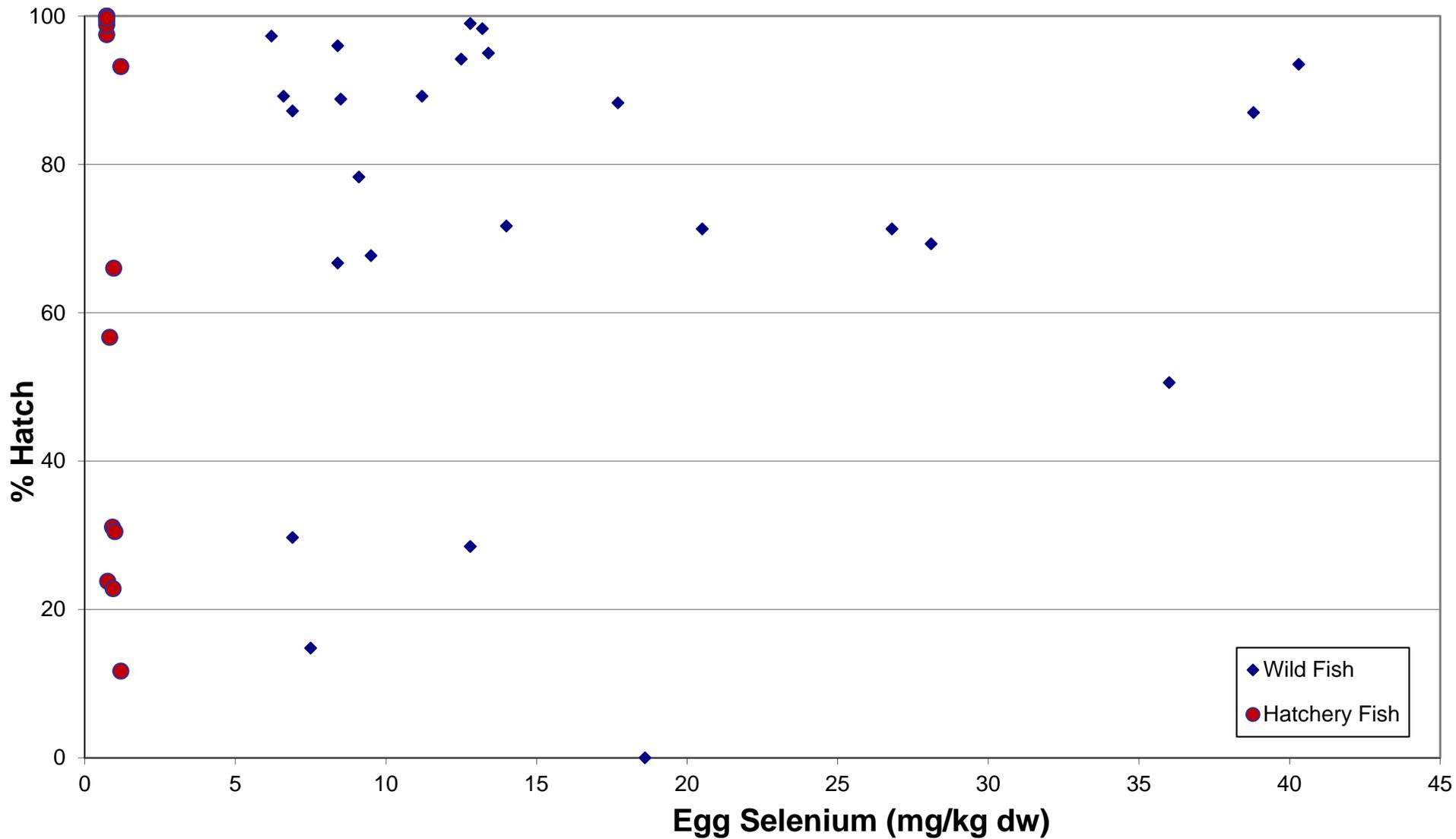


Figure 27
Egg Selenium Concentration Versus
Percent of Brown Trout Fry that Hatched

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC

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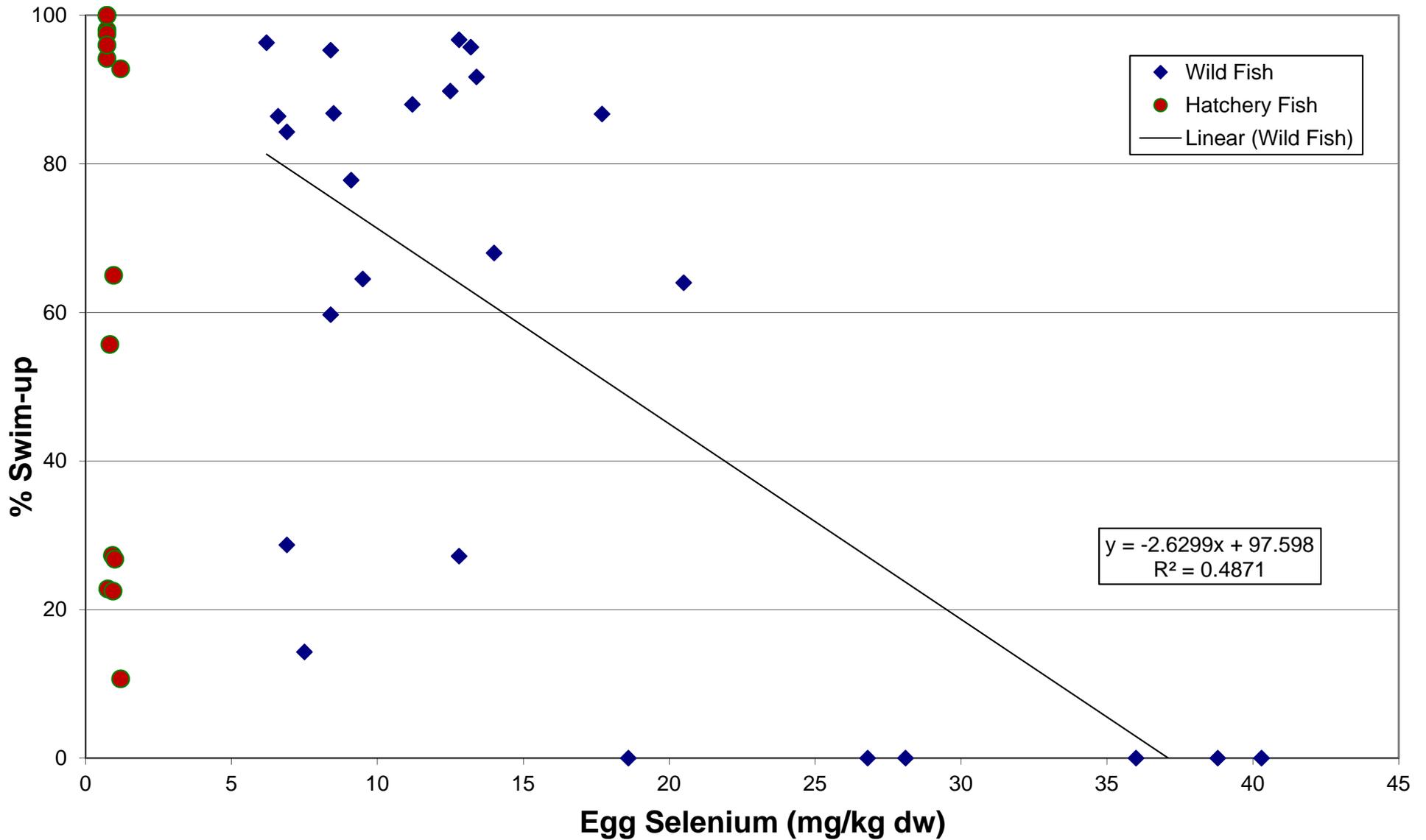


Figure 28
Wild Fish Egg Selenium Concentration Versus
Percent of Trout Achieving Swim-Up

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC

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Egg Selenium Versus Weight

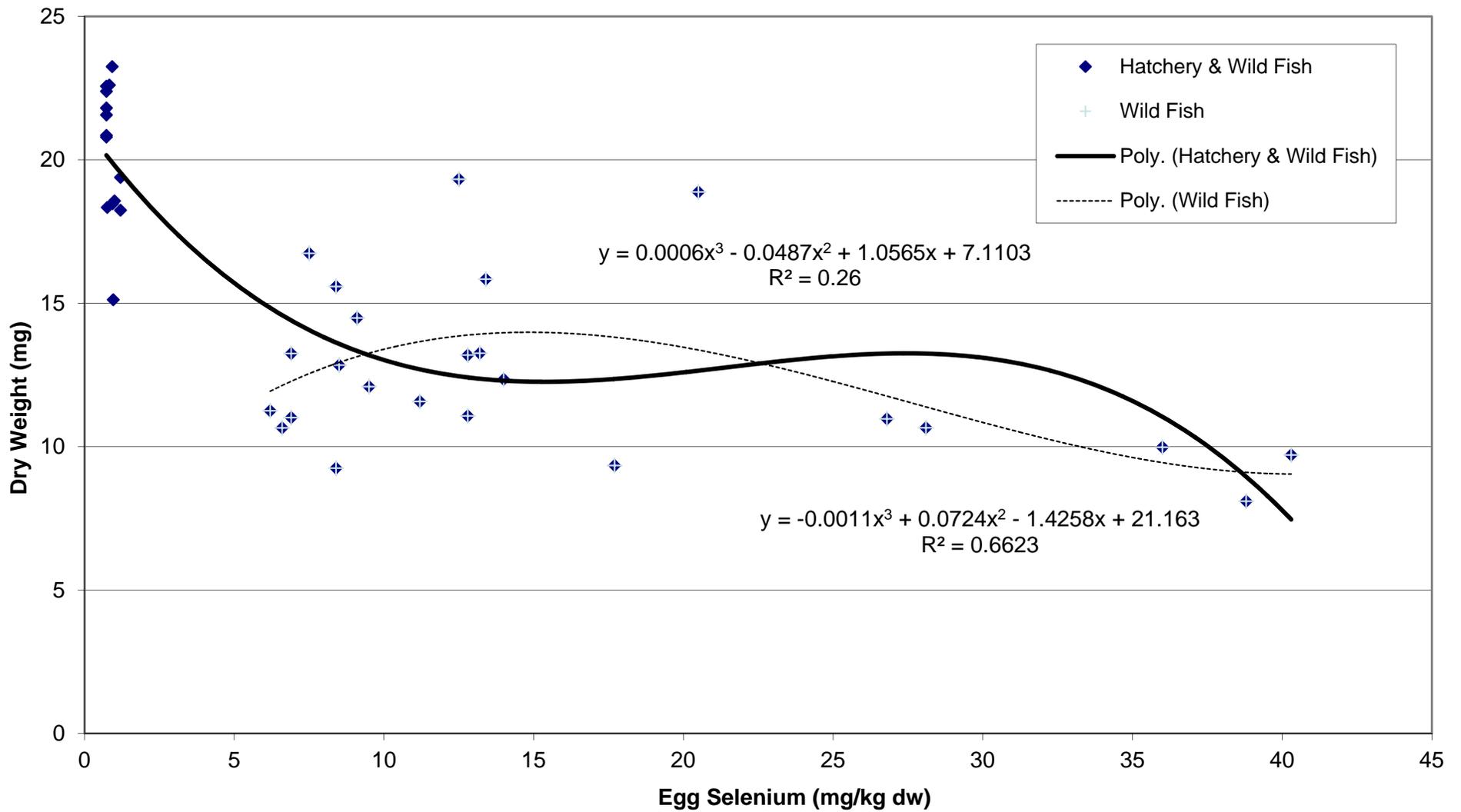


Figure 29
Egg Selenium Concentration Versus
Larval Fish Weight

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011
REV: 1	BY: SMC CHK: SMC



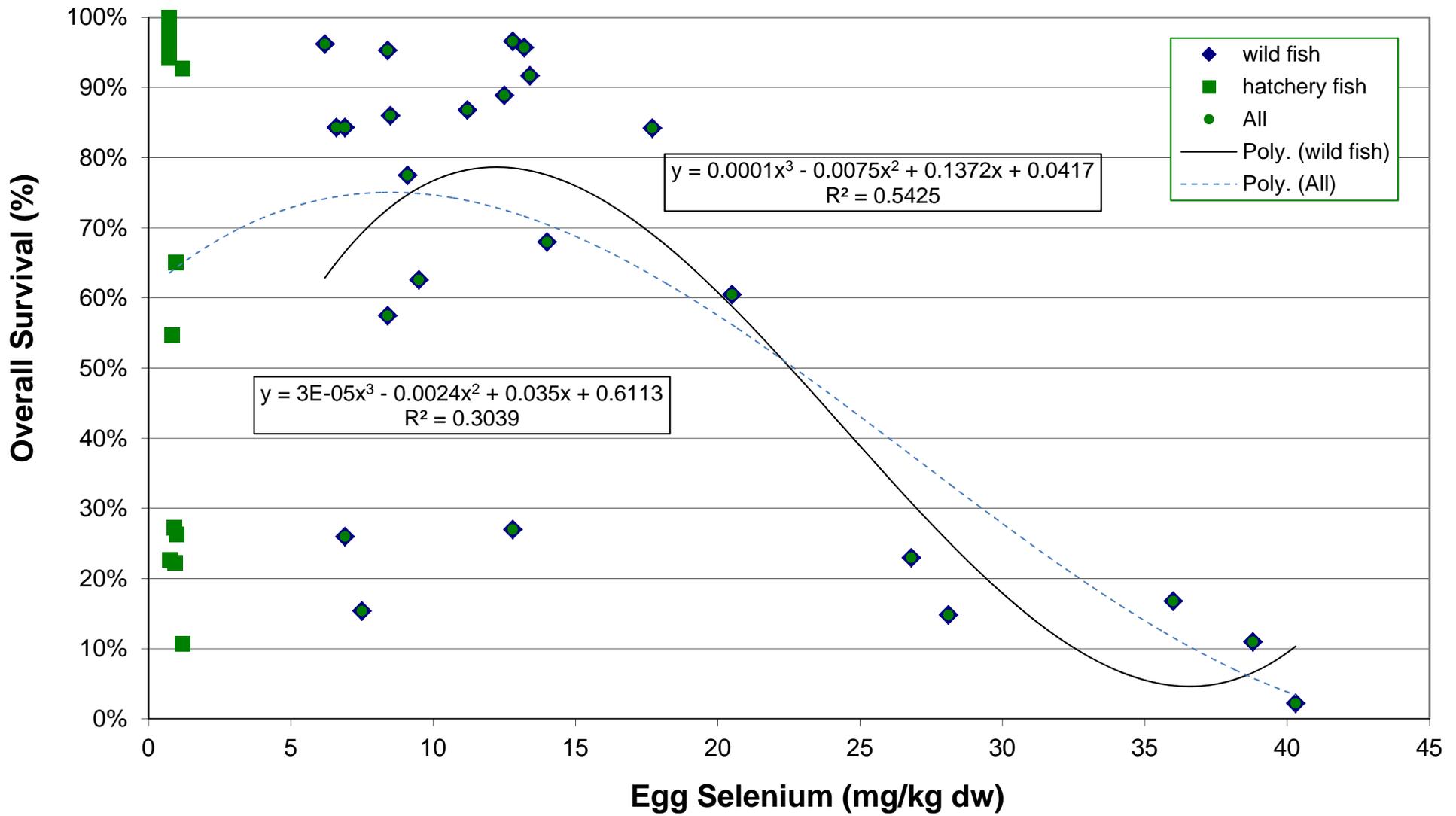


Figure 30
Egg Selenium Concentration Versus
Overall Percent Survival

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC

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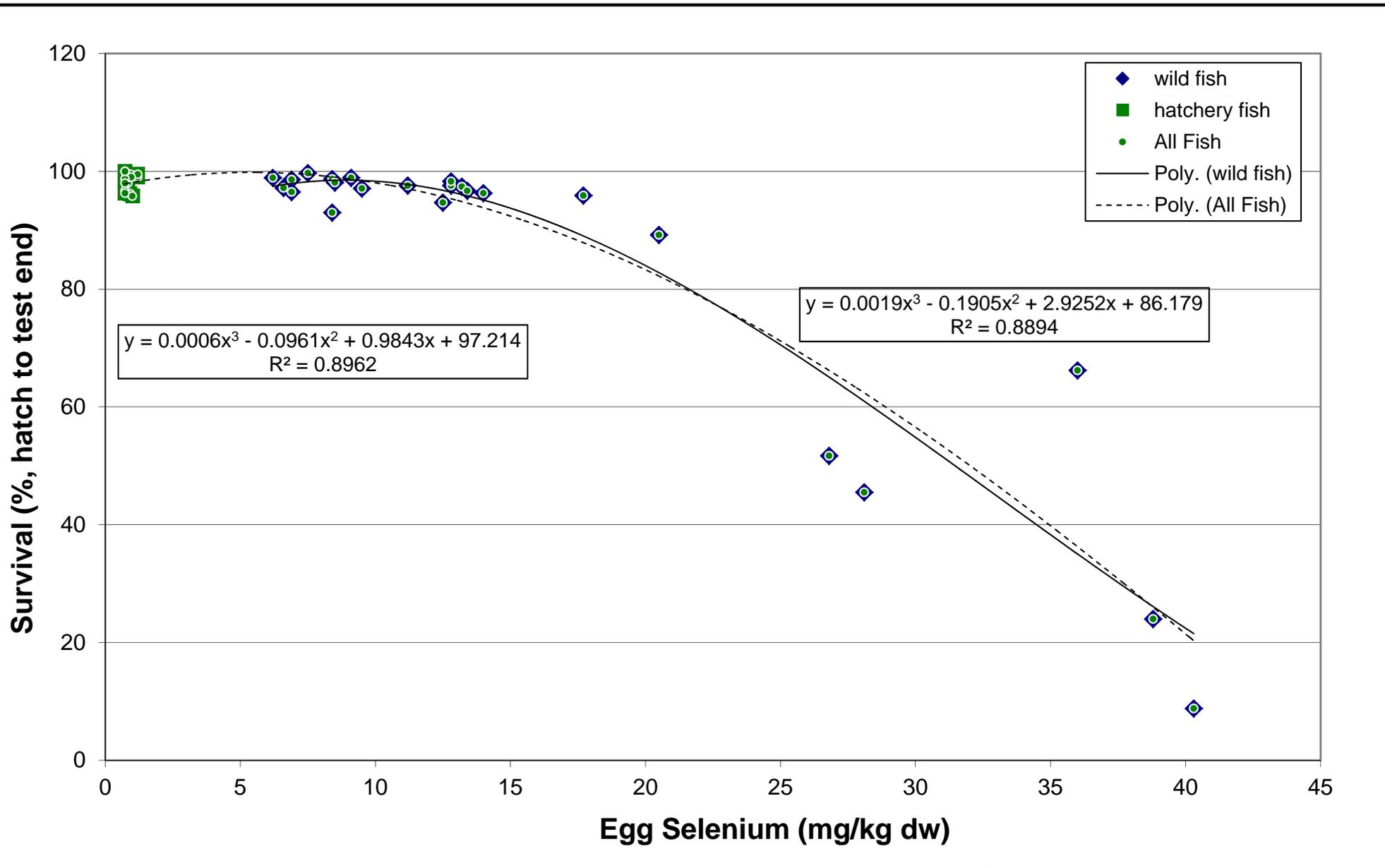


Figure 31
Egg Selenium Concentration Versus
Percent Survival (Hatch to Test End)

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC

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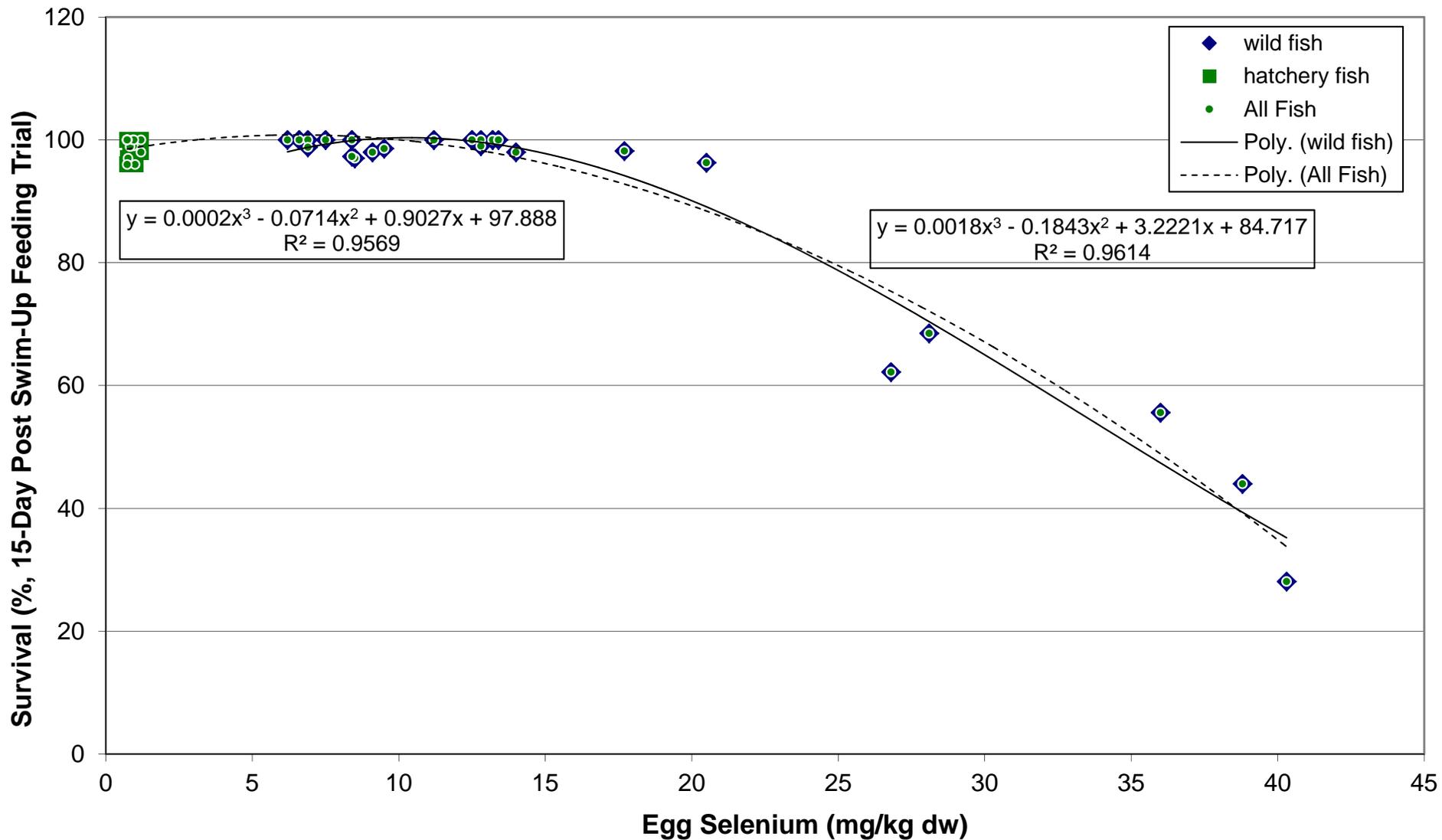


Figure 32
Egg Selenium Concentration Versus
Percent Survival in the 15-Day Post Swim-Up Feeding Trial

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC

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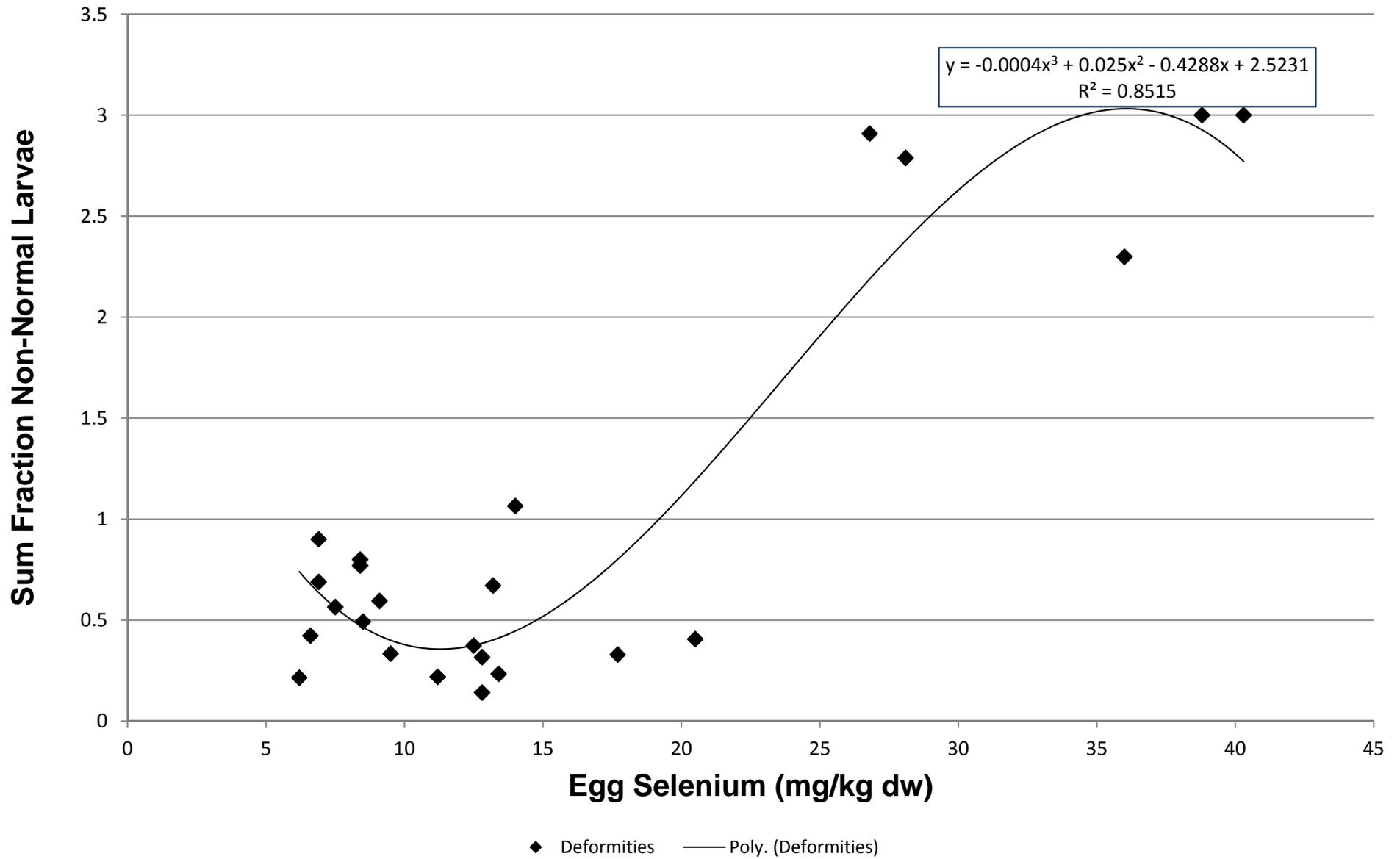


Figure 33
Egg Selenium Concentration Versus
Sum Fraction Non-Normal Larvae

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC



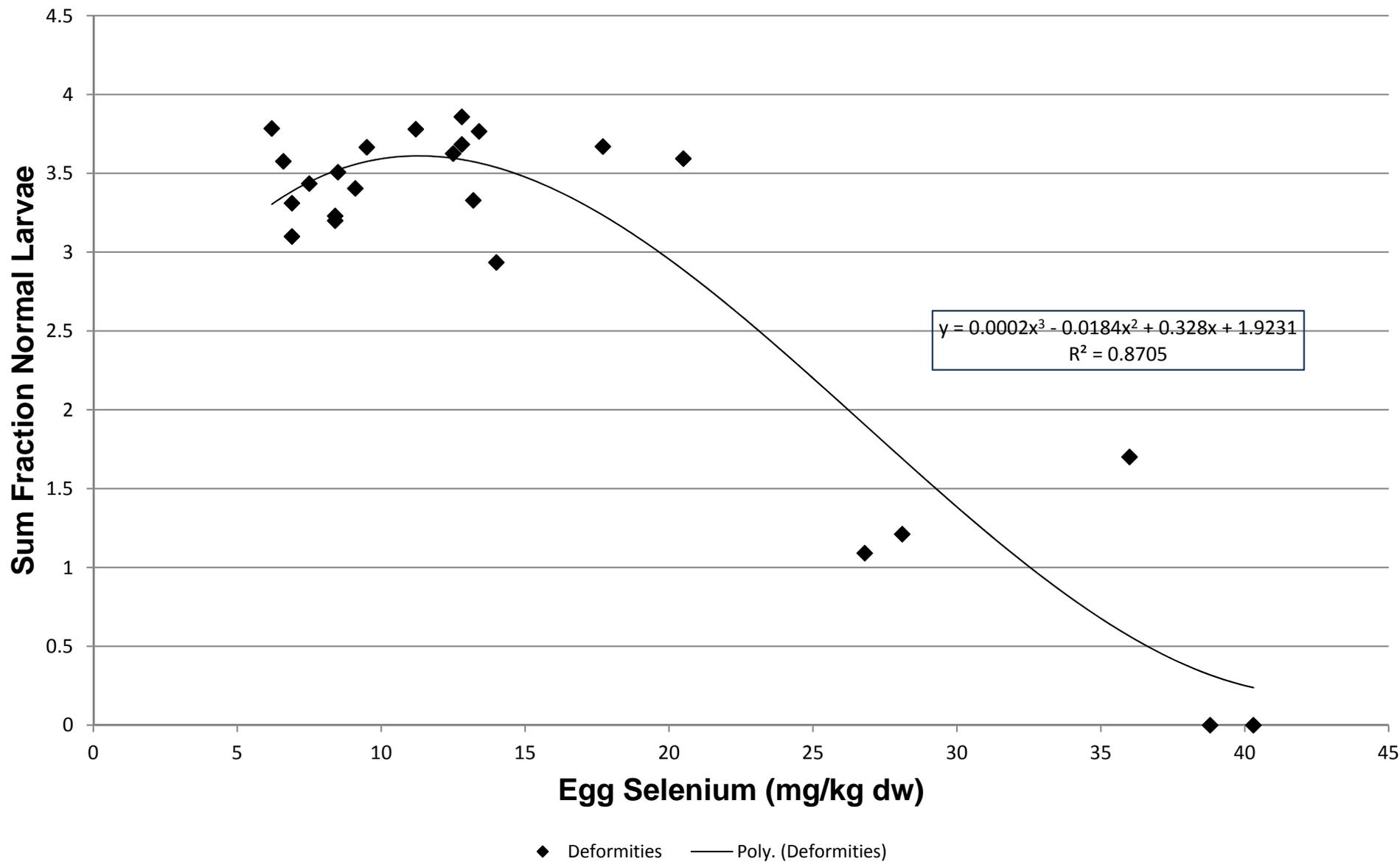


Figure 34
Egg Selenium Concentration Versus
Sum Fraction Normal Larvae

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011
REV: 1	BY: SMC CHK: SMC



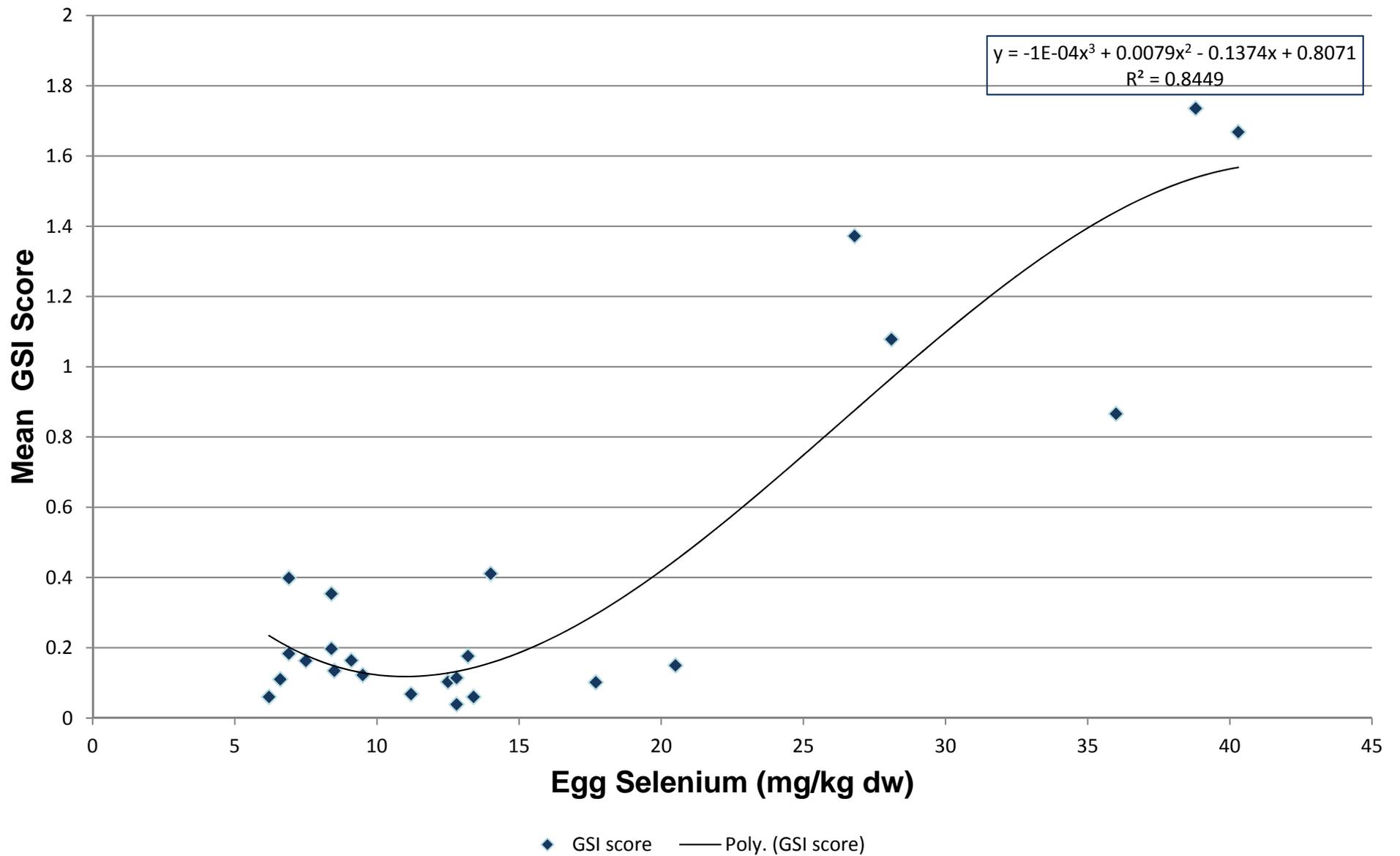


Figure 35
Egg Selenium Concentration Versus
Mean GSI Score

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC

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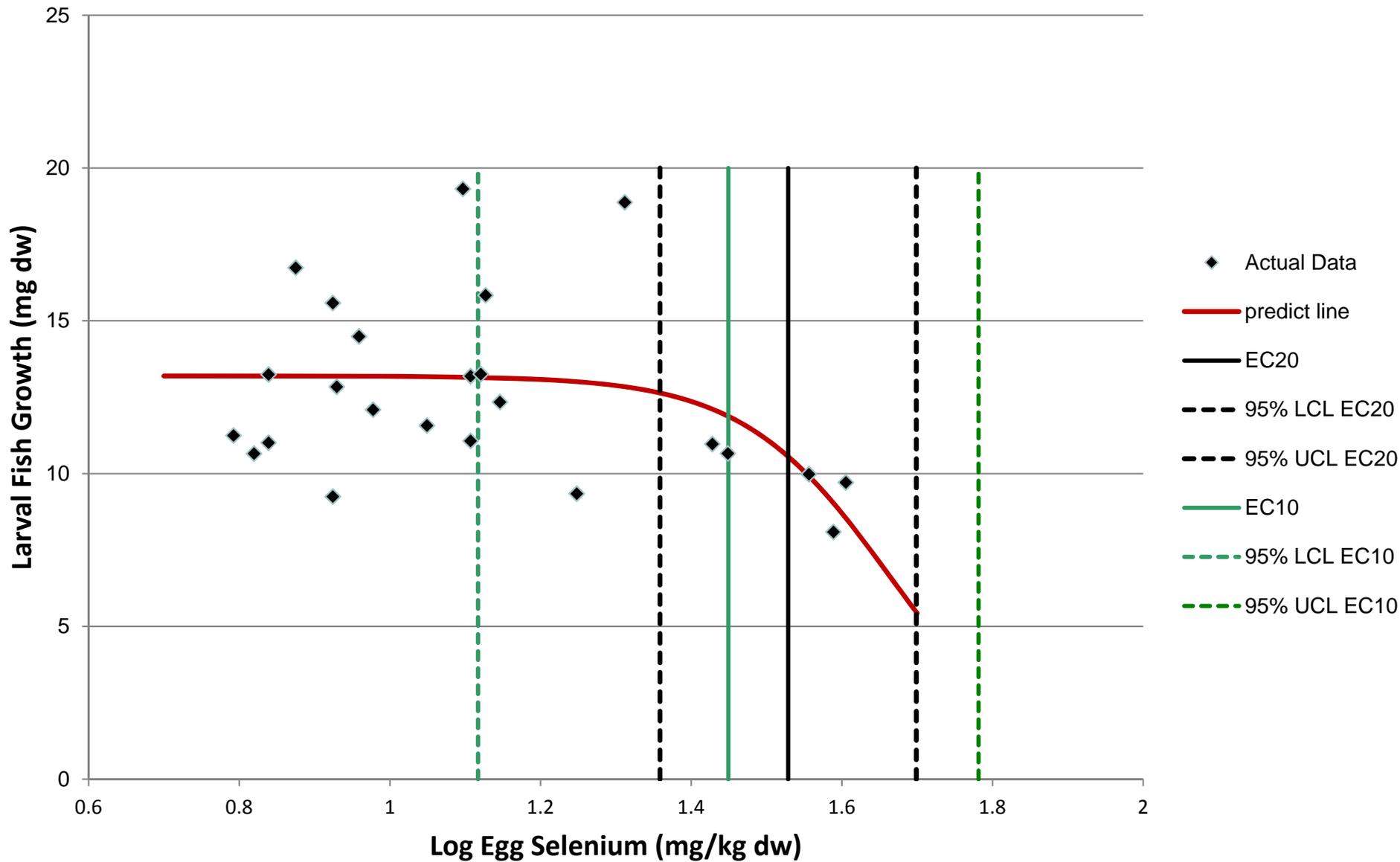


Figure 36
TRAP Model
Egg Selenium Concentration Versus Larval Fish Growth
15-Day Post Swim-Up Feeding Trial

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011
REV: 1	BY: SMC CHK: SMC



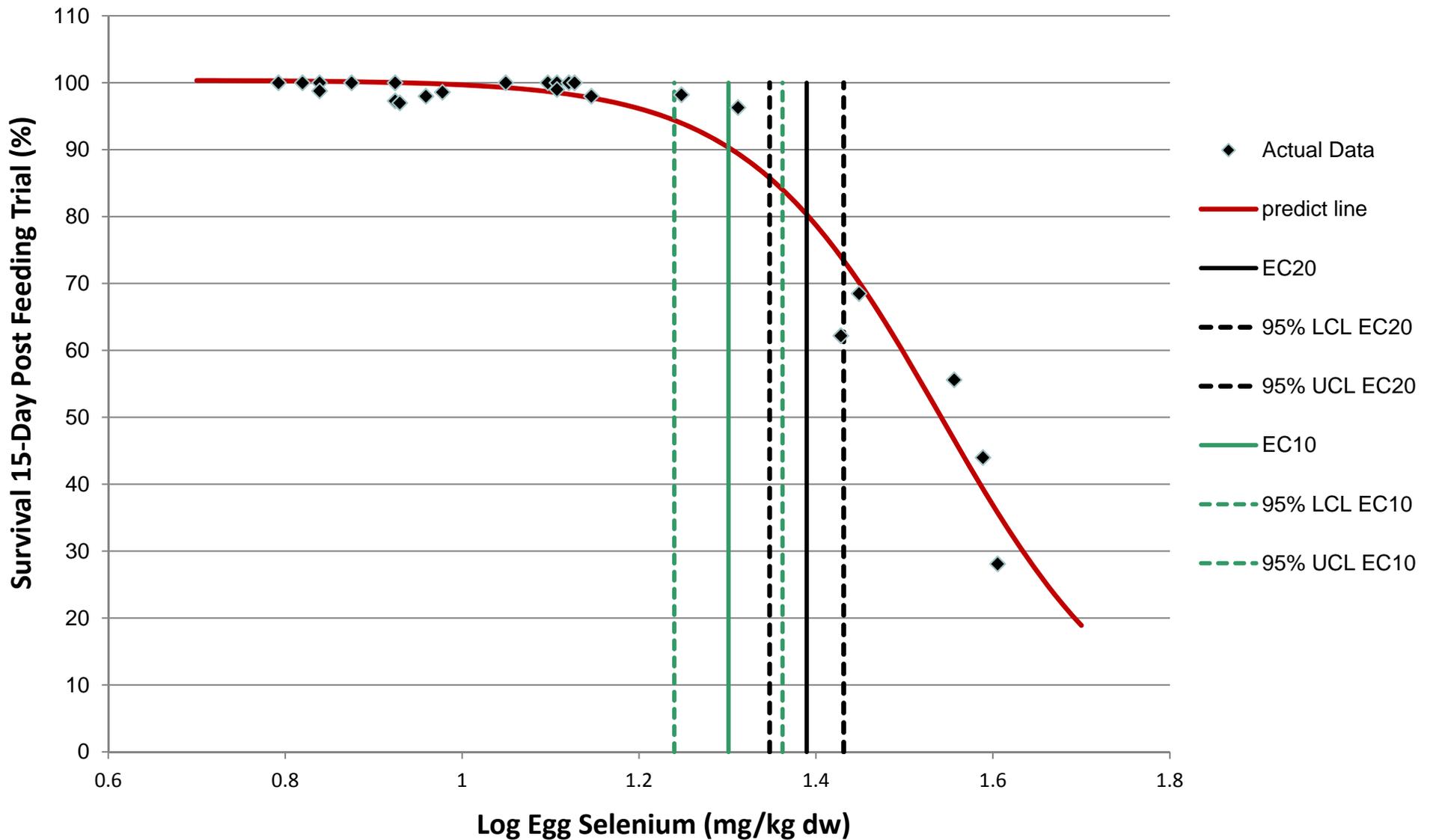


Figure 37
TRAP Model
Egg Selenium Concentration Versus Survival
15-Day Post Swim-Up Feeding Trial

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC

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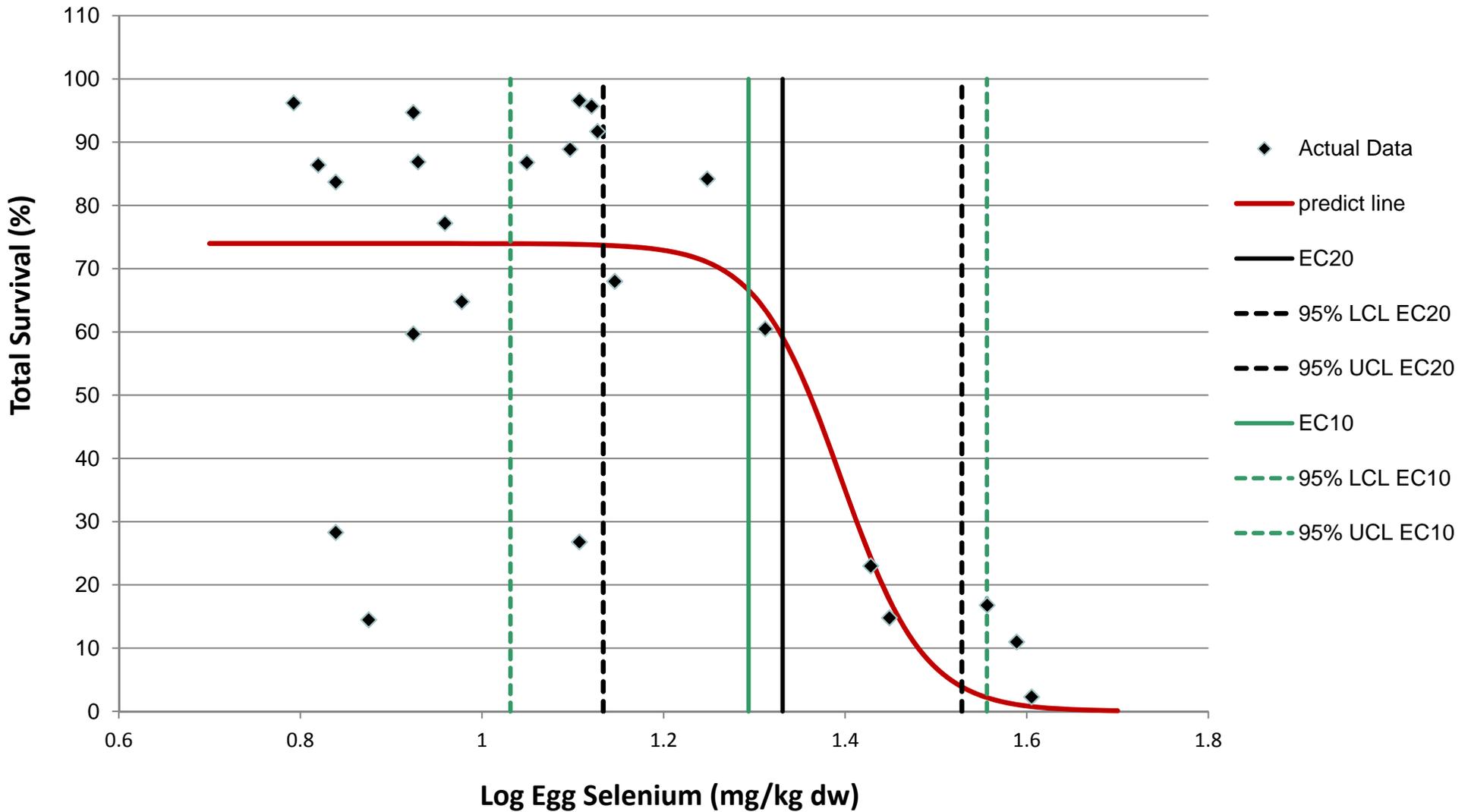


Figure 38
TRAP Model
Egg Selenium Concentration Versus Total Survival

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC

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Egg Selenium Versus % Survival (Hatch to Test End)

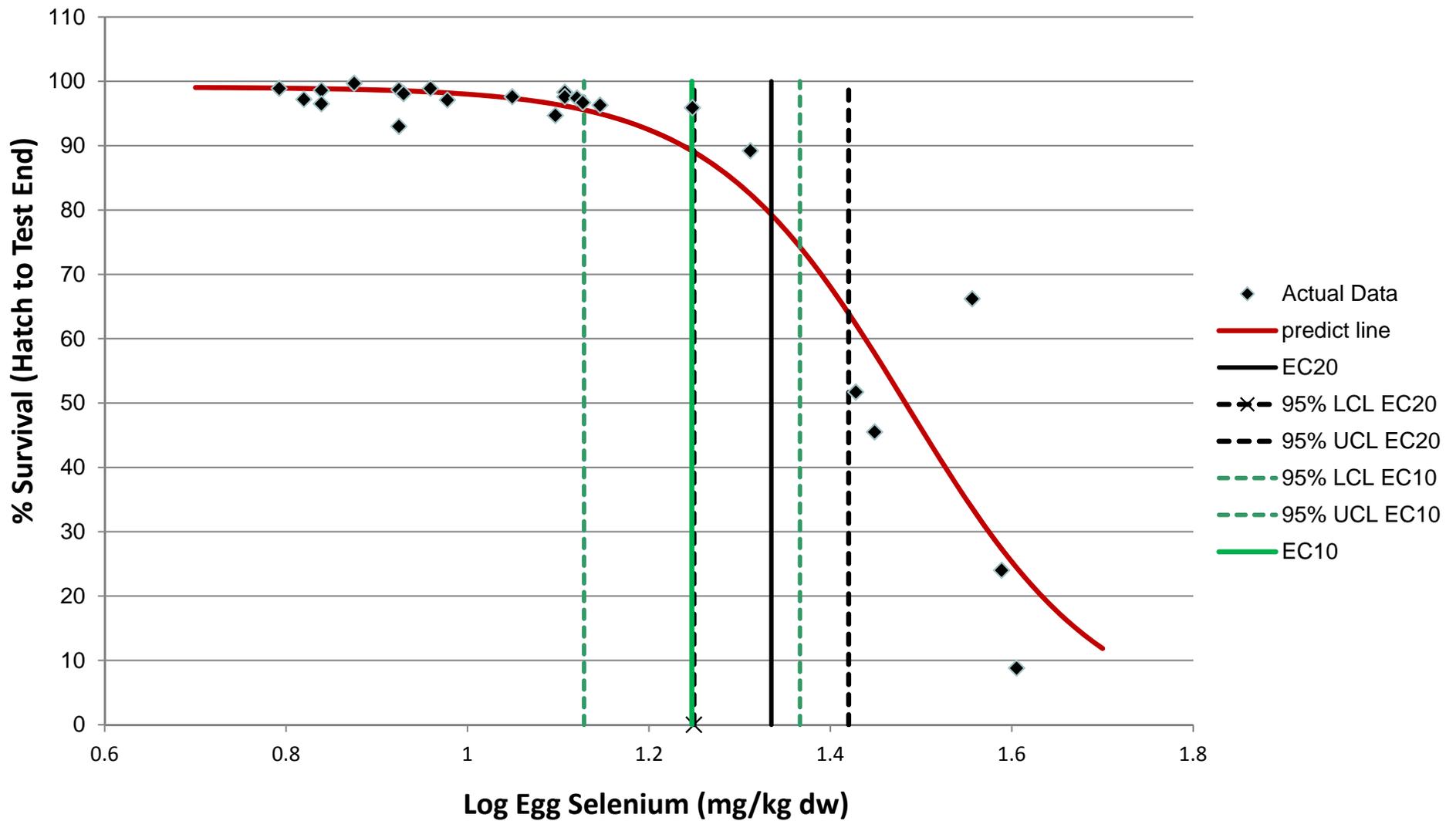


Figure 39
TRAP Model
Egg Selenium Concentration Versus Percent Survival
(Hatch to Test End)

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC



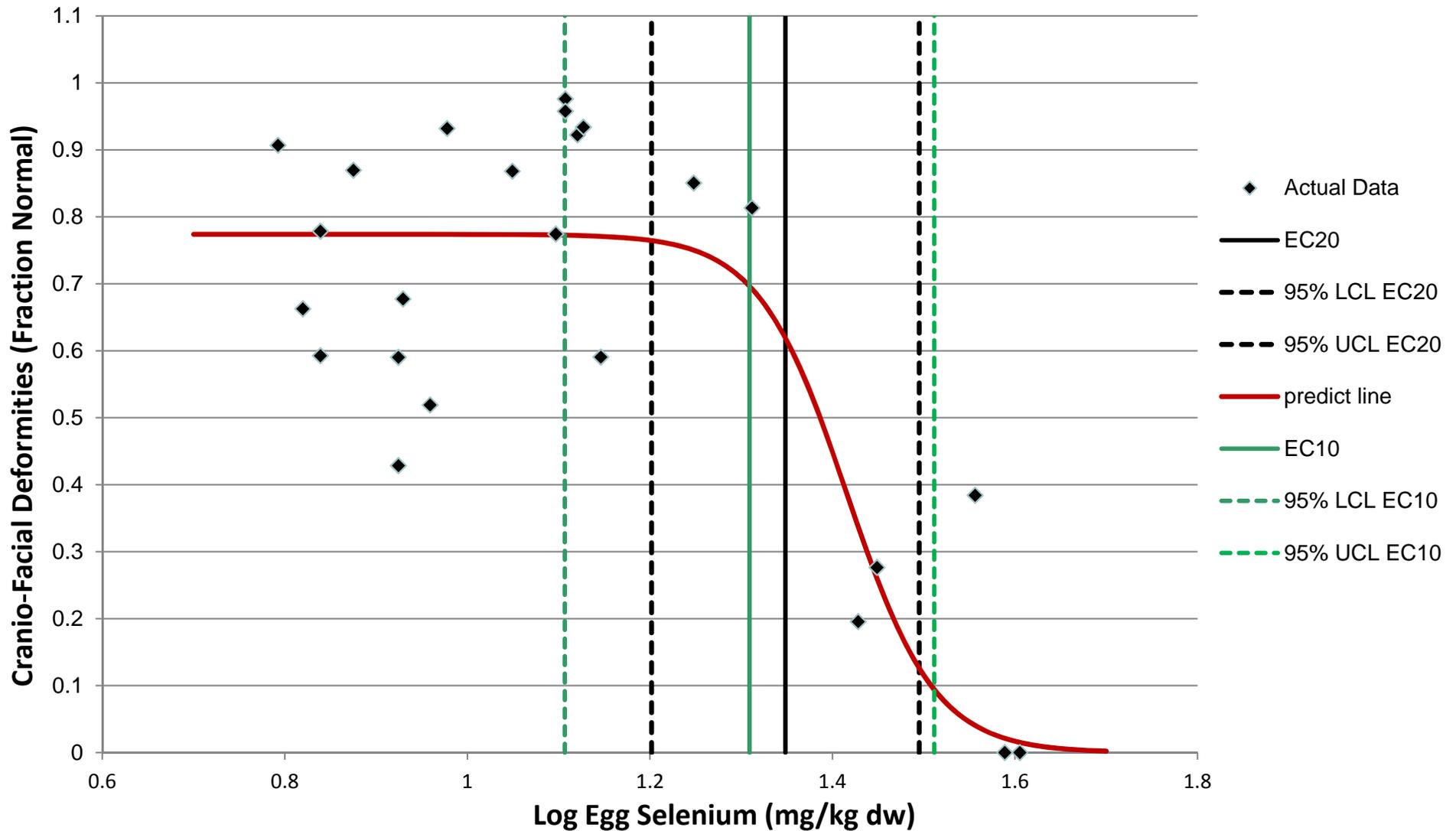


Figure 40
TRAP Model
Egg Selenium Concentration Versus
Cranio-Facial Deformities (Fraction Normal)

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC

FORMATION
 ENVIRONMENTAL

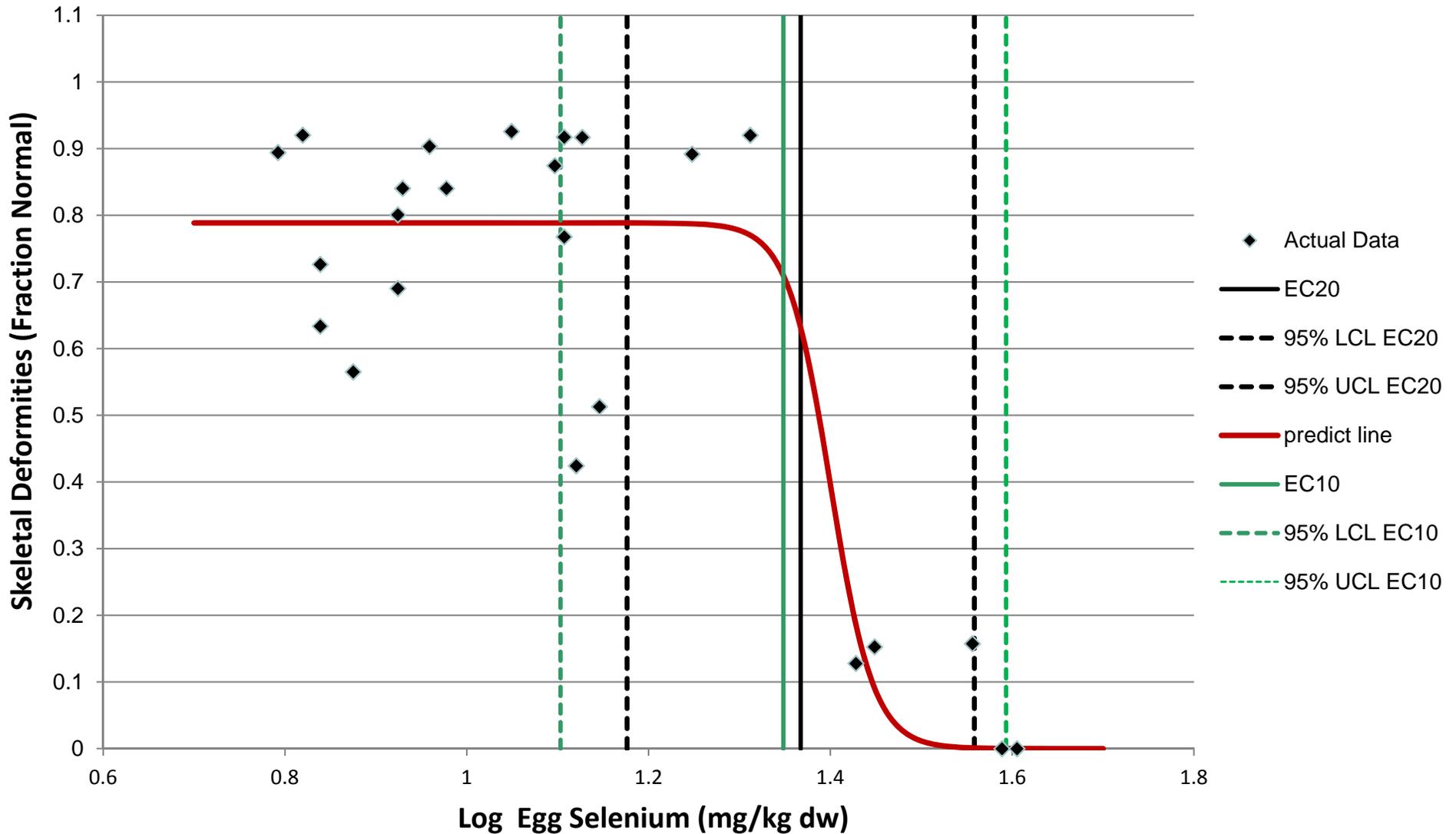


Figure 41
TRAP Model
Egg Selenium Concentration Versus
Skeletal Deformities (Fraction Normal)

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC

FORMATION
 ENVIRONMENTAL

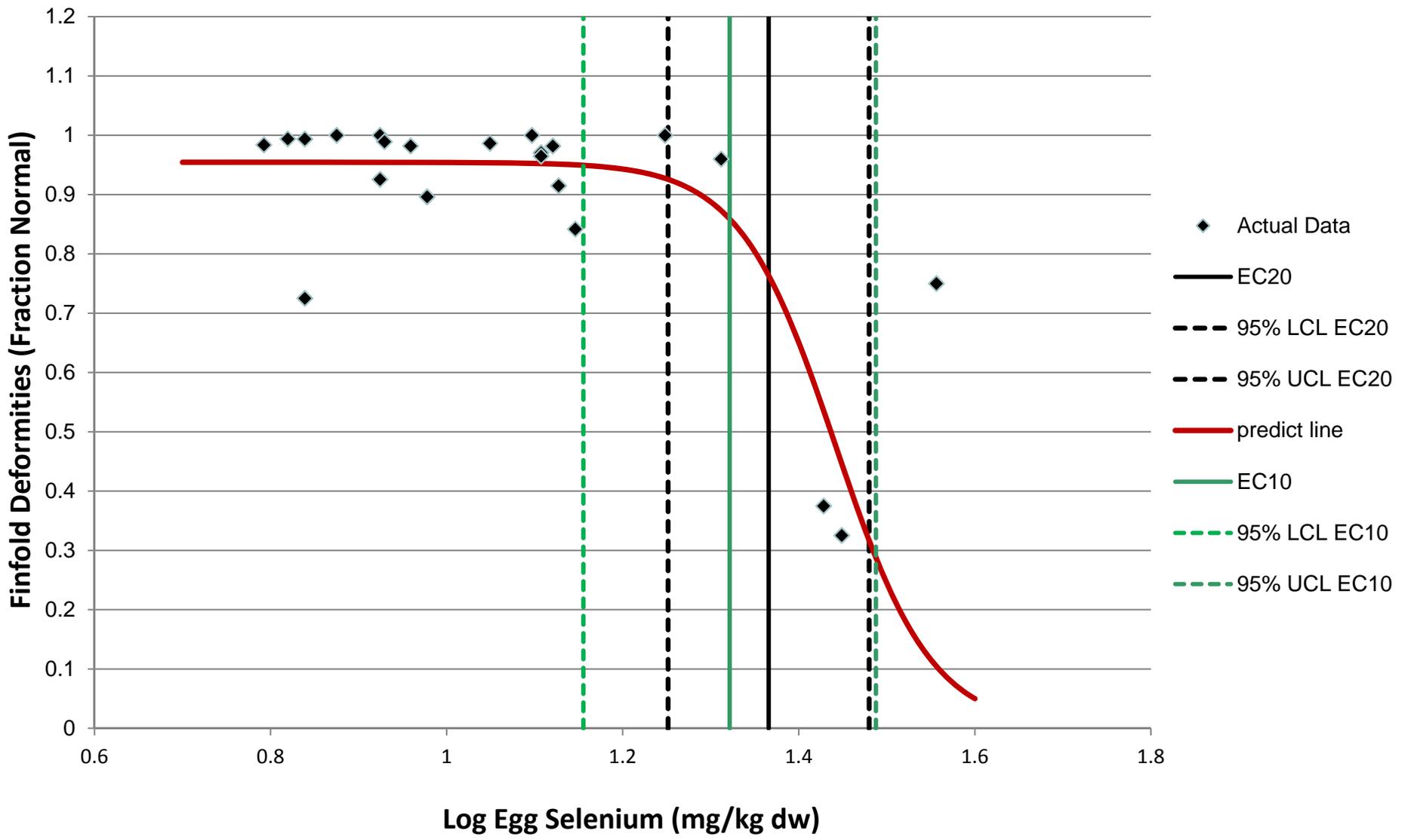


Figure 42
TRAP Model
Egg Selenium Concentration Versus
Finfold Deformities (Fraction Normal)

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70	DATE: June 2011	
REV: 1	BY: SMC	CHK: SMC



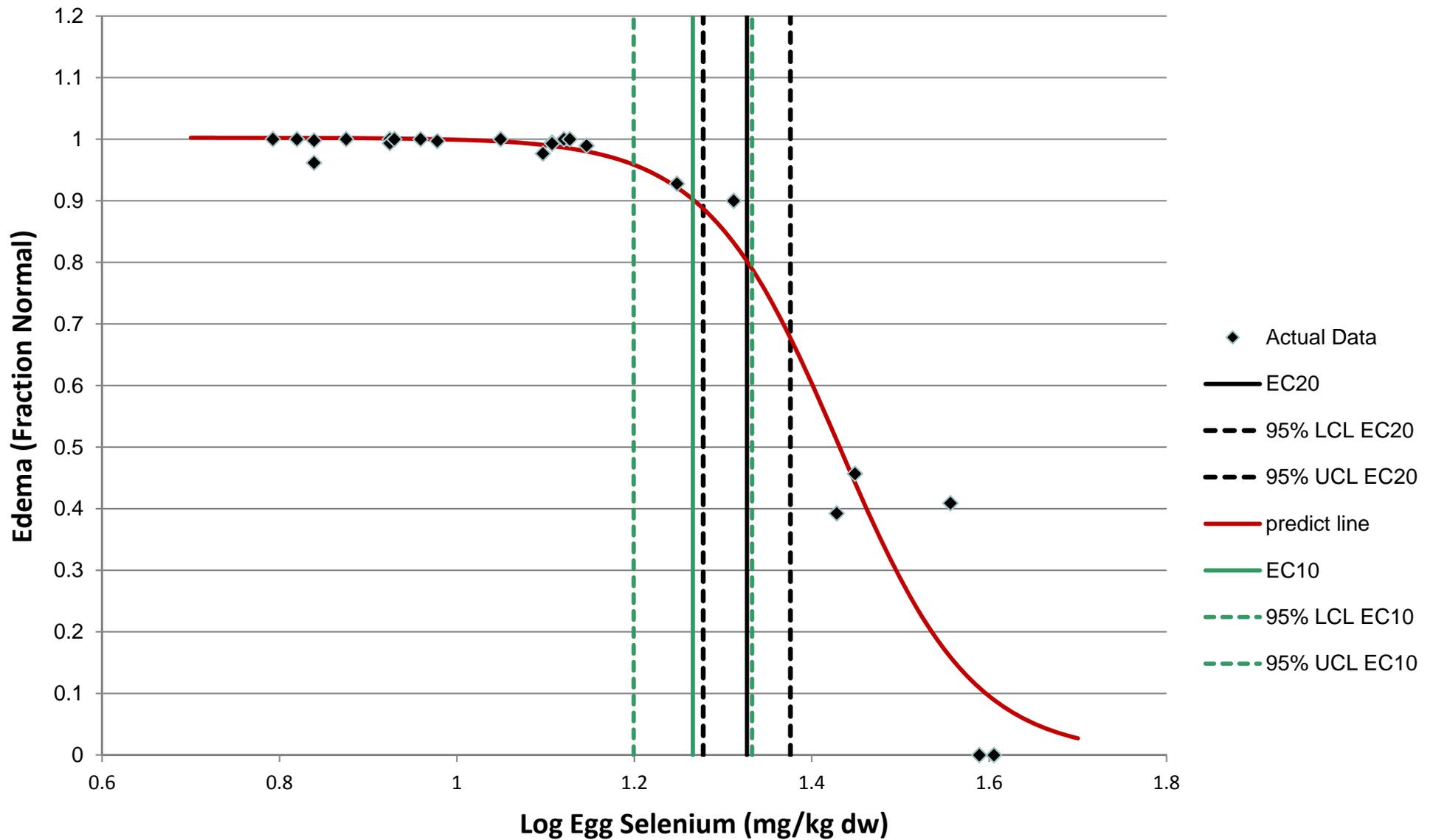


Figure 43
TRAP Model
Egg Selenium Concentration Versus
Edema (Fraction Normal)

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC



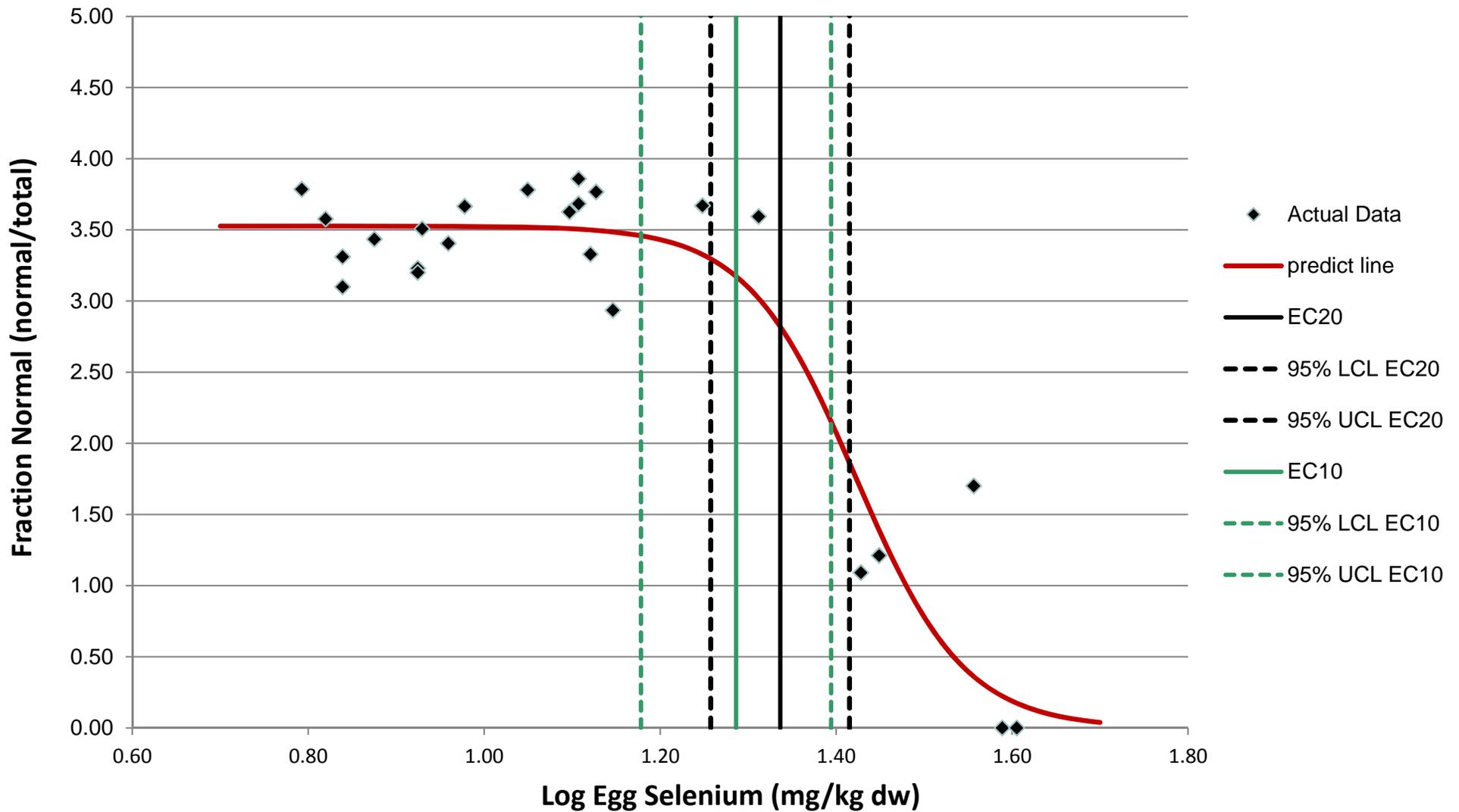


Figure 44
TRAP Model
Egg Selenium Concentration Versus
Sum Fraction Normal Fish Larvae

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC



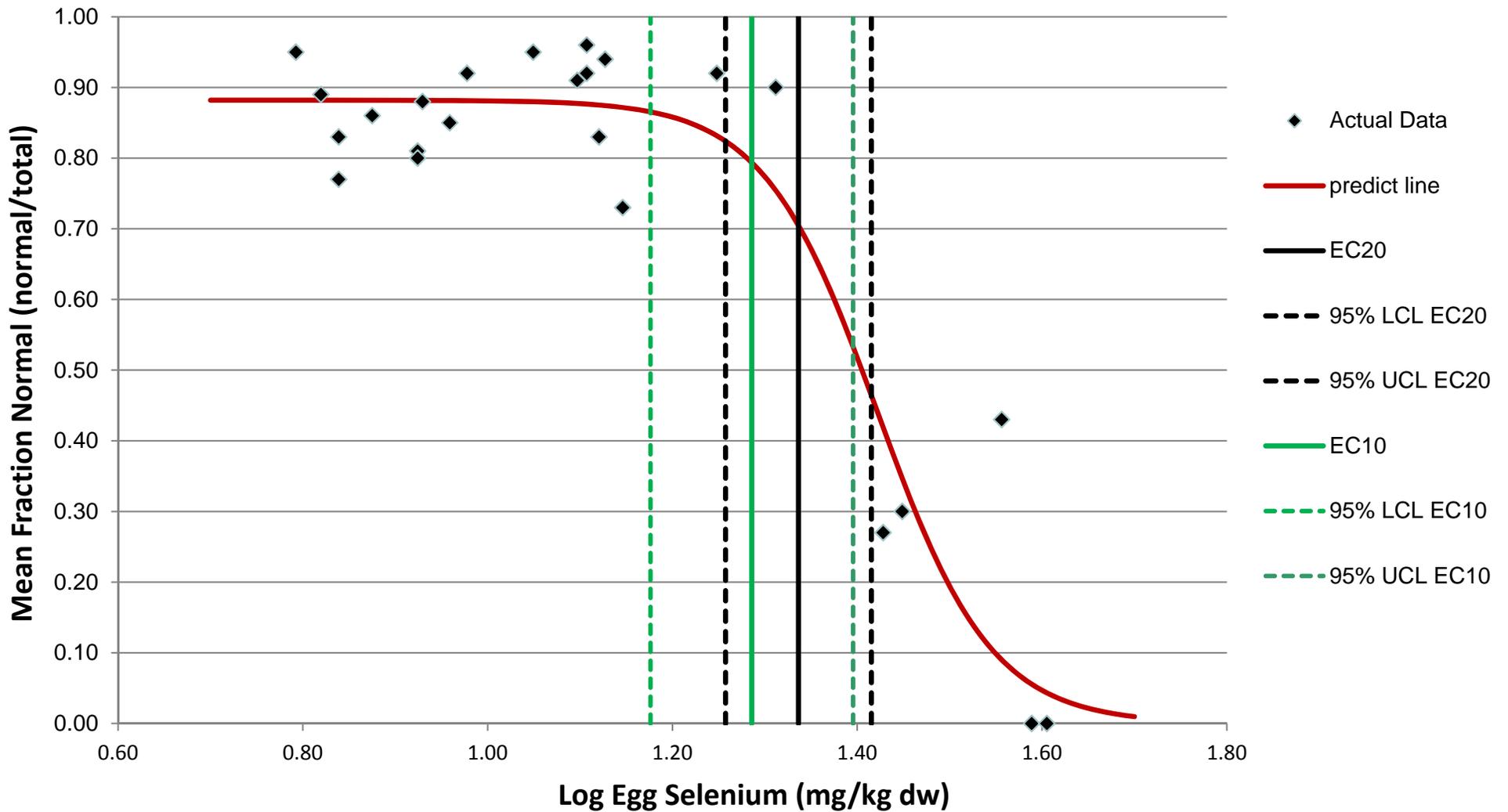


Figure 45
TRAP Model
Egg Selenium Concentration Versus
Mean Fraction Normal Fish Larvae

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC

FORMATION
 ENVIRONMENTAL

Egg Selenium Versus % Survival (Hatch to Test End)

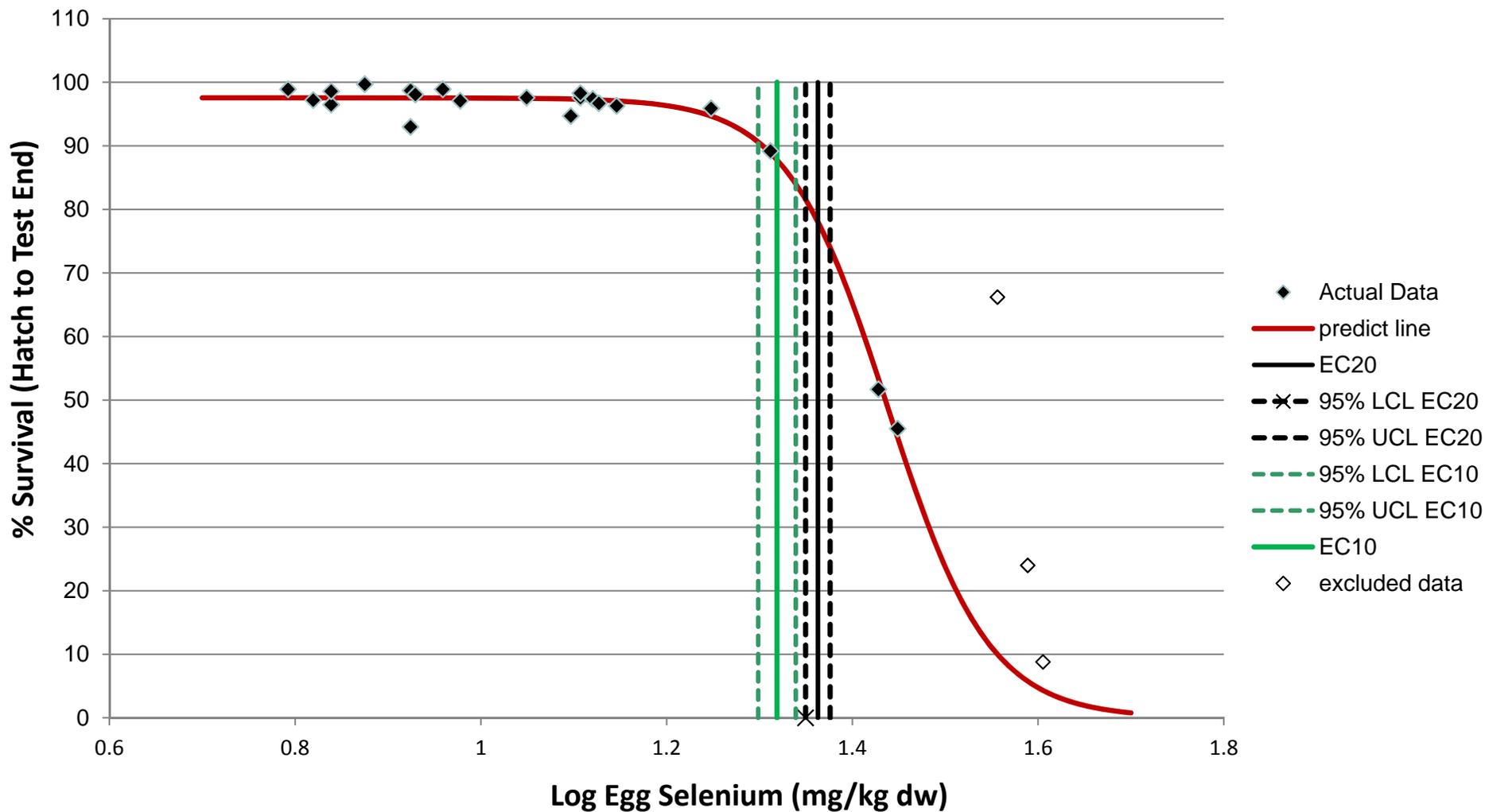


Figure 46
Revised TRAP Model
Egg Selenium Concentration Versus
Percent Survival (Hatch to Test End)

J.R. Simplot Company
 Site-Specific Selenium Criterion

PRJ: 0442-004-900.70

DATE: June 2011

REV: 1

BY: SMC

CHK: SMC



APPENDIX A
Data Report-Reproductive Success Study with Brown Trout (*Salmo trutta*)-ENSR

Prepared for:
JR Simplot Company and Newfields Company, LLC.
Pocatello, Idaho



Data Report

Reproductive Success Study with Brown Trout (*Salmo trutta*)

Final

October 2011

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1.0 Introduction

A study of brown trout (*Salmo trutta*) reproduction was conducted by Newfields for the JR Simplot Company to evaluate the parental transfer of selenium on the potential effects to offspring. ENSR's Fort Collins Environmental Toxicology Laboratory (FECTL), Fort Collins, CO was retained to conduct the laboratory biological exposure portions of this study according to the study design plan outlined in the Technical Memorandum – Methods for Testing Adult Brown Trout Reproductive Success (Newfields 2007). An assessment of larval trout deformities was performed by Dr. Kevin Bestgen at Colorado State University's Larval Fish Laboratory, which is described in a separate document. This report presents the results / data from the laboratory portion of this work.

2.0 Methods

ENSR FCETL personnel joined the Newfields team during the November 2007 sampling trip and provided assistance with fish collection, egg fertilization, and transport of egg samples to the laboratory. The time sensitive nature of transporting fertilized eggs from remote areas to the laboratory required integration of laboratory and field staff in this effort and near immediate transport of the samples to the laboratory.

Hatchery fish and gametes were obtained from Saratoga National Fish Hatchery (Saratoga, WY). A second set of hatchery fish were obtained in December 2007. This second batch of hatchery fish were later obtained (as eyed-eggs) from Spring Creek Trout Hatchery (Lewistown, MT).

Photographs taken at various points during the study are included in Appendix A.

2.1 Spawning of Brown Trout

Fertilization techniques for hatchery fish were slightly different from those of the field collected fish because of problems encountered with the SC hatchery eggs (lower than expected survival rates of eggs). ENSR engaged various hatchery and fishery personnel for recommendations on the fertilization technique. Based on these recommendations, the field methods were altered to fertilize eggs in the field instead of bringing the individually collected gametes under oxygen back to the laboratory and mixing them to achieve fertilization.

2.1.1 Hatchery Trout

Hatchery fish and gametes were obtained from Saratoga National Fish Hatchery, Saratoga, WY (courtesy of Lee Bender) on October 23, 2007. Throughout this study, fish from this hatchery are identified as SC. Because hatchery fish were obtained when they were ripe, which occurred prior to when fish were spawning in the field, the initial hatchery fish were obtained approximately 3 week prior to the first field collected fish.

Fertilization techniques for the first set of hatchery fish (SC) methods described by Holm et al. (2005). Eight adult female and eight male trout were anesthetized using tricaine methanesulfonate (MS-222) and stripped by hand (similar to treatment of field fish described below). Eggs from a given female were collected directly into a cleaned plastic pan and then transferred into a plastic bag. Bags were labeled for that individual. Milt from a single male was collected directly into a small plastic bag. Bags were labeled with the individual identifications for each fish and the collection location and date. Prior to transport to the laboratory, all bags with gametes were partially filled with oxygen, sealed, double bagged, and then placed on ice (~4 °C) in a cooler to keep gametes cold and out of direct sunlight. A min-max thermometer (Taylor® Digital Wireless Temperature System) was placed into the cooler with eggs to monitor temperature during transit to ENSR. The temperature range during transport of the SC eggs from Saratoga National Fish Hatchery and the FCETL was 6.1 – 9.7 °C; with 9.7 °C the initial reading when placed into the cooler.

After stripping, the individual adult females (n=8) from the Saratoga National hatchery were collected for determination of total length, weight, percent solids, and whole-body selenium analysis.

Sacrificed adult female trout were placed in large plastic bags, double bagged, and then stored on ice for transport to ENSR. Total length measurements were taken at ENSR prior to freezing samples for shipment to the analytical laboratory for analysis. Wet weight measurements were not measured at the laboratory but were estimated using the wet weight data from CAS and the wet weight data from the field fish (Appendix B).

Once received at ENSR's FCETL (Tuesday, October 23, 2007), eggs were maintained in coolers on ice overnight (temperature range overnight was 6-7 °C). The following day, eggs from each hatchery female were fertilized using the methods similar to those described by Holm et al. (2005; Appendix C). This method entailed placing 'green' or unfertilized eggs into a pre-cleaned plastic bowl (32 oz Rubbermaid plastic container). Milt (~1 ml, composite from all male fish) was combined with the eggs at a rate of 10 ul / 50 ml. The eggs and milt were gently stirred with a glass rod and allowed to stand for ~60 seconds. The egg/milt mixture was covered with ~100 ml of laboratory process water (i.e., hardness [~50 mg/L as CaCO₃] adjusted Horsetooth Reservoir water cooled to 6 °C), swirled for approximately three minutes, after which 500 ml of cooled laboratory water was poured over the eggs. The eggs were allowed to water harden for approximately five minutes. After water hardening, the fertilized eggs were kept in the Rubbermaid plastic containers with lids loosely fastened, placed back into the coolers, and maintained in the dark to gradually warm up to ~10 °C over the next 24-hours. On Thursday October 25, 2007, 600 eggs were collected from each batch of eggs using egg pickers and placed in prepared egg cups. Egg cups were then placed in individual test chambers. Remaining eggs not used for the test were then counted and frozen until they could be sent to Columbia Analytical Services (CAS), Inc. (Kelso, WA) for total selenium and percent solids analysis. Eggs for the SC hatchery fish were estimated using an egg counting technique developed for this purpose (Appendix C). Briefly, we counted the number of eggs for a given female that would fill a graduated cylinder to a particular volume (either 100 ml or 50 ml). Then we poured all the remaining eggs into a graduated cylinder to measure the total egg volume for that female. Using the number of eggs in either 50 or 100 ml, we determined the number of eggs in the total volume of eggs for that female. Since eggs from different females were of different size, this method was completed separately for each egg batch.

Because of low survival rates associated with the first set of hatchery eggs, a second set of hatchery eggs were obtained from Spring Creek Trout Hatchery (Lewistown, MT) on December 4, 2007. These eggs were obtained at the eyed-up stage because individual gametes were not available from Saratoga National fish hatchery or any other hatchery. These eggs served as method controls for the experimental system to ensure that we could expect acceptable survival and growth given the design. There were lower than expected survival rates for some treatments from the SC hatchery fish, which we believe were related to the fertilization and acclimation methods employed. Egg counts were not performed for the SPC eggs, as these were not obtained from individual female fish and thus not comparable with other treatments.

2.2 Laboratory Reproduction Tests

The temperature range for the cooler (#1) that held the trout from LSV2C (received at ENSR on November 15, 2007) was 3.4 – 8.7 °C (excluding the first 10 min acclimation period). The

temperature range for the second cooler that contained the remaining field eggs (received at ENSR on November 16, 2007) was 5.1 – 10.3 °C. Once back at the laboratory, eggs were allowed to slowly warm up over a 24-hour period to ~10°C (range of 6.2 – 8.4°C for cooler #1; range of 7.9 – 8.9 °C for cooler #2) prior to introduction into egg cups and test chambers. All remaining eggs were counted and frozen until they could be sent to the analytical laboratory for analysis of total selenium and solids. A list of the different locations from which fish were collected (i.e., treatments) and the individual identifications for each are provided in the table below (Table 2-1).

Table 2-1. Brown trout treatments and sample identifications for individual lots of fish eggs used in the reproductive study.

Saratoga NF Hatchery (SC)	Spring Creek Hatchery (SPC)	Background Se Field Location (CC-150)	Low Se Field Location (CC-350)	High Se Field Location (LSV2C)
SC-001	SPC-001	CC-150-009	CC-350-006	LSV2C-002
SC-002	SPC-002	CC-150-011	CC-350-007	LSV2C-003
SC-003	SPC-003	CC-150-012	CC-350-008	LSV2C-004
SC-004	SPC-004	CC-150-013		LSV2C-005
SC-005	SPC-005	CC-150-015		LSV2C-006
SC-006	SPC-006	CC-150-016		LSV2C-007
SC-007		CC-150-017		LSV2C-008
SC-008		CC-150-018		LSV2C-010
		CC-150-020		LSV2C-012
				LSV2C-016
				LSV2C-017
				LSV2C-019
				LSV2C-020
				LSV2C-021

Note: CC – Crow Creek; LSV – Lower Sage Creek

After the 24-h temperature acclimation period, a target of 600 eggs were transferred (in low ambient light) from each batch of field collected eggs using egg pickers, and placed in prepared egg cups. Egg cups were then placed in individual test chambers in the water bath. The remaining eggs for each treatment not used for the test were then counted by hand (Appendix C) and frozen until they could be sent to Columbia Analytical Services (CAS), Inc. (Kelso, WA) for total selenium and percent solids analysis.

Eggs were transferred from collection bags to egg cups in low ambient light using egg pickers. The egg cups were submersed in ~10°C water. Egg cups were constructed of polyvinyl chloride (PVC) schedule 40 pipe (approximately 5 cm ID and 3.8 cm depth) with a nitex screen bottom. Ten individual units were attached in a 2 x 5 layout design using silicon, so that each egg cup consisted of 10 individual cells (Figure 2-1). Eggs were evenly distributed into all 10 of the cells of the egg cups. For instance, the treatments initiated with 600 eggs had 60 eggs placed into each egg cup cell. While the original intent was to maintain the ten replicates for a given fish throughout the study, this was not feasible due to the water demands and space limitations. Therefore, organisms from all replicates were combined in the test chamber at hatch out.

Egg cups were hung with clips and fishing line in Sterilite® plastic test chambers (11.4 L). Each chamber was aerated for the duration of the test to maintain the dissolved oxygen concentration at

sufficient levels (>60%). The volume in each test chamber was approximately 5 L maintained at the level of the top of the drain pipe which consisted of a piece of 5-mm ID glass tubing inserted through a silicone stopper which is pressed into a small hole drilled in the side of the chamber. Chambers and water volume were of sufficient size to maintain a loading rate of < 5 g of fish per L of water in each test chamber. Spent water overflowed out of the glass standpipes and into the water bath before being discharged directly to a conduit connecting both baths. This water was treated with an ultraviolet light disinfection unit prior to discharge into the laboratory waste water. After swim-up occurred, the drain openings were covered with a small piece of nylon mesh to prevent loss of organisms. In general, methods employed for this study followed ASTM (2006) standard guidance for conducting early life stage tests with fish, although modifications were made to account for study-specific hypotheses and test design criteria (e.g., number of eggs).

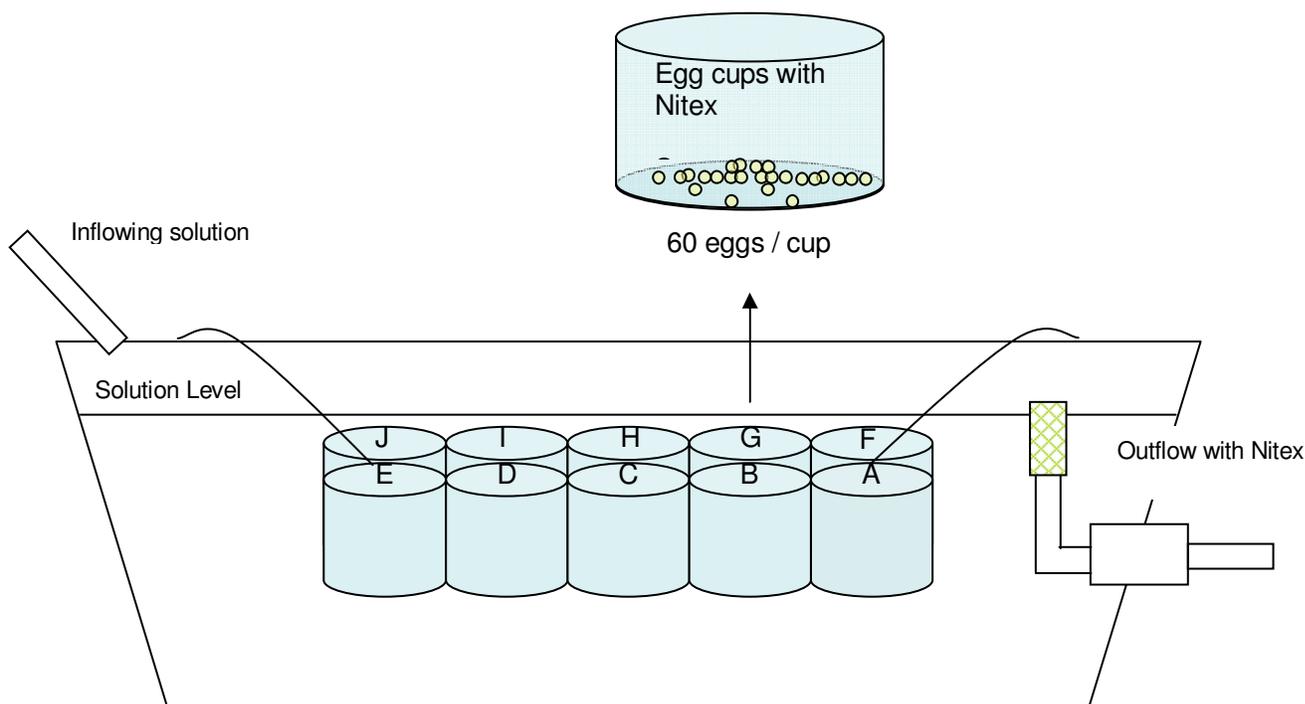


Figure 2-1. Schematic diagram of the test chamber and egg cups for brown trout reproductive study. Inset shows individual cells of egg cups ($n = 10$) within a chamber. Aeration tube not shown.

With the test solution volume of ~5 L and a flow rate of 20 ml/min, each test vessel received ~ 5.7 volume additions per day. Test chambers were held in two separate water baths with the temperature of the water baths controlled by chillers. Chambers were randomly placed in one of the two water baths so that each bath had an equal number of chambers (or as close as possible) from each treatment. Chambers within a given bath were then randomly placed into different locations within the water bath (Figure 2-2).

Figure 2-2. Location of egg treatments in water baths for brown trout reproductive study.

Bath #1		Bath #2	
HIGH002	BK013	SC004	HIGH004
LOW008	SC005	HIGH019	SC003
BKD020	HIGH010	HIGH008	BK012
HIGH016			HIGH005
HIGH021	SPC004*	SPC006	
SPC005	BK017		SC006
		HIGH017	BK016
	LOW006	BK015	BK009
SC007	BK011	LOW007	SC002
SPC001	BK018		
SC008	HIGH012	SPC003	SPC002*
SC001	HIGH003	HIGH020	HIGH007

*denotes smaller test chamber (~2.5 L)

Note: BKD = CC-150, LOW = CC-350, and HIGH = LSV2C

Two smaller glass chambers (2.5 L) used previously in ELS studies were also initiated with eggs from SPC (20 eggs per egg cup / chamber, 20 ml/min flow). These two chambers were included as an additional set of performance controls to compare with SPC eggs initiated using the regular design (i.e., 600 eggs per egg cup per chamber)

The exposure chambers were housed in temperature-controlled water baths. Target temperatures in the test chambers were $10 \pm 2^\circ\text{C}$. Dissolved oxygen concentrations were maintained at ≥ 60 percent of saturation (5.6 mg/L at 5,200 feet elevation and 10°C). Embryos and fry were maintained under dim lighting (approximately 0.8 foot-candles) until swim-up occurred, after which they were held in ambient lighting (approximately 16 ft-c) with 16 hours of light per 24-hour period.

Egg cups were maintained submerged in each test chamber until all eggs hatched or were noted as dead. Originally, dead eggs were not going to be removed until after all live eggs eyed up. However, because of the observance of fungus on some of the dead eggs, dead eggs were carefully removed from the egg cups prior to the eyed up stage in some instances. The number of dead eggs removed each day was recorded for each test chamber. Eggs or embryos were considered dead if they appeared opaque and/or developed visible fungal infections. As hatching occurred, the numbers of live alevins, as well as the dead alevins (or eggs) that were removed, were recorded on a daily basis. However, because of the movement of the alevins due to their own swimming ability as well as from the movement of the water from aeration it was difficult to get an accurate count on the number of newly hatched alevins on a daily basis. Since this measurement was unreliable, it was discontinued and counts were largely based on the number of dead organisms removed from a particular chamber. When eggs hatched, alevins were gently removed to the bottom of the surrounding test chamber using a large bore glass pipette and the remaining egg shell was removed. Organisms that died as eggs or while hatching were recorded and preserved in Stockard's solution. Eggs that had the amniotic fluid (e.g., yolk) leak out just about the time of hatching were termed, 'dead while hatching' (DWH). Any organisms that were not found during the test were considered dead, except during the 15-d swim-up study. For this phase of the

study, any missing organisms were considered missing and were excluded from survival analysis. Fish were considered dead if no gill movement or visible response was observed in response to gentle prodding. Egg cups were removed after all living eggs hatched. Test initiation and termination dates for each treatment are provided below (Table 2-2).

Table 2-2. Test initiation dates and termination dates for brown trout treatments in the reproduction study.

Fish Treatment / Location	Test Initiation Date	Test Termination Date(s)
SC	Oct. 25, 2007	Jan. 17, 2008
SPC	Dec. 4, 2007	Jan. 22, 2008
CC-150 (BKD)	Nov. 17, 2007	Feb. 7 & 12, 2008
CC-350 (LOW)	Nov. 17, 2007	Feb. 7 & 12, 2008
LSV2C (HIGH)	Nov. 16, 2007	Feb. 7 & 12, 2008

Eggs (primarily SC treatments) were treated with salt (NaCl) and, later, formalin in an attempt to reduce fungal growth. Days and type of treatment are located in Appendix D. Fungus appeared to affect the SC treatments more than later field-collected (i.e., wild) eggs and the second set of hatchery treatments as additional UV disinfection systems were incorporated at other locations prior to initiation of these other treatments.

Alevins (recently hatched young with yolk sacs) were monitored daily for mortality. Dead organisms were removed and placed in Davidson's solution. As alevins approached swim-up, trout chow was offered to the organisms to determine if they were actively feeding. The swim-up date was set based on when at least 80% of the alevins had absorbed their yolk sac and were actively feeding. At the swimup stage, organisms were thinned down to a target of 100 organisms per test chamber, preserving all the extra organisms in Davidson's solution for the deformities assessment. If there were less than 100 organisms in the test chamber then organisms were counted and left in the test chamber; however, no organisms were preserved at this stage for deformities analysis. All living larval fish were then maintained for the 15-d post swim-up stage of the study. Dead organisms were counted and removed daily, saved by placing them in Davidson's solution. Swim-up trout were started on a 4% body weight ration of salmon starter #1 (purchased from Aquatic Biosystems, Fort Collins, CO) over three feedings daily (i.e., morning, noon, evening) during the week, and at least two feedings daily on weekends. Weight of a swim-up fry was determined by sacrificing one fish out of seven hatchery treatments (SC) and determining a wet weight. The wet weight for these seven fish averaged 0.105 g; therefore, daily feeding was ~0.4 g trout chow assuming 100 fry.

At initiation of the swim-up stage the flow rate into each chamber (except the 2.5L chambers) was altered to 40 ml/min and taller stand pipes were added to adjust the total volume to ~9 L to account for loading requirements based on the anticipated growth of the organisms. Loading for the hatchery fish was < 2.5 g/L (assumes a wet wt of 0.2 g for 100 fish in 8 L of water). Once feeding

started, test chambers were siphoned daily (in p.m. prior to feeding) to remove remaining food and fecal material.

At the end of the 15-d post swim-up study, all remaining organisms were sacrificed via immersion in isopropyl alcohol. A sub-set of 20 organisms was rinsed with deionized water, blotted dry and measured for standard length (tip of snout to caudal peduncle). All remaining organisms were preserved in Davidson's solution for deformities assessment.

Length measurements were taken on the day of test termination for all organisms except the SC-001 through SC-007 fish. Length measurements for these organisms were taken after a few days in isopropyl alcohol because they could not all be measured on the day of test termination. Length and wet weight measurements on 10 fish from SC-008 chamber were taken prior to storage and after storage to see if storage in alcohol altered the length measurement statistically (pre-isopropyl storage avg = 22.6 mm, post-isopropyl storage avg. = 21.2 mm, $p = 0.0055$). Because of the difference in lengths pre- and post-preservation, all remaining organisms were measured for length prior to preservation. Following length measurements, organisms were preserved in isopropyl alcohol until dry weight could be determined. For dry weight analysis, each fish was transferred to a tared weight boat and dried at 100 °C for at least 48 hours. After removal from the drying oven, the weigh boats were placed in a dessicator to prevent absorption of moisture from the air, until weighed (dry weight) to the nearest 0.01 mg.

2.3 Dilution Water

The dilution/control water used in this study was FCETL process water obtained from Horsetooth Reservoir. The ambient incoming water is coarse-filtered (through a sand filter and polypropylene core filters [10 and 1 micron]) to remove indigenous organisms, particulate matter, and contaminants. Water then passes through an ultraviolet light disinfection system before being stored in large holding tanks. This water is periodically analyzed for contaminants. Horsetooth Reservoir process water is very soft to soft water according to USEPA (2002), with both hardness and alkalinity typically 20 - 30 mg/L as CaCO₃ (Table 2-3). Background sulfate levels in unaltered Horsetooth water are ~5.0 mg/L. Ambient (unheated) laboratory Horsetooth reservoir water was metered into a large holding vessel (~100 gallons) and chilled with a counter-current cooling process to help maintain the target water temperature in the test chambers of 10 °C. Water from the holding vessel was adjusted to increase the hardness and sulfate so that it would be higher than ambient levels and more similar to the field conditions (Table 2-3). Given the soft water conditions of the laboratory Horsetooth water and the volume of water used on a daily basis, it was impractical to match the water quality characteristics of the site.

Table 2-3. Water quality measurements for Horsetooth Reservoir process water (unamended) and Crow Creek Drainage.

Parameter	Horsetooth Reservoir ¹		Crow Creek ²	
	Average	Range	Average	Range
Hardness (mg/L)	33	26.5 – 41.8	171	129 – 220
Alkalinity (mg/L)	28.8	25 – 33	197	140 – 231
Sodium (mg/L)	3.5	2.7 – 5.5	3.3	1.0 – 6.5
Potassium (mg/L)	<1	---	<1	<1 – 1.8
Sulfate (mg/L)	5.6	3.4 – 10	27	7.5 – 48.7
Chloride (mg/L)	1.9	0.5 – 3.6	7.3	0.2 – 89
DOC (mg/L)	2.4	2.1 – 2.9	1.0	0.34 – 2.18

¹Horsetooth Reservoir laboratory process water (Fort Collins, CO) from 2000 to 2004 measured at ENSR's FCETL.

²Crow Creek drainage as characterized by surface water from Crow Creek, Sage Creek, and Hoopes Spring surface water quality data (Newfield 2007)

Calcium sulfate ($\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$; Ben Franklin[®] Aquacal[™], Plaster City, CA) and magnesium sulfate ($\text{MgSO}_4 \cdot 7\text{H}_2\text{O}$; The PQ Corporation, Valley Forge, PA) were added at a ratio of 1.82:1 calcium:magnesium (molar basis) to deionized water to prepare a super hardness stock solution of ~2,000 mg/L as CaSO_3 . This super stock was metered into the holding vessel to achieve a target hardness of ~50 mg/L as CaCO_3 and sulfate concentration of ~20 mg/L. Water hardness was measured daily during the study, while sulfate concentration was monitored periodically.

The super stock solution of extremely hard Horsetooth water (~2,000 mg/L hardness) was metered from two Mariotte bottles (5-gallon glass bottle) into a head tank which then was metered into the large holding vessel along with dilution water inflow. The inflow of the ambient laboratory Horsetooth water was approximately 1,400 ml/min. The super hardness stock solution was delivered into the holding vessel at a target rate of 14 ml/min. Batches of the super hard stock solution were prepared every three days and the Mariotte bottles were filled daily throughout the study. Flows on the main dilution water (unadjusted Horsetooth water) and the drip flowing from the head tank (Mariotte bottles fed this tank) were measured at least once daily throughout the study.

Water from the holding vessel, now adjusted to a water hardness of ~50 mg/L and slightly cooled, was then pumped over to a head tank set up above the dilutor panels and water baths holding all test chambers. Flows into the head boxes above each dilutor panel feeding the test chambers for that bath were maintained by providing a constant head pressure using a submersible pump from the holding vessel and an over-flow recirculating system (i.e., excess flow was returned from the head tank above the test chambers back to the 100 gal holding vessel).

Water from the head tank flowed into diluter panels constructed out of glass, silicone adhesive, and silicone stoppers. Adjusted Horsetooth process water was delivered to the test chambers through (3/8 I.D. x 1/2 O.D. x 1/16 thickness, inch) polyethylene tubing. The dilutor panel delivered modified Horsetooth water to up to 24 test chambers per water bath. Flow rate into each chamber was

adjusted in the splitter box to deliver a target rate of 20 ml of test solution per minute to each chamber. After swim-up had occurred and the 15-d post swim-up study was underway, the flow rates were adjusted to a target of 40 ml of test solution per minute per chamber.

2.4 Water Chemistry

Temperature (°C), pH (s.u.), dissolved oxygen (mg/L) concentrations, and conductivity (µS/cm) were measured and recorded in one chamber for each test treatment daily. Hardness (as mg/L CaCO₃) was measured from the dilutor panel or from a test chamber daily during the study. Total ammonia (mg/L as N) was measured in selected test chambers (LSV2C-010, CC-150-016, and CC-350-006) once feeding was initiated. Sulfate concentration was measured from water collected from the dilutor panel or from test chambers. Determinations of waterborne sulfate concentrations were made at Paragon Analytics, Inc. (Fort Collins, Colorado, USA) using ion chromatography (EPA Method 300.0).

Water samples for total recoverable and “dissolved” selenium analyses were collected, prepared, and preserved from selected test chambers during the course of the study. Briefly, approximately 50 to 250 ml of test solution was collected for analysis of either dissolved or total selenium analysis. Aqueous analytical samples were analyzed at either Paragon Analytics Inc. or ACZ Laboratories (Steamboat Springs, CO). Dissolved selenium samples were filtered through 0.45 µm filters (GHP Acrodisc Syringe Filters, Pall Gelman Scientific, Ann Arbor, MI, USA) prior to placing in the polypropylene sample containers and preserved with 1% nitric acid. Samples were analyzed at ACZ using an ICP-MS (EPA Method 200.8) or at Paragon Analytics, Inc. using method SW3005A (ICP-OES).

Aqueous water samples were also collected at the hatchery sites (SC and SPC) for analysis of total and dissolved selenium so background levels of selenium at the hatcheries could be compared with selenium levels from other fish areas. These samples were collected in April 2008 (SC) or October 2008 (SPC) and sent to ACZ for analysis. Total recoverable and dissolved Se concentrations in the water from the Saratoga National Fish Hatchery was 1.7 µg/L, while total recoverable and dissolved Se concentrations in the water from Spring Creek hatchery was <0.1 µg/L (Appendix E).

2.5 Deformities Assessment

Extra fry (excluding the target of 100 fry kept for the post swim-up phase of the study) were removed and preserved in Davidson’s Solution at swim-up for deformity examination. Any deformed fry were removed at this point and preserved as part of the extra fry. Upon test termination, an additional batch of fish (per treatment) were preserved similarly and saved for deformity assessment. Of the 100 organisms included in the 15-d post swim-up phase of the study, the target was to save 80 of these fry for deformity assessment (the other 20 were for length and dry weight analysis).

Dead fish and alevins were removed during the study and preserved for deformity assessment as well. However, many of these organisms did not preserve well because they were in various states of decay. Because of the poor tissue condition of these dead organisms they were not originally evaluated (i.e., necrotic tissue conditions and/or presence of fungus made analysis and observations difficult). A subsequent analysis was conducted on approximately 100 of these fish (per batch) on five samples in which there was no or little data. These additional samples were from LSV2C -003, -004, -005, -010, and -021. All samples for deformity analysis were sent to Dr. Kevin Bestgen at CSU. Data from these samples were incorporated in the deformity assessment performed.

2.6 Endpoints

Multiple test endpoints were utilized for this test at different times during the test. Fecundity, hatch, deformities, length, weight, survival (different times during the study), tissue concentrations (egg and whole body), and feeding success were proposed test endpoints. These endpoints were similar with those of Holm et al. 2005, Hardy 2005, and Kennedy et al. 2000 on which the test described herein was based.

Total egg production for each female was counted as a measure of fecundity. Survival was determined based on the number of surviving fish at hatch, swim-up, and at test termination compared to the number of eggs at test initiation. Percent hatch was determined as the number of live fish and alevins at day of first hatch compared to the number of eggs at test initiation. Other endpoints included day of swim-up, day of test termination, and measurements on survival larval fry at test termination (length and dry weight).

3.0 Results

3.1 Egg Analyses

The number of eggs used from a given female depended on the total number of eggs provided by that female. While the target was 600 eggs per female, certain organisms did not provide that many total eggs. Therefore, we attempted to maximize the number of eggs used in the reproduction study while leaving enough for selenium analysis. For treatments with fewer eggs (e.g., LSV2C-010), eggs were added to each replicate of the egg cup in small numbers (10 at a time) to ensure equal numbers in each replicate. Once that target number was added to each replicate egg cup cell, the number of eggs remaining was evaluated to see whether more eggs could be added to the egg cup. This process was repeated until no fewer than 61 eggs remained for Se analysis. For the field collected eggs, the most that remained was 732 eggs; however, the average number of eggs saved for Se analysis was 329. The number of eggs used in the study from a particular female, the total number of eggs the female produced, and the percent egg mortality are presented (Table 3-1).

One set of eggs were completely dead upon arrival (LSV2C-006). Because there were no viable eggs upon inspection, these eggs were not used in the reproductive study but were retained for selenium analysis. From 100 to 600 eggs were used to initiate the reproductive studies for the remaining fish sampled (Table 3-1). As mentioned, this corresponded to the total number of eggs produced by a particular female. The total number of eggs from field collected organisms ranged from 161 (LSV2C-010) to 1,658 (CC-150-016). By contrast, the range of eggs collected from the SC hatchery fish ranged from 1,248 to 5,448. The total number of eggs could not be counted individually for the SPC hatchery fish as they were previously composited from multiple females and delivered as eyed-up eggs.

Table 3-1. Estimated number of total brown trout eggs from adult female organisms used in the reproductive success study.

Location	Sample ID	#Eggs placed in study	Total # of eggs from fish	Egg Mortality (%)	
<i>Hatchery Fish</i>	SC-001	600	4,173	76.2	
	SC-002	600	4,005	77.2	
	SC-003	600	5,120	43.3	
	SC-004	600	1,248	68.9	
	SC-005	600	5,448	88.3	
	SC-006	600	3,176	6.8	
	SC-007	600	3,224	69.5	
	SC-008	600	4,005	34	
<i>Second set of</i>	SPC-001	600	--	0.7	
	SPC-002	20	--	0	
<i>Hatchery Fish</i>	SPC-003	600	--	2.5	
	SPC-004	21	--	0	
	SPC-005	600	--	1.2	
	SPC-006	600	--	0.3	
Wild	CC-150-009	600	1,215	71.5	
	CC-150-011	300	488	4.0	
	CC-150-012	350	556	11.2	
	CC-150-013	600	1,234	33.3	
	CC-150-015	600	1,003	21.7	
	CC-150-016	600	1,658	85.2	
	CC-150-017	250	414	10.8	
	CC-150-018	600	959	12.8	
	CC-150-020	600	1,332	2.7	
	CC-350-006	600	1,154	28.3	
	CC-350-007	600	1,174	70.3	
	CC-350-008	600	922	32.3	
	Fish	LSV2C-002	600	1,096	1.0
		LSV2C-003	400	474	6.5
		LSV2C-004	500	766	49.4
LSV2C-005		300	476	28.7	
LSV2C-006		--	--	100	
LSV2C-007		500	773	100	
LSV2C-008		300	372	11.7	
LSV2C-010		100	161	13	
LSV2C-012		600	1,031	1.7	
LSV2C-016		600	826	5.0	
LSV2C-017		300	447	28.7	
LSV2C-019	500	693	5.8		
LSV2C-020	400	525	10.8		
LSV2C-021	600	1,208	30.7		

3.2 Laboratory Study

3.2.1 Water Chemistry

The water quality parameters (pH, temperature and dissolved oxygen [DO]) monitored daily during the study were within acceptable ranges for the survival of brown trout (Table 3-2).

Table 3-2. Water hardness (avg \pm SD), dissolved oxygen (low and % saturation), pH (range), temperature, and conductivity measured in each treatment during the reproductive study using brown trout (*Salmo trutta*).

Fish Treatment	Water Hardness (mg/L)	Minimum Dissolved Oxygen (mg/L) & % Saturation	pH (s.u.)	Avg \pm SD Temp ($^{\circ}$ C)	Temp. Range ($^{\circ}$ C)	Conductivity (μ S/cm)
SC	49.3 \pm 3.2	7.8 / 83	7.4 – 7.9	10.1 \pm 0.7	8.5 – 11.9	104 – 196
SPC	48.6 \pm 3.6	7.5 / 80	7.4 – 7.9	10.5 \pm 0.7	8.6 – 11.9	106 – 146
CC-150 (BKD)	48.4 \pm 3.5	7.5 / 80	7.4 – 7.9	10.5 \pm 0.8	8.7 – 12.5	107 – 175
CC-350 (LOW)	48.4 \pm 3.5	7.4 / 79	7.5 – 7.9	10.6 \pm 0.8	8.8 – 12.2	104 – 167
LSV2C (HIGH)	48.4 \pm 3.5	7.1 / 76	7.3 – 7.8	10.6 \pm 0.8	8.7 – 12.5	107 – 161

Note: At 5,200 feet elevation and 10 $^{\circ}$ C, 60% dissolved oxygen saturation is 5.63 mg/L

Alkalinity was measured at least weekly in the laboratory Horsetooth dilution water and it averaged 28.2 ± 1.4 mg/L (as CaCO_3) between October 25, 2007 and February 12, 2008. Ammonia was measured in select treatments (LSV2C-010, CC-150-016, and CC-350-006) during the 15-d post-swim-up feeding portion of the study and was < 1.0 mg/L in all test chambers. Sulfate, measured 12 times over the course of the study, averaged 22.9 (range 17 – 27) mg/L (Appendix E). Water temperature measured in the chambers for each water bath and water hardness are presented over the course of the study (Appendix E).

Aqueous selenium measured in the hardness adjusted Horsetooth water or in specific test chambers from October 23, 2007 to February 12, 2008 were < 5 μ g/L (12 total measurements; Appendix E). The only time that selenium was detected was on January 22, 2008 for a collected sample from the SPC-001 chamber (result was 9.9 μ g/L). There was insufficient sample to have this value re-verified by a separate laboratory with lower detection limits.

3.2.2 Biological Endpoints

The day of first hatch for the SC hatchery fish ranged from 40 to 47 days, although all but two had started hatching by day 42 (Table 3-3). The second set of hatchery fish (received at the eyed up stage) hatched from nine to 13 days after receipt, while the treatment fish hatched from 36 to 43 days. The slightly lower temperatures for the SC hatchery fish likely explain the slightly longer day to first hatch.

Table 3-3. Day of first hatch, percent hatch, day of swim-up, percent swim-up, and percent survival at swim-up for brown trout fry from the reproductive success study. Eggs from the second set of hatchery fish were obtained at the eye-up stage.

Location	Sample ID	Day of 1 st hatch	% hatch ^a	Day of swim-up	% Swim-up	Survival (%) at Swim-up Stage	
<i>Hatchery Fish</i>	SC-001	42	23.8	69	22.8	22.8	
	SC-002	41	22.8	69	22.5	22.5	
	SC-003	40	56.7	69	55.7	55.7	
	SC-004	40	31.1	69	27.3	27.3	
	SC-005	42	11.7	69	10.7	10.7	
	SC-006	40	93.2	69	92.8	92.8	
	SC-007	47	30.5	69	26.8	26.8	
	SC-008	46	66.0	69	65.0	65.0	
<i>Second set of Hatchery Fish</i>	SPC-001	11	99.3	34	98.0	98.0	
	SPC-002	13	100	34	100	100	
	SPC-003	11	97.5	34	94.2	94.2	
	SPC-004	13	100	34	100	100	
	SPC-005	11	98.8	34	97.5	97.5	
	SPC-006	9	99.7	34	96.0	96.0	
Wild	CC-150-009	39	28.5	72	27.2	27.2	
	CC-150-011	42	96.0	67	95.3	95.3	
	CC-150-012	40	88.8	72	86.8	86.8	
	CC-150-013	39	66.7	72	59.7	59.7	
	CC-150-015	40	78.3	67	77.8	77.8	
	CC-150-016	43	14.8	72	14.3	14.3	
	CC-150-017	42	89.2	72	86.4	86.4	
	CC-150-018	41	87.2	72	84.3	84.3	
	CC-150-020	38	97.3	67	96.3	96.3	
	CC-350-006	41	71.7	72	68.0	68.0	
	CC-350-007	38	29.7	67	28.7	28.7	
	CC-350-008	37	67.7	67	64.5	64.5	
	Fish	LSV2C-002	36	99.0	68	96.7	96.7
		LSV2C-003	41	93.5	88	0 ^b	8.0
		LSV2C-004	40	50.6	88	0 ^b	30.2
		LSV2C-005	40	71.3	88	0 ^b	37
		LSV2C-006	NA	0.0	---	---	---
		LSV2C-007	NA	0.0	---	0	0
		LSV2C-008	39	88.3	68	86.7	86.7
		LSV2C-010	42	87.0	88	0 ^b	25
LSV2C-012		42	98.3	73	95.7	95.7	
LSV2C-016		39	95.0	73	91.7	91.7	
LSV2C-017		41	71.3	73	64.0	64.0	
LSV2C-019		38	94.2	73	89.8	89.8	
LSV2C-020	40	89.2	68	88.0	88.0		
LSV2C-021	38	69.3	88	0 ^b	21.7		

^a Percent hatch and percent survival at hatch were synonymous endpoints.

^b Did not achieve swim-up stage before test was terminated; remaining alevins still had yolk sacs (test Day 88).

Percent hatch and percent survival at hatch were synonymous endpoints for all treatments. The percent hatch of the SC treatments was lower than anticipated, ranging from 11.7 – 93.2% (average

of 42%), although this was likely due to the different fertilization technique used for the SC hatchery fish and some fungal problems. The percent hatch for the SPC hatchery fish was 97.5 – 100%, although the holding period was significantly shorter because they were received as eyed eggs. The percent hatch for the field collected eggs was typically better than the SC hatchery eggs. Eggs collected from CC-150 treatments ranged from 14.8 to 97.3%, with an average of 71.9%. Average hatch out for the eggs collected from fish at CC-350 was 56.4%, while that for the eggs from LSV2C was 83.9% (not including LSV2C-006 or -007 whose eggs were either dead upon arrival or were not fertilized). The lowest percent hatch from the LSV2C treatments was 50.6%, while the highest was 99.0%. The field organisms indicated that the change in fertilization technique resulted in a higher hatch success rate compared with the SC hatchery fish.

The day of swim-up for the SC hatchery fish was at 69 days and was 34 days for the SPC hatchery fish (Table 3-3). For the majority of the field treatments, the day of swim-up was between 67 to 73 days, regardless of the collection location. There were five treatments where the alevins did not absorb their yolk sac and reach the swim-up stage. These treatments were: LSV2C-003, LSV2C-004, LSV2C-005, LSV2C-010, and LSV2C-021. These organisms were maintained for the duration of the study and taken down on test day 88 with the last treatments that had finished the 15-d post swim-up portion of the study (Table 3-3).

The next two endpoints were very similar, the percentage of organisms that reached the swim-up stage and percent survival at the swim-up stage (i.e., on the day of swim-up). Because the fry on the day of swim-up had already absorbed their yolk sac, these values were the same for most treatments (Table 3-3). The only treatments for which these values were different were for the five in which the alevins did not reach swim-up (listed above). Survival at this point was determined for the remaining alevins and ranged between 8.0 and 37%.

The last phase of the studies consisted of the 15-d post swim-up study. The first three endpoints for this phase of the study consisted of survival in the 15-d study, total survival for the entire study, and day of test termination. For this phase, each treatment was initiated with a target of 100 of the surviving fry and maintained for 15-d to monitor growth to assess whether there were any latent effects post swim-up. All treatments were initiated with 100 fry per chamber except the following listed below. The number of organisms at initiation of this phase is listed in parenthesis:

- SC-005 (62)
- SPC-002 (20) & SPC-004 (21)
- CC-150-016 (86)
- CC-350-006 (101)
- LSV2C-003 (32), LSV2C-004 (151), LSV2C-005 (111), LSV2C-010 (25), LSV2C-016 (101), & LSV2C-021 (130)

For some of the treatments, i.e., SC-005, CC-150-016, LSV2C-003, LSV2C-004, LSV2C-005, LSV2C-010, and LSV2C-021, only this many fish (or alevins) were left alive at this point in the study. Since some of these were below the target of 100 or had not yet reached swim-up stage, they were

not thinned out, but maintained all the remaining live organisms through the duration of the study. The remaining treatments mentioned were either initiated with fewer eggs (i.e., SPC-002 and SPC-004) or had an extra organism above the target number. For some of the test chambers, fry were lost during this stage of the study due to food clogging the drain pipe, resulting in an overflow of the test chamber. In calculating survival during the 15-d study and overall survival, these organisms were excluded as a technician error. Other technician errors also occurred in the study, for example during thinning when two individual fish were killed (SC-003 and SC-005). A list of the treatments that lost organisms due to an overflow of the chamber and the number lost (i.e., not recovered at the end of the study) are listed below.

- SC-003 (9) & SC-008 (10)
- CC-150-013 (26), CC-150-016 (43), CC-150-017 (33)
- CC-350-007 (20) & CC-350-008 (28)
- LSV2C-002 (16), LSV2C-008 (46), LSV2C-017 (19), LSV2C-019 (39), & LSV2C-020 (36)

Survival during the 15-d post swim-up stage was relatively high (Table 3-4). Excluding the five LSV2C treatments that had poor survival at swim-up (-003, -004, -005, -010, and -021), all other treatments had survival above 96.0%. Survival for the five mentioned LSV2C treatments ranged from 28.1% to 68.5%, indicating that only these treatments had substantial mortality at this phase of the study (Appendix F).

Day of test termination for all but the SPC treatments ranged from 82 to 88 days (Table 3-4). The SC hatchery fish were terminated on day 84, while the SPC fish were terminated on day 49. Total survival for the entire study is presented in Table 3-4. Total survival for the SC hatchery fish ranged from 10.7% to 92.7%, and ranged from 94.2% to 100% for the SPC hatchery fish.

Table 3-4. Percent survival in the 15-d post swim-up phase of the study, total survival for the entire study, and day of test termination for brown trout reproductive study.

Location	Sample ID	Survival (%) in 15-d Post swim-up stage	Total Survival (%)	Survival (%) from Hatch until test term.	Day of test termination
<i>Hatchery Fish</i>	SC-001	97.0	22.7	98.9	84
	SC-002	99.0	22.3	99.5	84
	SC-003	98.9*	54.7	98.0	84
	SC-004	100	27.3	96.2	84
	SC-005	100	10.7	99.0	84
	SC-006	98.0	92.7	99.5	84
	SC-007	96.0	26.3	95.8	84
	SC-008	100*	65.0	99.0	84
<i>Second set of Hatchery Fish</i>	SPC-001	100	98.0	98.7	49
	SPC-002	100	100	100	49
	SPC-003	100	94.2	96.7	49
	SPC-004	100	100	100	49
	SPC-005	96	96.8	98.0	49
	SPC-006	100	96.0	96.3	49
<i>Wild Fish</i>	CC-150-009	99	27.0	98.3	87
	CC-150-011	100	95.3	98.7	82
	CC-150-012	97	86.0	98.1	87
	CC-150-013	97.3*	57.5	93.0	87
	CC-150-015	98	77.5	98.9	82
	CC-150-016	100*	15.4	99.7	87
	CC-150-017	100*	84.3	97.2	82
	CC-150-018	100	84.3	96.5	87
	CC-150-020	100	96.3	98.9	82
	CC-350-006	98.0	67.7	96.3	87
	CC-350-007	98.8*	26.0	98.6	82
	CC-350-008	98.6*	62.6	97.1	82
	LSV2C-002	100*	96.6	97.6	83
	LSV2C-003	28.1	2.25	8.8	88
	LSV2C-004	55.6	16.8	66.2	88
	LSV2C-005	62.2	23.0	51.7	88
	LSV2C-008	98.2*	84.2	95.9	83
	LSV2C-010	44.0	11.0	24.0	88
	LSV2C-012	100	95.7	97.4	88
	LSV2C-016	100	91.7	96.7	88
	LSV2C-017	96.3*	60.5	89.2	88
LSV2C-019	100*	88.9	94.7	88	
LSV2C-020	100*	86.8	97.6	83	
LSV2C-021	68.5	14.8	45.5	88	

* - missing organisms at test termination were not included in calculation.

The range of total survival for the CC-150 treatments was 15.4% to 96.3%, that for the CC-150 treatments was 26.0% to 67.7%, and for the LSV2C treatments was 2.25% to 96.6% (Table 3-4).

For most of these treatments, there was not a substantial difference between the survival rate at swim-up and the number of organisms that hatched (Figure 3-1). For most of the treatments, the number of organisms that hatched reached the swim-up stage. One group did not hatch or swim-up

(LSV2C-007), while five groups had substantial hatch (>50%) but did not reach swim-up (as mentioned previously). Percent survival, from hatch until test termination was included in Table 3-4.

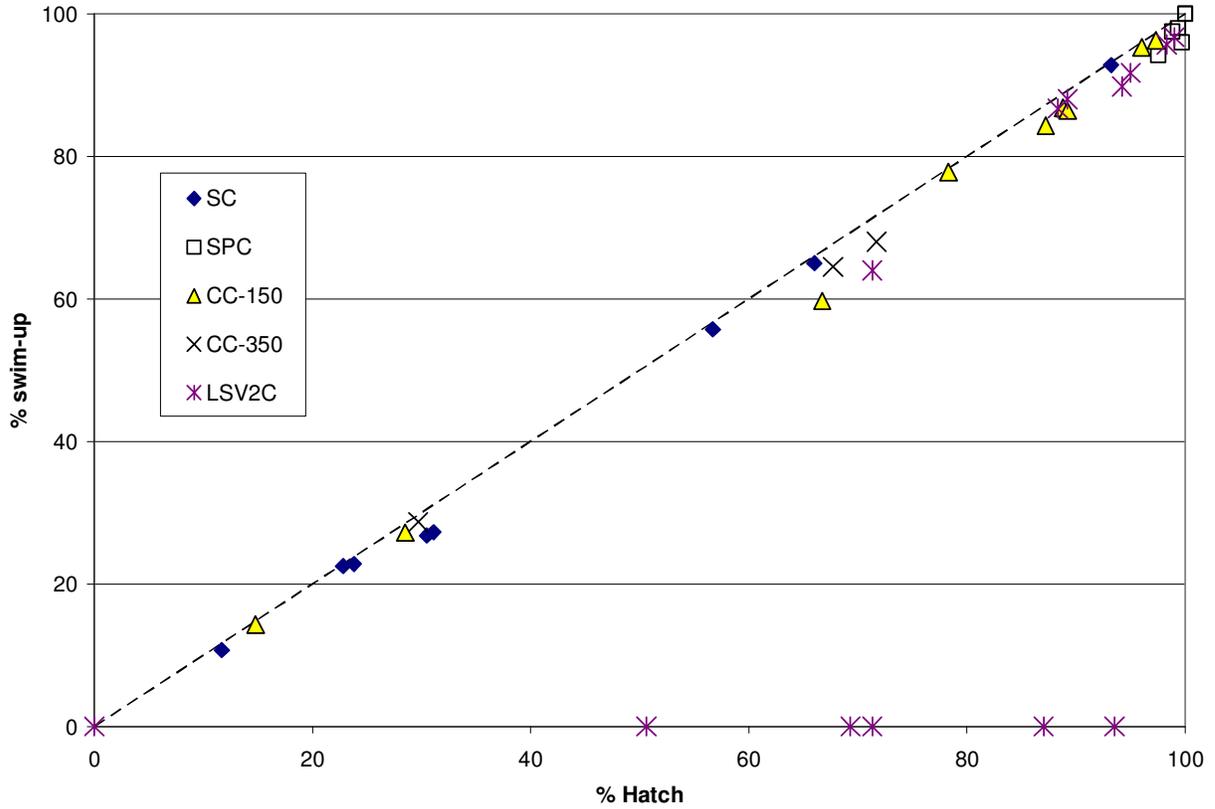


Figure 3-1. Relationship between the percentage of organisms that hatched and the percentage of organisms that reached swim-up. Note, the dashed line indicates a 1:1 agreement.

The results of length and dry weight analysis for the target of 20 organisms at the end of the 15-d post swim-up phase of the study are provided below (Table 3-5). Raw data are in Appendix G.

Table 3-5. Standard length and dry weight (avg \pm SD) of larval brown trout at test termination. The number of larval fish measured for each treatment is included (n).

Location	Sample ID	n	Average Standard Length (mm)	Average Dry Weight (mg)	
<i>Hatchery Fish</i>	SC-001	20	21.4 \pm 1.5	18.338 \pm 4.3	
	SC-002	20	22.2 \pm 0.93	18.470 \pm 3.3	
	SC-003	20	22.8 \pm 1.3	22.602 \pm 4.8	
	SC-004	20	22.2 \pm 1.9	23.246 \pm 7.5	
	SC-005	20	21.8 \pm 1.2	19.386 \pm 4.0	
	SC-006	20	21.8 \pm 0.83	18.240 \pm 3.3	
	SC-007	20	21.0 \pm 1.5	18.567 \pm 3.9	
	SC-008	20	21.2 \pm 1.1	15.118 \pm 3.3	
<i>Second set of Hatchery Fish</i>	SPC-001	20	24.4 \pm 2.1	22.564 \pm 6.1	
	SPC-002	20	22.8 \pm 1.5	20.852 \pm 4.1	
	SPC-003	20	23.3 \pm 2.1	21.807 \pm 5.4	
	SPC-004	20	23.4 \pm 2.0	21.564 \pm 5.3	
	SPC-005	20	23.0 \pm 1.9	20.792 \pm 5.8	
	SPC-006	20	23.2 \pm 1.6	22.386 \pm 6.1	
Wild	CC-150-009	20	21.8 \pm 1.3	13.188 \pm 3.2	
	CC-150-011	20	20.1 \pm 0.85	9.248 \pm 2.3	
	CC-150-012	20	21.4 \pm 0.89	12.840 \pm 2.8	
	CC-150-013	20	22.6 \pm 1.3	15.582 \pm 3.4	
	CC-150-015	20	22.6 \pm 1.2	14.486 \pm 3.2	
	CC-150-016	20	22.8 \pm 1.2	16.736 \pm 4.0	
	CC-150-017	20	21.0 \pm 0.83	10.652 \pm 2.4	
	CC-150-018	20	21.3 \pm 1.0	13.244 \pm 3.3	
	CC-150-020	20	20.2 \pm 1.4	11.248 \pm 3.6	
	CC-350-006	20	21.4 \pm 1.1	12.342 \pm 3.1	
	CC-350-007	20	21.3 \pm 1.3	11.009 \pm 3.7	
	CC-350-008	20	20.6 \pm 0.94	12.089 \pm 3.1	
	Fish	LSV2C-002	20	20.1 \pm 1.2	11.070 \pm 3.2
		LSV2C-003	9	21.3 \pm 0.71	9.710 \pm 2.4
LSV2C-004		20	21.2 \pm 0.89	9.979 \pm 2.1	
LSV2C-005		20	20.2 \pm 1.1	10.967 \pm 2.7	
LSV2C-008		20	20.4 \pm 0.51	9.342 \pm 1.1	
LSV2C-010		11	19.9 \pm 0.83	8.092 \pm 2.5	
LSV2C-012		20	22.0 \pm 0.46	13.256 \pm 0.8	
LSV2C-016		20	21.8 \pm 1.5	15.836 \pm 5.0	
LSV2C-017		20	23.7 \pm 1.0	18.878 \pm 4.0	
LSV2C-019		20	23.6 \pm 1.4	19.320 \pm 5.8	
LSV2C-020	20	21.8 \pm 1.0	11.574 \pm 2.7		
LSV2C-021	20	20.2 \pm 1.5	10.656 \pm 3.9		

3.2.3 Deformity Assessment

Below is a list of the number of specimens preserved and analyzed at either swim-up or test termination for deformities (Table 3-6). The majority of fish that had died during the test were preserved but were not evaluated because of the poor state that they were in by the time death had occurred. As mentioned, a subset of these dead organisms were evaluated for deformities and included with the results of the assessment conducted on organisms that were alive when preserved.

Table 3-6. Number of brown trout fry preserved and assessed for deformities. Samples were preserved at swim-up, at test termination, or upon death. All organisms preserved at swim-up and test termination were assessed for deformities; however, only select samples from organisms that died during the study were evaluated.

Location	Field Sample ID	Number of fish assessed that were preserved at swim-up	Number of fish assessed that were preserved at test termination	Number of fish assessed that had died during the study
<i>Hatchery Fish</i>	SC-001	38	77	--
	SC-002	34	79	--
	SC-003	233	69	--
	SC-004	60	80	--
	SC-005	0	42	--
	SC-006	457	78	--
	SC-007	61	76	--
	SC-008	289	70	--
<i>Second set of Hatchery Fish</i>	SPC-001	488	80	--
	SPC-002	0	0	--
	SPC-003	465	80	--
	SPC-004	0	0	--
	SPC-005	485	76	--
	SPC-006	476	80	--
Wild	CC-150-009	62	80	--
	CC-150-011	185	81	--
	CC-150-012	204	77	--
	CC-150-013	258	52	--
	CC-150-015	367	78	--
	CC-150-016	0	23	--
	CC-150-017	116	47	--
	CC-150-018	405	81	--
	CC-150-020	478	80	--
	CC-350-006	307	79	--
	CC-350-007	72	59	--
	CC-350-008	287	51	--
	Fish	LSV2C-002	480	64
LSV2C-003		0	0	~100 ^a
LSV2C-004		0	64	~100 ^a
LSV2C-005		0	49	~100 ^a
LSV2C-008		160	34	--
LSV2C-010		0	0	~84 ^a
LSV2C-012		474	80	--
LSV2C-016		449	81	--
LSV2C-017		92	58	--
LSV2C-019		349	41	--
LSV2C-020		252	44	--
LSV2C-021	0	69	~100	

^aWhile a subset of ~100 organisms were evaluated, scoring criteria were not always possible due to the poor physical state at preservation.

For this assessment, the scoring criteria results of the fry preserved at swim-up, the fry preserved at test termination, and the fry preserved upon death (select samples) were combined. A summary of the raw data can be found in Appendix D of the main Brown Trout report.

4.0 References

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Appendix A

Select photographs of different phases of the brown trout reproductive study

Photo 1: Egg cups used for hatching of brown trout eggs; 10 replicates with 60 (target) eggs / replicate.



Photo 2: Egg cup with eggs at test initiation. Photo includes egg pickers, container of remaining eggs for analytical, and counter.

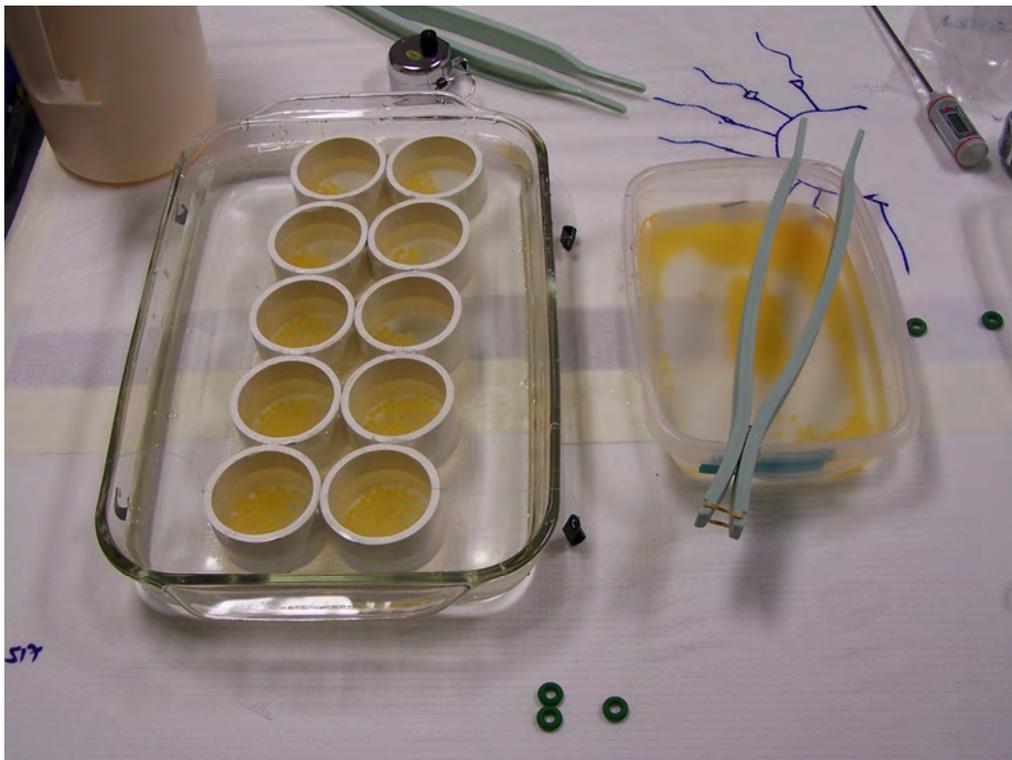


Photo 3: Eggs in egg cups for CC-350-008 brown trout eggs; 10 replicates with 60 eggs / replicate.



Photo 4: Close up of eggs in egg cup at test initiation.

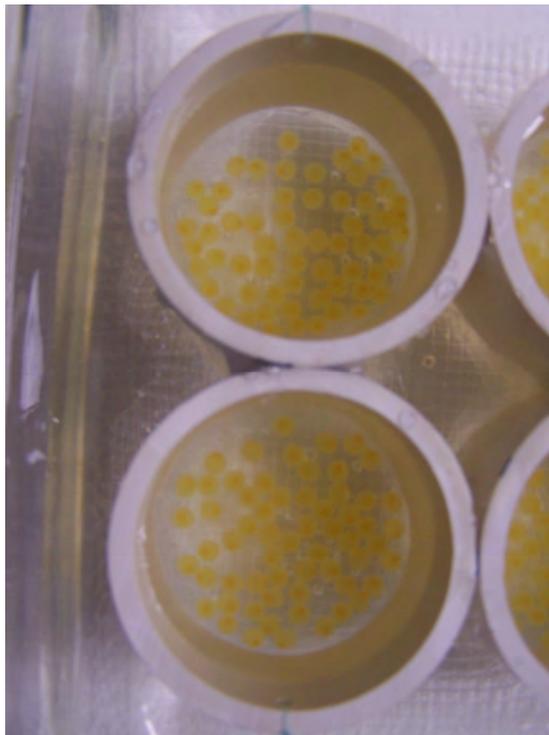


Photo 5: Water bath covered with black curtain to keep brown trout eggs in the dark during initial stage.



Photo 6: Test chambers in water bath for brown trout reproductive study during swim-up stage.

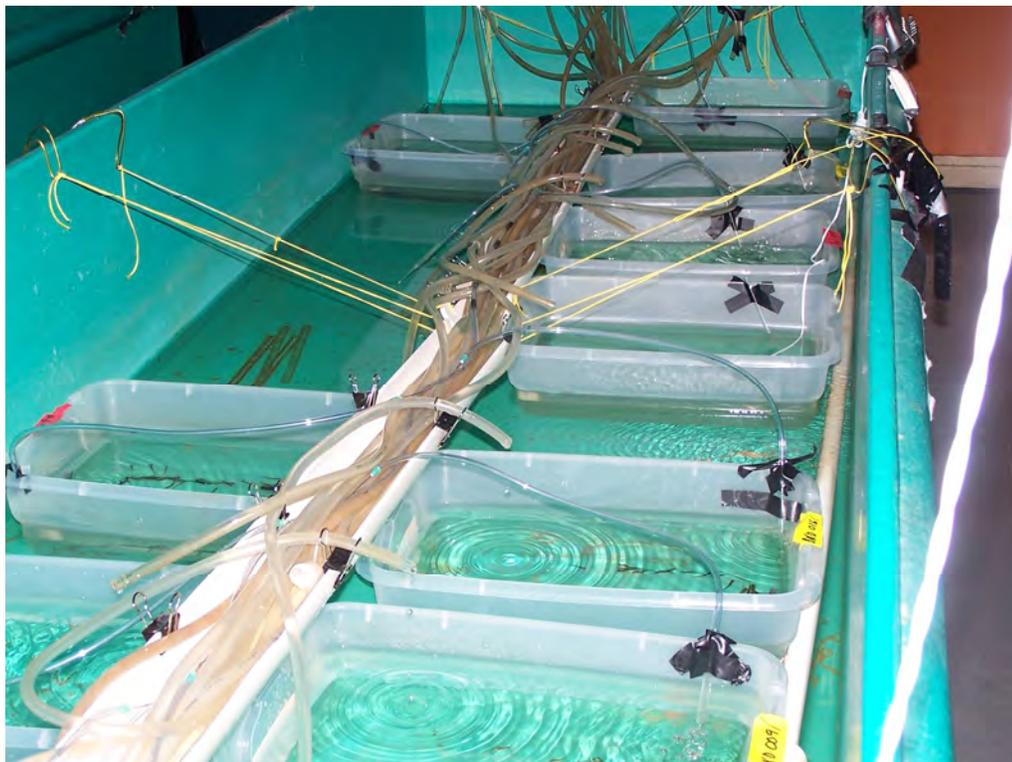


Photo 7: Dilutor panel (one of two) used to feed adjusted Horsetooth water to each testing chamber.



Photo 8: Separation of water bath into light conditions for swimup fry and dark for pre swimup yolk sac fry.



Photo 9: Brown trout fry in test chamber at swim-up (prior to thinning).



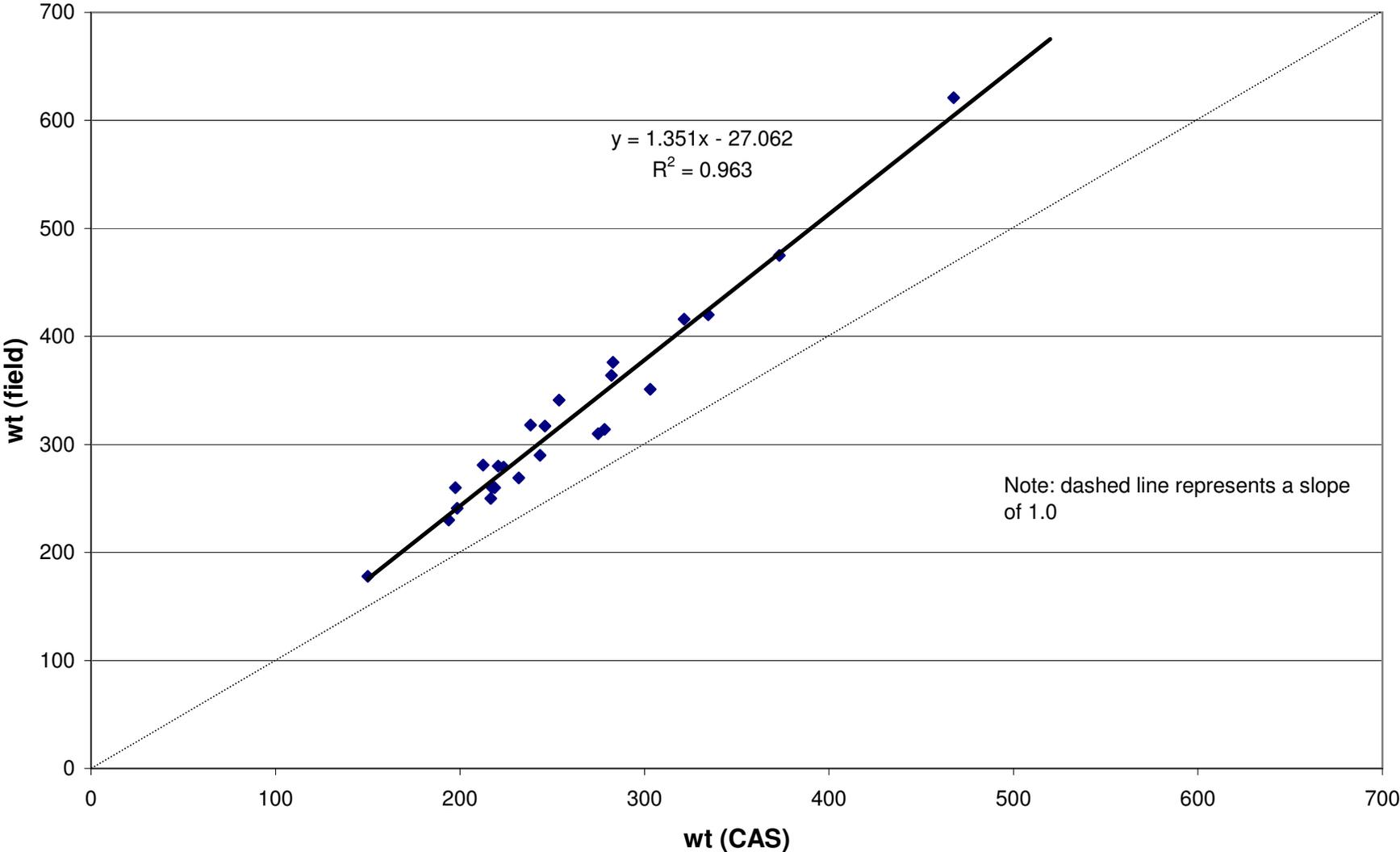
Photo 10: Brown trout fry in test chamber at swim-up (after thinning to 100 organisms).



Appendix B

Wet weight estimation for adult fish from Saratoga hatchery and raw data for adult fish

Relationship between wet wt data of field fish from CAS and from field measurements.



Appendix C

Egg counts for Saratoga hatchery fish (SC) and field collected fish

**Estimation of Brown trout eggs numeration using volume-estimating technique, 12699-001
 October 25, 2007
 Hatchery Eggs from Saratoga National Fish Hatchery (SNFH, Saratoga, WY)**

The number of eggs placed into each study was counted manually. For all of the treatments, the remaining number of eggs was estimated using a volume technique to develop a #egg/volume ratio for that particular female. The technique consisted of counting the number of eggs that filled a graduated cylinder to a particular volume (e.g., 50 mL) to determine the # of eggs per mL for that female. Initially we conducted two separate counts by two different staff personnel. Based on the agreement of these counted numbers for the particular volume, we subsequently only conducted counts once per female. Using this ratio, we then calculated the total number of remaining eggs for the total volume of eggs measured in a graduated cylinder. The total number of eggs used to initiate the studies (i.e., 600) was then added to the estimated number of remaining eggs to determine the total number of eggs for that particular female trout.

<u>Treatment</u>	<u># Eggs placed in study</u>	<u>Count#</u>	<u># eggs counted to est. #eggs/vol</u>	<u>Avg # eggs counted</u>	<u>Vol of eggs counted (ml)</u>	<u>Avg # eggs/mL</u>	<u>Total vol (ml) of remaining eggs</u>	<u>Total # eggs</u>
SC-001	600	1 2	1058 1044	1051	100 100	10.51	340	4,173
SC-002	600	1 2	570 565	567.5	50 50	11.35	300	4,005
SC-003	600	1	452	452	50	9.04	500	5,120
SC-004	600	1	506	506	50	10.12	64	1,248
SC-005	600	1	480	480	50	9.6	505	5,448
SC-006	600	1	477	477	50	9.54	270	3,176
SC-007	600	1	495	495	50	9.9	265	3,224
SC-008	600	1	532	532	50	10.64	320	4,005
							Avg	3,800
							Geomean	3,525
							SD	1,305
							CV	34%

**Estimation of Brown trout eggs numeration using direct counts and volume-estimation technique, 12699-001
November 16 & 17, 2007**

Treatment eggs from background, low, and high treatment areas (ID)

The number of eggs placed into each study was counted manually. For almost all of the treatments, the remaining eggs were also counted manually. When there were too many eggs to count, the number of remaining eggs were estimated using a volume technique to develop a #eggs/volume ratio for that particular female. The technique consisted of counting the number of eggs that fill a graduated cylinder to a particular volume (e.g., 50 mL) to determine the # eggs per mL for that female. Using this ratio, we then calculated the total number of remaining eggs for the total volume of eggs measured in the graduated cylinder. The total number of eggs used to initiate the studies (e.g., 600) was then added to this estimated number of remaining eggs to determine the total number of eggs for that particular female trout.

<u>Treatment</u>	<u># Eggs placed in study</u>	<u># Remaining eggs counted</u>	<u># eggs counted to est. #eggs/vol</u>	<u>Vol of eggs counted (ml)</u>	<u>Avg # eggs/mL</u>	<u>Total vol (ml) of remaining eggs</u>	<u>Total # eggs</u>	
High								
LSV2C-002	600	496					1,096	
LSV2C-003	400	74					474	
LSV2C-004	500	266					766	
LSV2C-005	300	176					476	
LSV2C-006	eggs dead / dying upon arrival (not counted)							
LSV2C-007	500	273					773	
LSV2C-008	300	72					372	
LSV2C-010	100	61					161	
LSV2C-012	600	431					1,031	
LSV2C-016	600	226					826	
LSV2C-017	300	147					447	
LSV2C-019	500	193					693	
LSV2C-020	400	125					525	Avg
LSV2C-021	600	608					1,208	681
Bkd								
CC-150-009	600	615					1,215	
CC-150-011	300	188					488	
CC-150-012	350	206					556	
CC-150-013	600	634					1,234	
CC-150-015	600	403					1,003	
CC-150-016	600		460	50	9.2	115	1,658	
CC-150-017	250	164					414	
CC-150-018	600	359					959	Avg
CC-150-020	600	732					1,332	984
Low								
CC-350-006	600	554					1,154	
CC-350-007	600	574					1,174	Avg
CC-350-008	600	322					922	1,083

10/24/07

BT Fertilization

SUBJECT: DAILY LOG

ALL ENTRIES MUST BE INITIALED WITH DATE AND TIME:

- Checked temp in cooler in am
 still OK - 6-7°C

Internal

- Temp range of the cooler
 from Saratoga until Wed am
 was 6.1 - 9.7°C
 (9.7°C was initial reading taken on
 10/30 before thermometer had equilibrated
 in cooler)

gametes obtained on 10/23/08

Treatments

SC-001

-002

-003

-004

-005

-006

-007

-008

General
 Fertilization process

1) - place eggs in plastic container
 - estimate vol, using surrogate
 container w/ H₂O (similar levels
 (Rubbermaid 32oz plastic container))

2) Add semen (we composited
 semen from 6 of the 8 males
 too much yellow color in those 2)
 2 ml of semen to each batch
 of eggs, spread around
 container

3) mix by gently swirling
 container

4) sit for ~ 60 sec (sometimes this
 time includes mixing)

5) added 100 ml of 6°C HT modified
 water (from PBT tank)
 i.e. enough to cover eggs

6) gently swirl container (w/ eggs/semen)
 for 3 min

7) add ~ 500 ml more of 6°C
 HT modified water to water
 harden

(still in tupperware)
 8) placed in cooler w/ ice in
 bottom to slowly acclimate
 eggs to ~ 10°C (over piece of ice
 will slowly melt & temp will rise)

9) Added eggs to Egg cups
 10/25 & placed in water bath

Egg estimation technique

10/25/07

SUBJECT: DAILY LOG

ALL ENTRIES MUST BE INITIALED WITH DATE AND TIME:

#1 100 mL of eggs = 1058 eggs (340 mL)

#2 50 mL of eggs = 520 eggs (300 mL)
1044 eggs
565 eggs

#3 50 mL of eggs = 452 eggs (500 mL)

#4 50 mL of eggs = 506 (64 mL) eggs more yellow than
orange (others more orange)

#5 50 mL of eggs = 480 eggs (505 mL)

#6 50 mL of eggs = 477 (270 mL)

#7 50 mL of eggs = 495 (265 mL)

#8 50 mL of eggs = 532 (320 mL)

Results of egg estimation technique

Tried to count using an egg counter (Marisource, Inc) but too large & eggs too large - doubled up in holes. Came up with this technique.

Measurement of Adult SC trout

SUBJECT: DAILY LOG

ALL ENTRIES MUST BE INITIALED WITH DATE AND TIME:

10/25/07@1600

6mm @ TD

females

	C.p.	total length *
SC-001	428 mm	498 mm
SC-002	360 mm	420 mm
SC-003	440 mm	520 mm
SC-004	470 mm	562 mm
SC-005	475 mm	558 mm
SC-006	365 mm	439 mm
SC-007	385 mm	449 mm
SC-008	411 mm	494 mm
SC-001 male with 006	238 mm	290 mm

length measurements for adult Brown trout
 used in reproductive study

C.p. = caudal peduncle

11/16/07

SUBJECT: DAILY LOG

ALL ENTRIES MUST BE INITIALED WITH DATE AND TIME:

1315 start placing fert eggs in ϕ egg cups 60/cup \rightarrow target
 1335 & 1415 = 600 fert

measured DO in LSU2C-002 \approx 10.0 mg/L

- raise off eggs w/ modified HT, transfer w/ egg pickers

eggs in egg cups

Container	Count	Notes	Target
LSU2C-002	60 eggs each	rest to count in cooler	600
-003	40 eggs each	"	400
-004	50 eggs each	"	500
-005	30 eggs each	"	300
-006	eggs turned white	freeze for tissue analysis	
-007	50 eggs each		500
-008	30 eggs each		300
yellowish - 010	10 eggs each		100
yellowish - 012 (eggs looked larger)	60 eggs each		600
greyish - 016	60 eggs each		600
-017	30 eggs each		300
-019 (eggs looked larger)	50 eggs each		500
yellowish - 020	40 eggs each		400
yellowish 021	60 eggs each		600

start small bags

(161 total)

end e 1800 hrs

still need to count remaining eggs

min/max temp in cooler 6.2-8.4 during overnight equilibration

① 1335 11/16/07 E. ② 1335 10/23/08 CF; # of organisms from fert indicate 600 organisms were counted at initiation

11/17/07 cooler #2 for ID

SUBJECT: DAILY LOG

ALL ENTRIES MUST BE INITIALED WITH DATE AND TIME:

eggs collect 11/15

1038 start placing fert eggs in egg cups

cooler min-max during equilibration period 7.9-8.9°C
 all in smaller bags

ID

CC-350-006	60 eggs each	600 eggs	CC-350-006
↓ -007	smaller eggs	60 eggs each	↓ -007
↓ -008	larger eggs	60 eggs each	↓ -008
CC-150-009	smaller yellowish	60 eggs each	CC-150-009
↓ -011	few yellowish	30 eggs each	↓ -011
↓ -012	yellowish	35 eggs each	↓ -012
↓ -013	smaller yellowish	60 eggs each	↓ -013
↓ -015	"	60 eggs each	↓ -015
↓ -016	larger brownish	60 eggs each	↓ -016
↓ -017	few yellowish	25 eggs each	↓ -017
↓ -018		60 eggs each	↓ -018
↓ -020		60 eggs each	↓ -020

low site
↓

BKD site
↓

finished @ 1545

collected
 11/15/07
 @ 1605
 ↓

11/15/07
 1300
 ↓

Appendix D

Summary of fungal treatment methods for SC Hatchery eggs and field collected eggs

Treatment of brown trout eggs for fungal control. Study # 12699-001

Different methods were employed in order to help control fungal growth and improve overall success rate of the brown trout eggs in the reproductive study. As mentioned in the report, this primarily affected only the SC hatchery eggs because they were the first batch to get in. And additional methods were employed to reduce fungal growth with the field collected eggs (initiated Nov. 16 & 17, 2008) and the second set of hatchery eggs (initiated Dec. 4, 2007).

November 2, 2007:

1% salt (as NaCl) solution was added to SC-002 and SC-004 chambers (i.e., 50 g of NaCl was mixed with small amount of water from each test chamber until all salt was dissolved and added). Measured chloride concentration at 5,620 mg/L. Time = 1700 hrs.

Conductivity check at 1920 hrs:

SC-004 @ 12,800 $\mu\text{S}/\text{cm}^2$

SC-002 @ 11,440 $\mu\text{S}/\text{cm}^2$

Background conductivity @ 123 $\mu\text{S}/\text{cm}^2$

November 3, 2007:

Conductivity check in chambers @ 0550 hrs

SC-004 @ 1,770 $\mu\text{S}/\text{cm}^2$

SC-002 @ 2,670 $\mu\text{S}/\text{cm}^2$

Background conductivity @ 118 $\mu\text{S}/\text{cm}^2$

1% salt (as NaCl) solution was added to SC-003 (@0630 hrs) and SC-001 (@0700 hrs) chambers (i.e., 50 g of NaCl was mixed with small amount of water from each test chamber until all salt was dissolved and added).

Conductivity check in chambers @ 1620 hrs

SC-004 @ 343 $\mu\text{S}/\text{cm}^2$

SC-002 @ 602 $\mu\text{S}/\text{cm}^2$

SC-001 @ 3,660 $\mu\text{S}/\text{cm}^2$

SC-003 @ 3,830 $\mu\text{S}/\text{cm}^2$

November 6, 2007:

Installed small UV disinfection system on HT water line prior to going into 5-gallon head tank bucket above dilutor panel. This treats all water going into the study. Because there was algal / fungal growth in 5-gallon head tank, treated head tank with NaCl (189 g mixed in 5 gallons). Salt solution was allowed to flush through the dilutor and into the test chambers (SC test chambers only).

November 9, 2007:

Added 50 g of NaCl to each SC chamber because still seeing fungus (*Saprolegnia?*).

November 12, 2007:

Now that eggs were eyed up, salt treatment consisted of removing egg cups from test chamber (SC treatments) and soaking in 3% salt treatment for 1 hour, and then transferring

back to cleaned test chamber. Treated drip lines with (~1,700 ppm) formalin. The formalin solution did not go into test chambers with eggs.

November 16, 2007:

A 1.5% NaCl solution (75 g NaCl) was added to 5 L water in test container for SC hatchery eggs.

November 24, 2007:

Saw some dead eggs in newly received field egg treatments. Add 190 g of NaCl to 5 gal head tank. Let it flush through dilutor and into all chambers.

November 25, 2007:

Measured conductivity @ 0800 hrs, ~200 $\mu\text{S}/\text{cm}^2$. Moved all SC hatchery eggs into new egg cups. Let soak in 3% NaCl solution (60 g / 2 L) for ~ 1 h while being transferred into new egg cups. Some eggs were very covered by fungus. SC005 treatment appeared to have a lot of unfertilized (non-eyed up) eggs. Cleaned (soap and water wash) SC test chambers while eggs were in NaCl soak (all SC treatments).

Treated all test chambers in bath #2 (ENSR water bath #3) with 1.5% NaCl, except SC chambers. Mixed 75 g NaCl with deionized water (Milli-Q) and poured into each chamber. 15 test chambers in all for bath #2.

Consisted of: CC-350-(LOW)-007
 CC-150-(BKD)-009, -012, -015, & -016
 LSV2C-(HIGH)-004, -005, -007, -008, -017, -019, & -020
 SPC-(2ND HATCHERY)-002, -003, & -006

November 26, 2007:

Treated all test chambers in bath #1 (ENSR water bath #4) with 1.5% NaCl, except SC chambers. Mixed 75 g NaCl with deionized water (Milli-Q) and poured into each chamber. 16 test chambers in all for bath #1.

Consisted of: CC-350-(LOW)-006 & -008
 CC-150-(BKD)-011, -013, -017, -018, & -020
 LSV2C-(HIGH)-002, -003, -010, -012, -016, & -021
 SPC-(2ND HATCHERY)-001, -004, & -005

Drained and cleaned large circular (trout tank) hardness mixing tank. Chambers were static for ~2 hours.

November 28, 2007:

Soaked SC treatments by placing egg cups in 3% NaCl for one hour (120 g / 4 L) in separate chamber. Removed and placed back into test chambers. Cleaned as necessary.

November 29, 2007:

After discussions with S. Covington and hatchery personnel, decided to use formalin to help minimize fungal growth on SC eggs. Treated SC-001 eggs with formalin (~1700 ppm, 6.8 ml in 4 L), by placing egg cup in separate chamber with formalin (next to original test chamber in water bath). From 1054 hrs to 1115 hrs.

Also treated with following test chambers with formalin in a similar way.

SC-002 (1124 hrs to 1140 hrs)
 SC-003 (1142 hrs to 1157 hrs)
 LSV2C-(HIGH)-007 (1720 hrs to 1738 hrs).

Noticed a lot of dead / dying eggs in the LSV2C-(HIGH)-007 treatment at the end of the day so decided to treat with formalin, even though it did not appear that there was fungus on these eggs¹.

November 30, 2007:

Treated the following egg treatments with formalin (~1700 ppm, as 7 ml in 4 L). Performed in secondary chamber as before, soaked for ~30 min, and returned to original chamber. Started soak at 1520 hrs, finish at 1550 hrs.

SC-001, SC-005, SC-007, SC-008 (bath #1)
 SC-004, SC-002, SC-003, SC-006 (bath #2)

December 1, 2007:

Placed egg cup for SC-001 in old formalin solution (prepared on Nov. 30, but still in water bath) for 15 minutes. Returned to original test chamber.

December 2, 2007:

Treat SC-001 with formalin by mixing 8.5 ml in 200 ml water (i.e., cold hardness adjusted Horsetooth water, similar as all preps before). Poured this mixture directly into test chamber (assumes ~5L so target is ~1700 ppm). After 15 min, pur 4 L HT water into test chamber to flush out formalin.

Still seeing fungus on control eggs, try a 4% NaCl solution on 1 SC batch for 1 hour. Prepared by dissolving 200 g NaCl in 1 L of water and pouring directly into test chamber (assumes 5 L of water in chamber). Added to SC-005 @ 1340 hrs.

Treated the following test chambers with formalin (8.5 ml into test chamber for 15 minutes, followed by 4-5L flush): SC-004, SC-002, SC-006

December 3, 2007:

Observed first hatched out alevin in SC-004 and 1 egg that appeared to explode (termed 'dead while hatching').

¹ None of these eggs appeared to be fertilized and all eventually died.

No more treatments for SC test chambers.

December 5, 2007:

Added 1% NaCl (50 g NaCl, assuming 5 L water in each chamber) to all test chambers except SC treatments (-001 through -008), and SPC-002, and SPC-004 (small SPC test chambers, i.e., ~ 2.5 L chambers).

December 7, 2007:

Clean out test chambers for LSV2C-008 and -019 with soap and water and replaced in bath.

December 8, 2007:

Cleaned egg cups by brushing outside surface with toothbrush. Did not disturb eggs. After cleaning egg cup, placed egg cup into a cleaned test chamber. This was done for the following treatments.

LSV2C-(HIGH)-017, -020, -007 (transferred few remaining eggs to a new egg cup),
-005, -004, -010, -002, & -016

December 9, 2007:

Cleaned egg cups by brushing outside surface with toothbrush. Did not disturb eggs. After cleaning egg cup, placed egg cup into a cleaned test chamber. Also replaced aeration pipettes. This was done for the following treatments.

LSV2C-(HIGH)-021, -012, & -003

December 10, 2007:

Cleaned egg cups by brushing outside surface with toothbrush. Did not disturb eggs. After cleaning egg cup, placed egg cup into a cleaned test chamber. Also replaced aeration pipettes. This was done for the following treatments.

CC-350-(LOW)-006, -007, & -008
CC-150-(BKD)-009, -015, & -016

December 7 – 18, 2007:

Noticed that flows on some treatments were reduced due to fouling of tubing lines (from splitter box to test chamber). Flushed out lines with hydrogen peroxide (1.5%). Test chambers were static during the flush out and either overnight or during the day (~ 8 hr) while water was allowed to flush through the tubing to waste. Also blew biofilm out of some lines, but this did not work for all chambers. This was done over a period of time because there were only a few extra tubing lines that did not directly feed a treatment. Therefore, a few were cleaned each day and allowed to flush prior to using for a test chamber.

December 31, 2007:

Egg cup for LSV2C-(HIGH)-019 was very dirty so changed out with clean egg cup.

January 1, 2008:

Egg cups for the following treatments were very dirty so changed out with clean egg cups:

LSV2C-(HIGH)-021, -004 & -012
CC-150-(BKD)-013 & -012
CC-350-(LOW)-006

January 2, 2008:

Cleaned dilutor panels (scrubbing only) to remove biofilm buildup. Test chambers were static for ~ 1 hr during this time.

January 18, 2008:

Exchanged test chamber for LSV2C-(HIGH)-003 with clean one due to fungal growth and high mortality.

Appendix E

Summary of water quality data selenium and sulfate analysis in water and selenium concentrations in eggs measure during reproductive study

ENSRProject ID: 12699-001
Sample ID: SPC-TOTACZ Sample ID: **L72820-01**
Date Sampled: 10/30/08 15:10
Date Received: 11/04/08
Sample Matrix: Ground Water

Inorganic Prep

Parameter	EPA Method	Result	Qual	XO	Units	MDL	PQL	Date	Analyst
Total Recoverable Digestion	M200.2 ICP-MS							11/07/08 11:28	jws

Metals Analysis

Parameter	EPA Method	Result	Qual	XO	Units	MDL	PQL	Date	Analyst
Selenium, total recoverable	M200.8 ICP-MS		U		mg/L	0.0001	0.0005	11/09/08 8:44	erf

ENSRProject ID: 12699-001
Sample ID: SPC-DISSACZ Sample ID: **L72820-02**
Date Sampled: 10/30/08 15:10
Date Received: 11/04/08
Sample Matrix: Ground Water

Metals Analysis

Parameter	EPA Method	Result	Qual	XQ	Units	MDL	PQL	Date	Analyst
Selenium, dissolved	M200.8 ICP-MS		U		mg/L	0.0001	0.0005	11/12/08 2:33	erf

Total Recoverable SELENIUM

Method SW6010B

Sample Results

Lab Name: Paragon Analytics
Client Name: ENSR Consulting and Engineering
Client Project ID: Brown Trout 12698-001
Work Order Number: 0711270 Final Volume: 50 g
Reporting Basis: As Received Matrix: WATER
Prep Method: SW3005A Result Units: mg/l

Client Sample ID	Lab ID	Date Collected	Date Prepared	Date Analyzed	Percent Moisture	Dilution Factor	Result	Reporting Limit	Flag	Sample Allquot
HT + metals	0711270-2	10/26/2007	12/04/2007	12/05/2007	N/A	1	0.005	0.005	U	50 g
Modified HT	0711270-4	11/17/2007	12/04/2007	12/05/2007	N/A	1	0.005	0.005	U	50 g
HT + metals	0711270-7	11/29/2007	12/04/2007	12/05/2007	N/A	1	0.005	0.005	U	50 g

Comments:

1. ND or U = Not Detected at or above the client requested detection limit.

Data Package ID: IT0711270-1

Date Printed: Tuesday, December 11, 2007

Paragon Analytics
LIMS Version: 6.095A

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Total Recoverable ICP Metals

Method SW6010B

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0712111

Client Name: ENSR Consulting and Engineering

Client Project ID: Brown Trout 12699-001

Field ID: HT+ metals
Lab ID: 0712111-2

Sample Matrix: WATER
% Moisture: N/A
Date Collected: 13-Dec-07
Date Extracted: 17-Dec-07
Date Analyzed: 16-Dec-07
Prep Method: SW3005 Rev A

Prep Batch: IP071217-2
QCBatchID: IP071217-2-1
Run ID: IT071218-2A1
Cleanup: NONE
Basis: As Received

Sample Allquot: 50 g
Final Volume: 50 g
Result Units: mg/l
Clean DF: 1
File Name: 071218A.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
7762-49-2	SELENIUM	1	0.005	0.005	U	

Data Package ID: IT0712111-1

Date Printed: Friday, December 21, 2007

Paragon Analytics
LIMS Version: 6.097A

Page 1 of 1

Total Recoverable ICP Metals

Method SW6010B

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0801009

Client Name: ENSR Consulting and Engineering

ClientProject ID: Brown Trout 12699-001

Field ID: HT + metals

Lab ID: 0801009-2

Sample Matrix: WATER

% Moisture: N/A

Date Collected: 03-Jan-08

Date Extracted: 04-Jan-08

Date Analyzed: 07-Jan-08

Prep Method: SW3005 Rev A

Prep Batch: IP080104-3

QCBatchID: IP080104-3-1

Run ID: IT080107-2A1

Cleanup: NONE

Basis: As Received

Sample Aliquot: 50 g

Final Volume: 50 g

Result Units: mg/l

Clean DF: 1

File Name: 080107A.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
7782-49-2	SELENIUM	1	0.005	0.005	U	

Data Package ID: IT0801009-1

Date Printed: Friday, January 11, 2008

Paragon Analytics

LIMS Version: 6.105A

Page 1 of 1

Total Recoverable ICP Metals

Method SW6010B

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0801127

Client Name: ENSR Consulting and Engineering

Client Project ID: Brown Trout 12699-001

Field ID:	HT
Lab ID:	0801127-1

Sample Matrix: WATER

% Moisture: N/A

Date Collected: 16-Jan-08

Date Extracted: 17-Jan-08

Date Analyzed: 18-Jan-08

Prep Method: SW3005 Rev A

Prep Batch: IP080117-1

QC Batch ID: IP080117-1-1

Run ID: IT080118-2A1

Cleanup: NONE

Basis: As Received

Sample Allquot: 50 g

Final Volume: 50 g

Result Units: mg/l

Clean DF: 1

File Name: 080118A.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
7782-49-2	SELENIUM	1	0.005	0.005	U	

Data Package ID: IT0801127-1

Dissolved ICP Metals

Method SW6010B

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0801254

Client Name: ENSR Consulting and Engineering

ClientProject ID: Brown Trout 12699-001

Field ID: SPC control 001
Lab ID: 0801254-1

Sample Matrix: WATER
% Moisture: N/A
Date Collected: 22-Jan-08
Date Extracted: 31-Jan-08
Date Analyzed: 31-Jan-08
Prep Method: SW3005 Rev A

Prep Batch: IP080131-10
QCBatchID: IP080131-10-1
Run ID: IT080131-2A1
Cleanup: NONE
Basis: As Received

Sample Aliquot: 5 ml
Final Volume: 5 ml
Result Units: mg/l
Clean DF: 1
File Name: 080131A.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
7782-49-2	SELENIUM	1	0.0099	0.005		

Data Package ID: IT0801254-1

Total Recoverable ICP Metals

Method SW6010B

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0801228

Client Name: ENSR Consulting and Engineering

Client Project ID: Brown Trout 12899-001

Field ID:	HT+Ions & Metals
Lab ID:	0801228-1

Sample Matrix: WATER
% Moisture: N/A
Date Collected: 28-Jan-08
Date Extracted: 29-Jan-08
Date Analyzed: 30-Jan-08
Prep Method: SW3005 Rev A

Prep Batch: IP080129-4
QCBatchID: IP080129-4-2
Run ID: IT080130-2A1
Cleanup: NONE
Basis: As Received

Sample Allquot: 50 g
Final Volume: 50 g
Result Units: mg/l
Clean DF: 1
File Name: 080130A.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
7782-49-2	SELENIUM	1	0.005	0.005	U	

Data Package ID: IT0801228-1

Date Printed: Monday, February 11, 2008

Paragon Analytics
LIMS Version: 6.114A

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Total Recoverable SELENIUM

Method SW6010B

Sample Results

Lab Name: Paragon Analytics
Client Name: ENSR Consulting and Engineering
Client Project ID: Brown Trout 12699-001
Work Order Number: 0802095 Final Volume: 50 g
Reporting Basis: As Received Matrix: WATER
Prep Method: SW3005A Result Units: mg/l

Client Sample ID	Lab ID	Date Collected	Date Prepared	Date Analyzed	Percent Moisture	Dilution Factor	Result	Reporting Limit	Flag	Sample Aliquot
HT + METALS	0802095-2	02/12/2008	02/19/2008	02/20/2008	N/A	1	0.005	0.005	U	50 g
HIGH 003	0802095-3	02/12/2008	02/19/2008	02/20/2008	N/A	1	0.005	0.005	U	50 g
HIGH 010	0802095-4	02/12/2008	02/19/2008	02/20/2008	N/A	1	0.005	0.005	U	50 g
BKD 009	0802095-5	02/12/2008	02/19/2008	02/20/2008	N/A	1	0.005	0.005	U	50 g

Comments:

1. ND or U = Not Detected at or above the client requested detection limit.

Data Package ID: *IT0802095-1*

Ion Chromatography

Method EPA300.0 Revision 2.1

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0710227

Client Name: ENSR Consulting and Engineering

Client Project ID: Brown Trout1269 9-001

Field ID: HT from panel
Lab ID: 0710227-1

Sample Matrix: WATER

% Moisture: N/A

Date Collected: 23-Oct-07

Date Extracted: 23-Oct-07

Date Analyzed: 23-Oct-07

Prep Method: NONE

Prep Batch: IC071023-1

QCBatchID: IC071023-1-1

Run ID: ic071023-1a

Cleanup: NONE

Basis: As Received

Sample Aliquot: 5 ml

Final Volume: 5 ml

Result Units: mg/l

Clean DF: 1

File Name: 71023_020

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
14808-79-8	SULFATE	1	17	1		

Data Package ID: ic0710227-1

Date Printed: Thursday, October 25, 2007

Paragon Analytics

LIMS Version: 8.079A

Page 1 of 1

SULFATE

Method EPA300.0 Revision 2.1

Sample Results

Lab Name: Paragon Analytics

Client Name: ENSR Consulting and Engineering

Client Project ID: Brown Trout 12699-001

Work Order Number: 0711270

Final Volume: 5 ml

Reporting Basis: As Received

Matrix: WATER

Prep Method: NONE

Result Units: mg/l

Client Sample ID	Lab ID	Date Collected	Date Prepared	Date Analyzed	Percent Moisture	Dilution Factor	Result	Reporting Limit	Flag	Sample Aliquot
HT + Ions	0711270-1	10/26/2007	11/30/2007	11/30/2007	N/A	1	28	1		5 ml
Modified HT Ions	0711270-3	11/17/2007	11/30/2007	11/30/2007	N/A	1	26	1		5 ml
HT + Ions	0711270-5	11/21/2007	11/30/2007	11/30/2007	N/A	1	24	1		5 ml
HT + Ions	0711270-6	11/29/2007	11/30/2007	11/30/2007	N/A	1	23	1		5 ml

Comments:

1. ND or U = Not Detected at or above the client requested detection limit.

Data Package ID: *ic0711270-1*

Ion Chromatography

Method EPA300.0 Revision 2.1

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0711046

Client Name: ENSR Consulting and Engineering

Client Project ID: Brown Trout1289 9-001

Field ID: HT from dilutor panel
Lab ID: 0711046-1

Sample Matrix: WATER
% Moisture: N/A
Date Collected: 01-Nov-07
Date Extracted: 06-Nov-07
Date Analyzed: 06-Nov-07
Prep Method: NONE

Prep Batch: IC071106-1
QCBatchID: IC071106-1-1
Run ID: ic071106-1a
Cleanup: NONE
Basis: As Received

Sample Allquot: 5 ml
Final Volume: 5 ml
Result Units: mg/l
Clean DF: 1
File Name: 71106_044.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
14808-79-8	SULFATE	1	22	1		

Data Package ID: ic0711046-1

Date Printed: Monday, November 12, 2007

Paragon Analytics
LIMS Version: 6.088A

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Ion Chromatography

Method EPA300.0 Revision 2.1

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0712041

Client Name: ENSR Consulting and Engineering

ClientProject ID: Brown Trout 12699-001

Field ID: HT + Ions
Lab ID: 0712041-1

Sample Matrix: LIQUID
% Moisture: N/A
Date Collected: 08-Dec-07
Date Extracted: 10-Dec-07
Date Analyzed: 10-Dec-07
Prep Method: NONE

Prep Batch: IC071210-1
QCBatchID: IC071210-1-1
Run ID: ic071210-1a
Cleanup: NONE
Basis: As Received

Sample Allotment: 5 ml
Final Volume: 5 ml
Result Units: mg/l
Clean DF: 1
File Name: 71210_023.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
14808-79-8	SULFATE	1	27	1		

Data Package ID: ic0712041-1

Ion Chromatography

Method EPA300.0 Revision 2.1

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0712111

Client Name: ENSR Consulting and Engineering

Client Project ID: Brown Trout 12699-001

Field ID: HT+ Ions

Lab ID: 0712111-1

Sample Matrix: WATER

% Moisture: N/A

Date Collected: 13-Dec-07

Date Extracted: 14-Dec-07

Date Analyzed: 14-Dec-07

Prep Method: NONE

Prep Batch: IC071214-1

QCBatchID: IC071214-1-1

Run ID: ic071214-1a

Cleanup: NONE

Basis: As Received

Sample Allquot: 5 ml

Final Volume: 5 ml

Result Units: mg/l

Clean DF: 1

File Name: 71214_020.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
14808-79-8	SULFATE	1	27	1		

Data Package ID: ic0712111-1

Date Printed: Thursday, December 20, 2007

Paragon Analytics

LIMS Version: 6.097A

Page 1 of 1

Ion Chromatography

Method EPA300.0 Revision 2.1

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0801009

Client Name: ENSR Consulting and Engineering

ClientProject ID: Brown Trout 12699-001

Field ID:	HT + Ions
Lab ID:	0801009-1

Sample Matrix: WATER

% Moisture: N/A

Date Collected: 03-Jan-08

Date Extracted: 04-Jan-08

Date Analyzed: 04-Jan-08

Prep Method: NONE

Prep Batch: IC080104-1

QC Batch ID: IC080104-1-1

Run ID: ic070104-1a

Cleanup: NONE

Basis: As Received

Sample Aliquot: 5 ml

Final Volume: 5 ml

Result Units: mg/l

Clean DF: 1

File Name: 80104_069.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
14808-79-8	SULFATE	1	22	1		

Data Package ID: ic0801009-1

Date Printed: Wednesday, January 09, 2008

Paragon Analytics

LIMS Version: 6.103A

Page 1 of 1

Ion Chromatography

Method EPA300.0 Revision 2.1

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0801127

Client Name: ENSR Consulting and Engineering

ClientProject ID: Brown Trout12699-001

Field ID: HT
Lab ID: 0801127-1

Sample Matrix: WATER

% Moisture: N/A

Date Collected: 16-Jan-08

Date Extracted: 17-Jan-08

Date Analyzed: 17-Jan-08

Prep Method: NONE

Prep Batch: IC080117-1

QCBatchID: IC080117-1-1

Run ID: ic080117-1a

Cleanup: NONE

Basis: As Received

Sample Aliquot: 5 ml

Final Volume: 5 ml

Result Units: mg/l

Clean DF: 1

File Name: 80117_041.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
14808-79-8	SULFATE	1	19	1		

Data Package ID: ic0801127-1

Ion Chromatography

Method EPA300.0 Revision 2.1

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0801228

Client Name: ENSR Consulting and Engineering

ClientProject ID: Brown Trout1269 9-001

Field ID:	HT+Ions & Metals
Lab ID:	0801228-1

Sample Matrix: WATER

% Moisture: N/A

Date Collected: 28-Jan-08

Date Extracted: 30-Jan-08

Date Analyzed: 30-Jan-08

Prep Method: NONE

Prep Batch: IC080130-1

QCBatchID: IC080130-1-1

Run ID: ic080130-1a

Cleanup: NONE

Basis: As Received

Sample Aliquot: 5 ml

Final Volume: 5 ml

Result Units: mg/l

Clean DF: 1

File Name: 80130_025.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
14808-79-8	SULFATE	1	23	1		

Data Package ID: ic0801228-1

Date Printed: Friday, February 08, 2008

Paragon Analytics

LIMS Version: 6.114A

Page 1 of 1

Ion Chromatography

Method EPA300.0 Revision 2.1

Sample Results

Lab Name: Paragon Analytics

Work Order Number: 0802095

Client Name: ENSR Consulting and Engineering

ClientProject ID: Brown Trout1269 9-001

Field ID:	HT + IONS
Lab ID:	0802095-1

Sample Matrix: WATER

% Moisture: N/A

Date Collected: 12-Feb-08

Date Extracted: 19-Feb-08

Date Analyzed: 20-Feb-08

Prep Method: NONE

Prep Batch: IC080219-1

QC Batch ID: IC080219-1-1

Run ID: ic080219-1a

Cleanup: NONE

Basis: As Received

Sample Aliquot: 5 ml

Final Volume: 5 ml

Result Units: mg/l

Clean DF: 1

File Name: 80219_062.

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	Result Qualifier	EPA Qualifier
14808-79-8	SULFATE	1	19	1		

Data Package ID: ic0802095-1

Date Printed: Friday, February 22, 2008

Paragon Analytics

LIMS Version: 6.117A

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COLUMBIA ANALYTICAL SERVICES, INC.

Analytical Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0712111
Date Collected: 10/25-11/14/07
Date Received: 12/21/07

Solids, Total

Prep Method: NONE
Analysis Method: Freeze Dry
Test Notes:

Units: PERCENT
Basis: Wet

Sample Name	Lab Code	Date Analyzed	Result	Result Notes
SM1007-SNFH-FT0058	K0712111-001	1/10/08	31.2	
SM1007-SNFH-FT0059	K0712111-002	1/10/08	26.0	
SM1007-SNFH-FT0060	K0712111-003	1/10/08	31.1	
SM1007-SNFH-FT0061	K0712111-004	1/10/08	23.8	
SM1007-SNFH-FT0062	K0712111-005	1/10/08	31.0	
SM1007-SNFH-FT0063	K0712111-006	1/10/08	29.5	
SM1007-SNFH-FT0064	K0712111-007	1/10/08	27.9	
SM1007-SNFH-FT0065	K0712111-008	1/10/08	29.8	
SM1107-LSV2c-FT0066	K0712111-009	1/10/08	32.4	
SM1107-LSV2c-FT0067	K0712111-010	1/10/08	33.9	
SM1107-LSV2c-FT0068	K0712111-011	1/10/08	32.1	
SM1107-LSV2c-FT0069	K0712111-012	1/10/08	32.9	
SM1107-LSV2c-FT0070	K0712111-013	1/10/08	25.8	
SM1107-LSV2c-FT0071	K0712111-014	1/10/08	32.2	
SM1107-LSV2c-FT0072	K0712111-015	1/10/08	32.1	
SM1107-LSV2c-FT0073	K0712111-016	1/10/08	33.4	
SM1107-LSV2c-FT0074	K0712111-017	1/10/08	30.6	
SM1107-LSV2c-FT0075	K0712111-018	1/10/08	31.4	
SM1107-LSV2c-FT0076	K0712111-019	1/10/08	31.7	
SM1107-LSV2c-FT0077	K0712111-020	1/10/08	31.7	

COLUMBIA ANALYTICAL SERVICES, INC.

Analytical Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0712111
Date Collected: 11/14-12/4/07
Date Received: 12/21/07

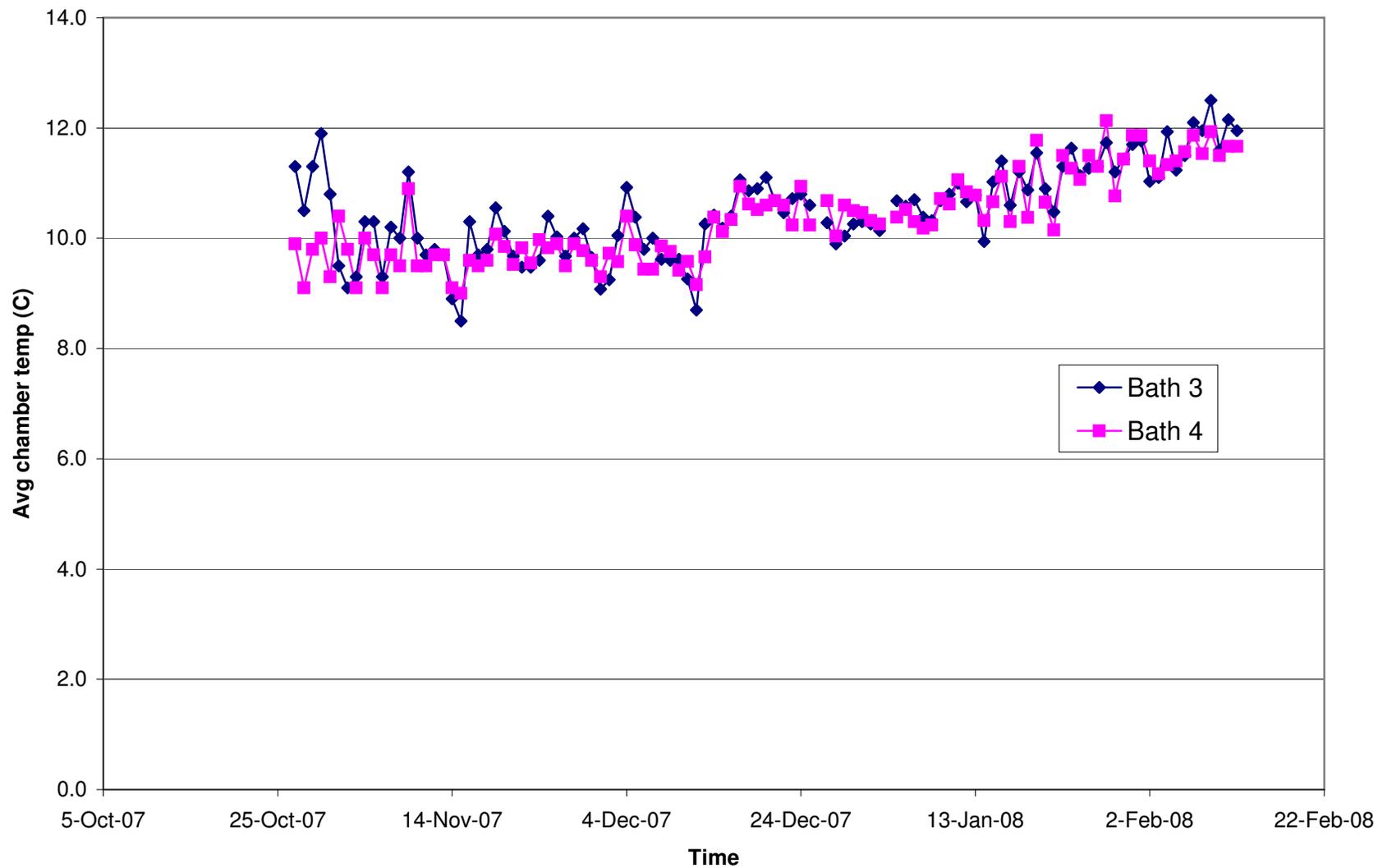
Solids, Total

Prep Method: NONE
Analysis Method: Freeze Dry
Test Notes:

Units: PERCENT
Basis: Wet

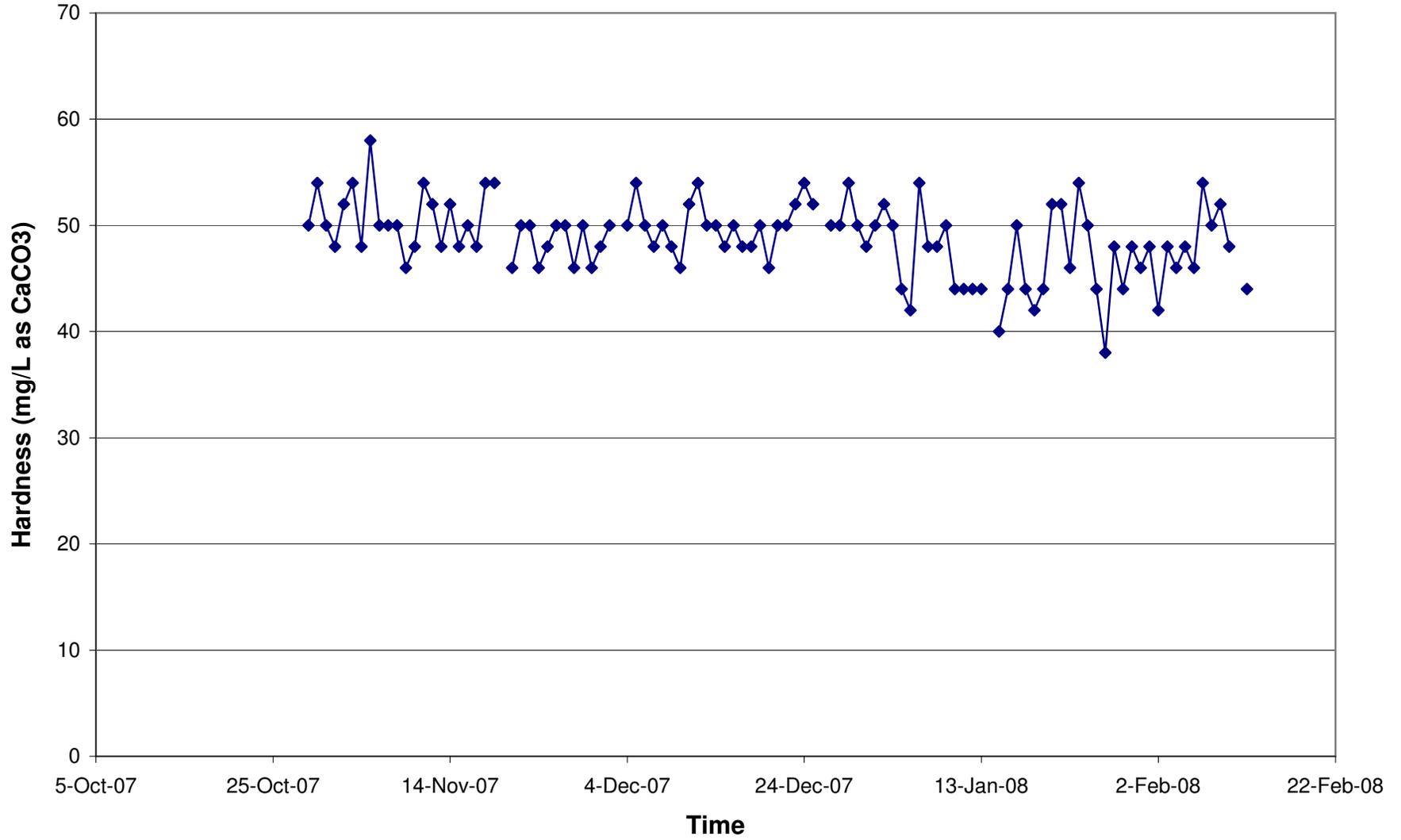
Sample Name	Lab Code	Date Analyzed	Result	Result Notes
SM1107-LSV2c-FT0078	K0712111-021	1/10/08	32.6	
SM1107-LSV2c-FT0079	K0712111-022	1/10/08	33.9	
SM1107-CC150-FT0080	K0712111-023	1/10/08	31.9	
SM1107-CC150-FT0081	K0712111-024	1/10/08	30.6	
SM1107-CC150-FT0082	K0712111-025	1/10/08	30.5	
SM1107-CC150-FT0083	K0712111-026	1/10/08	30.4	
SM1107-CC150-FT0084	K0712111-027	1/10/08	31.0	
SM1107-CC150-FT0085	K0712111-028	1/10/08	32.2	
SM1107-CC150-FT0086	K0712111-029	1/10/08	32.2	
SM1107-CC150-FT0087	K0712111-030	1/10/08	33.1	
SM1107-CC150-FT0088	K0712111-031	1/10/08	31.9	
SM1107-CC350-FT0089	K0712111-032	1/10/08	29.9	
SM1107-CC350-FT0090	K0712111-033	1/10/08	31.9	
SM1107-CC350-FT0091	K0712111-034	1/10/08	33.5	
SM1207-SPCFH-FT0092	K0712111-035	1/10/08	33.2	

Average Daily Chamber temp



Water temperature in test chambers for Brown trout studies																
12699-001																
Filename: water qual.xls																
Date	Test Day	SC treatments		SPC treatments		CC-150 (BKD) treatments		CC-350 (LOW) treatments		LSV2C (HIGH) treatments		Daily Avg bath #3	Temp (°C)	bath #3	bath #4	
		Water temperature (°C)		Water temperature (°C)		Water temperature (°C)		Water temperature (°C)		Water temperature (°C)						
		bath #3	bath #4	bath #3	bath #4	bath #3	bath #4	bath #3	bath #4	bath #3	bath #4					
25-Oct-07	0															
26-Oct-07	1															
27-Oct-07	2	11.3	9.9													11.3 9.9
28-Oct-07	3	10.5	9.1													10.5 9.1
29-Oct-07	4	11.3	9.8													11.3 9.8
30-Oct-07	5	11.9	10.0													11.9 10.0
31-Oct-07	6	10.8	9.3													10.8 9.3
1-Nov-07	7	9.5	10.4													9.5 10.4
2-Nov-07	8	9.1	9.8													9.1 9.8
3-Nov-07	9	9.3	9.1													9.3 9.1
4-Nov-07	10	10.3	10.0													10.3 10.0
5-Nov-07	11	10.3	9.7													10.3 9.7
6-Nov-07	12	9.3	9.1													9.3 9.1
7-Nov-07	13	10.2	9.7													10.2 9.7
8-Nov-07	14	10.0	9.5													10.0 9.5
9-Nov-07	15	11.2	10.9													11.2 10.9
10-Nov-07	16	10.0	9.5													10.0 9.5
11-Nov-07	17	9.7	9.5													9.7 9.5
12-Nov-07	18	9.8	9.7													9.8 9.7
13-Nov-07	19	9.7	9.7													9.7 9.7
14-Nov-07	20	8.9	9.1													8.9 9.1
15-Nov-07	21	8.5	9.0													8.5 9.0
16-Nov-07	22	10.3	9.6								0	10.3	9.6			10.3 9.6
17-Nov-07	23	9.7	9.5					0								9.7 9.5
18-Nov-07	24	9.8	9.6					1								9.8 9.6
19-Nov-07	25	10.6	9.7					2	10.8	10.6		3	10.5	9.7		10.6 10.1
20-Nov-07	26	10.2	9.7					3	9.9	10.3		4	10.1	10.2		10.1 9.9
21-Nov-07	27	9.4	9.1					4	9.7	9.5		5	9.7	9.3		9.7 9.5
22-Nov-07	28	9.3	9.4					5	9.4	10.1		6	9.6	10.4		9.5 9.8
23-Nov-07	29	9.8	9.3					6	9	9.8		7	9.3	8.8		9.5 9.6
24-Nov-07	30	9.7	9.3					7	9.1	9.6		8	9.8	10.6		9.6 10.0
25-Nov-07	31	10.0	9.3					8	10.7	10.2		9	10.5	10.2		10.4 9.8
26-Nov-07	32	9.8	9.6					9	9.8	9.5		10	10.4	10.1		10.0 9.9
27-Nov-07	33	9.7	9.3					10	9.2	9.4		11	9.6	9.3		9.7 9.5
28-Nov-07	34	10.3	9.6					11	9.8	10.1		12	10.1	10.5		10.0 9.9
29-Nov-07	35	10.1	9.5					12	9.9	9.7		13	10.3	10.4		10.2 9.8
30-Nov-07	36	9.2	9.4					13	10.3	9.5		14	9.8	9.8		9.7 9.6
1-Dec-07	37	9.4	8.7					14	8.7	9.8		15	9.1	9.2		9.1 9.3
2-Dec-07	38	9.1	9.3					15	8.8	9.1		16	9.6	10.4		9.3 9.7
3-Dec-07	39	9.9	9.6					16	10.4	9.9		17	9.9	9.4		10.1 9.6
4-Dec-07	40	11.2	10.2	0	11.3	10.1		17	10.3	10.4		18	11	11.2		10.9 10.4
5-Dec-07	41	10.2	9.4	1	10.2	10.1		18	10.7	9.5		19	10.4	10.1		10.4 9.9
6-Dec-07	42	9.6	9.7	2	9.4	9.4		19	9.8	9.3		20	10.1	9.3		9.8 9.4
7-Dec-07	43	10.0	9.3	3	10.5	9.3		20	9.7	9.3		21	9.5	9.2		10.0 9.4
8-Dec-07	44	10.1	9.8	4	9.3	10		21	9.1	9.8		22	9.8	10.1		9.6 9.9
9-Dec-07	45	9.7	9.4	5	9.3	9.7		22	9.7	10.1		23	9.8	9.8		9.6 9.8
10-Dec-07	46	9.7	9.3	6	9.8	9.5		23	9.9	9.5		24	9.6	9.4		9.6 9.4
11-Dec-07	47	9.8	9.4	7	9.2	9.3		24	8.9	9.3		25	9.1	10.1		9.3 9.6
12-Dec-07	48	8.5	9.1	8	8.6	9		25	8.9	9.5		26	8.7	9.1		8.7 9.2
13-Dec-07	49	10.0	9.6	9	10	9.7		26	10.5	9.9		27	10.4	9.6		10.3 9.7
14-Dec-07	50	10.7	10.0	10	10	10.2		27	10.4	10.1		28	10.7	10.5		10.4 10.4
15-Dec-07	51	10.5	10.3	11	9.8	10		28	9.7	10.1		29	10.5	10.2		10.2 10.1
16-Dec-07	52	10.1	10.5	12	10.7	10.1		29	10.3	10		30	10.5	9.8		10.4 10.3
17-Dec-07	53	11.3	10.4	13	10.8	10.7		30	11.3	11.1		31	10.4	12.1		11.1 10.9
18-Dec-07	54	11.4	10.1	14	10.2	10.3		31	10.7	10.3		32	11	10.9		10.9 10.6
19-Dec-07	55	11.8	10.2	15	11.2	10.3		32	9.7	10.3		33	11.1	10.9		10.9 10.5
20-Dec-07	56	11.3	10.6	16	10.9	10.6		33	11.3	11.2		34	10.7	10.2		11.1 10.6
21-Dec-07	57	10.6	10.6	17	10.1	10.4		34	11.2	10.6		35	10.9	11.4		10.7 10.7
22-Dec-07	58	10.3	10.5	18	10.1	11		35	10.9	10.6		36	10.1	9.9		10.5 10.6
23-Dec-07	59	10.0	10.5	19	11.1	10.3		36	10.9	10		37	10.7	10.3		10.7 10.2
24-Dec-07	60	10.9	10.2	20	10.6	10.4		37	10.9	11.2		38	10.7	11.4		10.8 10.9
25-Dec-07	61	10.2	10.1	21	10.9	10.7		38	10.9	10.1		39	10.2	10.1		10.6 10.2
26-Dec-07	62			22				39				40				
27-Dec-07	63	9.8	10.5	23	10.4	10.3		40	10.2	10.5		41	10.7	10.9		10.3 10.7
28-Dec-07	64	9.7	10.4	24	9.7	9.9		41	10.2	10.3		42	10	9.9		9.9 10.0
29-Dec-07	65	10.7	10.7	25	9.5	10.4		42	9.9	10.4		43	10	10.7		10.0 10.6
30-Dec-07	66	10.5	11.1	26	10.4	10		43	10.2	10.5		44	9.9	10.7		10.3 10.5
31-Dec-07	67	10.5	10.8	27	9.9	11.1		44	10.3	9.6		45	10.3	9.8		10.3 10.5
1-Jan-08	68	11.1	10.8	28	10.3	10.5		45	10.3	9.7		46	9.5	10.5		10.3 10.3
2-Jan-08	69	11.1	10.5	29	9.8	9.8		46	9.9	9.9		47	9.7	10.4		10.1 10.3
3-Jan-08	70			30				47				48				
4-Jan-08	71	10.7	10.6	31	11.1	10.6		48	10.6	10.5		49	10.5	10.4		10.7 10.4
5-Jan-08	72	10.9	11.0	32	10.9	10.6		49	10.3	10.4		50	10.6	10.6		10.6 10.5
6-Jan-08	73	11.1	10.6	33	10.8	10.9		50	10.9	10.4		51	10.4	9.8		10.7 10.3
7-Jan-08	74	10.8	10.5	34	10.6	10.5		51	10	9.7		52	10.3	10.3		10.4 10.2
8-Jan-08	75	10.7	10.9	35	10.9	10.6		52	10.3	9.5		53	9.5	10.3		10.3 10.2
9-Jan-08	76	11.1	11.2	36	11.1	11.3		53	10.2	11		54	10.7	10		10.7 10.7
10-Jan-08	77	11.4	11.1	37	11.1	10.9		54	10.7	10.5		55	10.1	10.3		10.8 10.6
11-Jan-08	78	11.3	11.3	38	11.4	11.1		55	10.6	10.6		56	10.9	11.2		11.0 11.1
12-Jan-08	79	10.9	11.4	39	11.3	11.4		56	10.1	11		57	10.5	10.2		10.7 10.8
13-Jan-08	80	11.2	11.3	40	11.3	10.7		57	10.3	10.7		58	10.6	10.9		10.8 10.8
14-Jan-08	81	10.1	10.9	41	9.9	10		58	9.9	10.2		59	10.1	10.5		9.9 10.3
15-Jan-08	82	11.3	10.9	42	11.3	11.1		59	10.9	10.3		60	10.9	10.7		11.0 10.7
16-Jan-08	83	11.4	11.1	43	11.6	11.6		60	11.4	10.7		61	11.3	10.9		11.4 11.1
17-Jan-08	84	10.8	11.1	44	11	10.7		61	10.3	10		62	10.1	9.7		10.6 10.3
18-Jan-08				45	11.7	11.6		62	10.5	11.7		63	11.1	10.5		11.2 11.3
19-Jan-08				46	11	10.6		63	11	10.7		64	10.5	10.1		10.9 10.4
20-Jan-08				47	11.5	11.9		64	11.7	11.8		65	11.5	11.7		11.6 11.8
21-Jan-08				48	11.2	10.7		65	10.4	10.3		66	10.9	10.9		10.9 10.7
22-Jan-08				49	10.5	10.3		66	10.3	10.3		67	10.7	9.8		10.5 10.2
23-Jan-08								67	11.1	11.2		68	11.1	11.9		11.3 11.5
24-Jan-08								68	11.6	11.4		69	11.6	10.7		11.6 11.3
25-Jan-08								69	10.6	10.7		70	11.3	11.4		11.1 11.1
26-Jan-08								70	11	11.5		71	11.2	11.1		11.3 11.5
27-Jan-08								71	10.9	11.2		72	11.8	11.7		11.3 11.3
28-Jan-08								72	11.5	12.4		73	11.5	12.4		11.7 12.1
29-Jan-08								73	11.3	10.6		74	10.9	10.1		11.2 10.8
30-Jan-08								74	11							

Water hardness measured in dilution water during BT study



Water hardness (mg/L) measurements in Brown trout study (12699-001)					
filename: water qual.xls					
		Water hardness			
Date	Test Day	(mg/L)			
25-Oct-07	D0				
26-Oct-07	D1				
27-Oct-07	D2				
28-Oct-07	D3				
29-Oct-07	D4	50			
30-Oct-07	D5	54			
31-Oct-07	D6	50			
1-Nov-07	D7	48			
2-Nov-07	D8	52			
3-Nov-07	D9	54			
4-Nov-07	D10	48			
5-Nov-07	D11	58			
6-Nov-07	D12	50			
7-Nov-07	D13	50			
8-Nov-07	D14	50			
9-Nov-07	D15	46			
10-Nov-07	D16	48			
11-Nov-07	D17	54			
12-Nov-07	D18	52			
13-Nov-07	D19	48			
14-Nov-07	D20	52			
15-Nov-07	D21	48			
16-Nov-07	D22	50			
17-Nov-07	D23	48			
18-Nov-07	D24	54			
19-Nov-07	D25	54			
20-Nov-07	D26				
21-Nov-07	D27	46			
22-Nov-07	D28	50			
23-Nov-07	D29	50			
24-Nov-07	D30	46			
25-Nov-07	D31	48			
26-Nov-07	D32	50			
27-Nov-07	D33	50			
28-Nov-07	D34	46			
29-Nov-07	D35	50			
30-Nov-07	D36	46			
1-Dec-07	D37	48			
2-Dec-07	D38	50			
3-Dec-07	D39				
4-Dec-07	D40	50			
5-Dec-07	D41	54			
6-Dec-07	D42	50			
7-Dec-07	D43	48			
8-Dec-07	D44	50			
9-Dec-07	D45	48			
10-Dec-07	D46	46			
11-Dec-07	D47	52			
12-Dec-07	D48	54			
13-Dec-07	D49	50			
14-Dec-07	D50	50			
15-Dec-07	D51	48			
16-Dec-07	D52	50			
17-Dec-07	D53	48			
18-Dec-07	D54	48			
19-Dec-07	D55	50			
20-Dec-07	D56	46			
21-Dec-07	D57	50			
22-Dec-07	D58	50			
23-Dec-07	D59	52			
24-Dec-07	D60	54			
25-Dec-07	D61	52			

Water hardness (mg/L) measurements in Brown trout study (12699-001)							
filename: water qual.xls							
		Water hardness					
<u>Date</u>	<u>Test Day</u>	<u>(mg/L)</u>					
26-Dec-07	D62						
27-Dec-07	D63	50					
28-Dec-07	D64	50					
29-Dec-07	D65	54					
30-Dec-07	D66	50					
31-Dec-07	D67	48					
1-Jan-08	D68	50					
2-Jan-08	D69	52					
3-Jan-08	D70	50					
4-Jan-08	D71	44					
5-Jan-08	D72	42					
6-Jan-08	D73	54					
7-Jan-08	D74	48					
8-Jan-08	D75	48					
9-Jan-08	D76	50					
10-Jan-08	D77	44					
11-Jan-08	D78	44					
12-Jan-08	D79	44					
13-Jan-08	D80	44					
14-Jan-08	D81						
15-Jan-08	D82	40	SC fish (Oct. 25 - Jan. 17)				
16-Jan-08	D83	44	<u>Avg</u>	<u>StDev</u>	<u>Min</u>	<u>Max</u>	
17-Jan-08	D84	50	49.32	3.19	40	58	
18-Jan-08		44					
19-Jan-08		42					
20-Jan-08		44	SPC fish (Dec. 4 - Jan. 22)				
21-Jan-08		52	<u>Avg</u>	<u>StDev</u>	<u>Min</u>	<u>Max</u>	
22-Jan-08		52	48.63	3.55	40	54	
23-Jan-08		46					
24-Jan-08		54					
25-Jan-08		50					
26-Jan-08		44					
27-Jan-08		38					
28-Jan-08		48					
29-Jan-08		44					
30-Jan-08		48					
31-Jan-08		46					
1-Feb-08		48					
2-Feb-08		42					
3-Feb-08		48					
4-Feb-08		46					
5-Feb-08		48					
6-Feb-08		46					
7-Feb-08		54					
8-Feb-08		50					
9-Feb-08		52					
10-Feb-08		48	CC-150 & CC-350 fish (Nov. 17- Feb. 12)				
11-Feb-08			<u>Avg</u>	<u>StDev</u>	<u>Min</u>	<u>Max</u>	
12-Feb-08		44	48.36	3.51	38	54	
Average (overall)		48.78					
	StDev	3.506	LSV2C fish (Nov. 16- Feb. 12)				
	Min	38	<u>Avg</u>	<u>StDev</u>	<u>Min</u>	<u>Max</u>	
	Max	58	48.38	3.49	38	54	

Appendix F

Number of organisms and survival rates at different stages during the brown trout reproduction study

Appendix G

Length and dry weight measurements for juvenile brown trout from reproductive study

**Length and dry weight data for juvenile brown trout from each treatment
12699-001**

filename: length & wt.xls

Saratoga Hatchery (SC)

	SC008 std length (mm)	SC008 dry wt (mg)	SC008 (2) std length (mm)	SC008 (2) std length before isopropyl	SC008 (2) dry wt (mg)	SC008 (2) wet wt (mg)
A	21	13.780	21	23	17.360	131.000
B	19	10.360	22	23	19.090	142.100
C	23	18.880	22	22	13.920	104.300
D	21	16.370	21	23	16.010	114.700
E	22	15.450	23	24	20.430	153.300
F	23	22.020	22	23	15.120	118.800
G	22	20.260	21	23	15.290	113.300
H	23	21.530	19	20	12.640	94.700
I	21	14.800	20	22	15.540	111.700
J	20	13.200	21	23	19.190	131.300
K	21	15.320				
L	21	12.310				
M	20	12.750				
N	21	12.000				
O	22	15.710				
P	20	12.230				
Q	20	11.200				
R	22	16.320				
S	21	14.630				
T	20	13.250				
Avg	21.15	15.1185	21.20	22.60	16.4590	121.52
Std	1.137	3.338	1.135	1.075	2.499	17.804
n	20		10		10	

**Length and dry weight data for juvenile brown trout from each treatment
12699-001**

filename: length & wt.xls

CC-350

	CC-350 006 std length (mm)	CC-350 006 dry wt (mg)	CC-350 007 std length (mm)	CC-350 007 dry wy (mg)	CC-350 008 std length (mm)	CC-350 008 dry wt (mg)
A	23	13.250	20	12.130	22	9.470
B	20	11.680	21	8.570	21	10.040
C	22	10.240	23	8.040	21	8.890
D	20	14.390	21	7.700	22	17.650
E	22	14.430	23	15.750	21	11.710
F	22	9.330	21	12.850	19	17.750
G	22	16.810	24	17.980	22	10.830
H	20	8.820	21	6.890	20	9.000
I	23	17.060	23	10.410	20	14.820
J	20	16.820	22	9.170	21	8.870
K	21	8.760	23	13.370	19	10.820
L	20	9.440	20	16.580	20	10.050
M	20	14.070	21	18.080	22	9.470
N	22	10.310	19	11.150	20	11.760
O	21	17.500	20	10.740	20	13.120
P	23	10.380	21	7.660	20	9.370
Q	22	8.000	20	10.480	20	10.300
R	21	14.170	20	8.010	20	14.400
S	23	9.300	21	7.260	21	16.860
T	21	12.090	22	7.360	20	16.600
Avg	21.40	12.3425	21.30	11.0090	20.55	12.0890
Std	1.142	3.148	1.342	3.676	0.945	3.130
n	20		20		20	

**Length and dry weight data for juvenile brown trout from each treatment
12699-001**

filename: length & wt.xls

CC-150

	CC-150 018 std length (mm)	CC-150 018 dry wt (mg)	CC-150 020 std length (mm)	CC-150 020 dry wt (mg)
A	21	16.610	20	12.040
B	20	14.100	21	5.700
C	22	16.740	19	15.550
D	20	10.730	22	6.790
E	19	5.700	20	14.280
F	22	14.280	21	7.590
G	23	15.640	22	7.050
H	22	13.500	22	18.360
I	20	16.560	20	11.320
J	20	11.270	21	10.960
K	21	10.500	20	7.690
L	22	12.970	22	13.100
M	22	8.000	20	14.180
N	22	13.310	18	10.600
O	22	18.330	21	8.360
P	21	8.330	18	13.110
Q	21	14.540	19	14.810
R	22	15.130	19	7.290
S	22	12.720	21	14.980
T	22	15.930	18	11.190
Avg	21.30	13.2445	20.20	11.2475
Std	1.031	3.302	1.361	3.553
n	20		20	

**Length and dry weight data for juvenile brown trout from each treatment
12699-001**

filename: length & wt.xls

LSV2C

	LSV2C 002 std length (mm)	LSV2C 002 dry wt (mg)	LSV2C 003 std length (mm)	LSV2C 003 dry wt (mg)	LSV2C 004 std length (mm)	LSV2C 004 dry wt (mg)	LSV2C 005 std length (mm)	LSV2C 005 dry wt (mg)	LSV2C 008 std length (mm)	LSV2C 008 dry wt (mg)	LSV2C 010 std length (mm)	LSV2C 010 dry wt (mg)	LSV2C 012 std length (mm)	LSV2C 012 dry wt (mg)
A	18	11.850	22	11.760	22	9.620	19	7.080	21	8.870	20	8.010	22	13.810
B	20	17.480	20	7.080	21	11.310	21	12.370	20	9.980	20	6.230	21	12.540
C	20	12.730	21	7.310	21	8.910	19	7.400	20	10.300	20	10.640	22	12.370
D	21	8.140	22	8.010	22	7.600	23	11.710	21	8.950	21	6.770	22	15.000
E	19	9.720	21	12.110	20	11.960	20	10.580	21	8.370	20	7.600	22	12.700
F	20	8.020	21	7.890	22	6.700	20	7.370	21	11.420	21	7.290	22	13.280
G	20	11.680	21	9.420	21	9.520	20	16.890	21	9.490	19	9.570	23	12.550
H	22	10.130	22	9.850	21	10.130	20	8.140	20	12.150	21	14.100	22	13.390
I	21	13.820	22	13.960	23	13.810	21	15.330	21	9.710	19	7.370	22	12.400
J	19	6.820			20	10.720	19	10.990	21	8.670	19	4.690	22	13.470
K	20	10.440			21	9.640	20	13.220	20	8.420	19	6.740	22	12.810
L	20	6.510			20	9.310	20	9.950	20	8.510			21	13.450
M	19	9.840			20	10.450	19	10.270	21	8.760			22	13.460
N	21	12.090			23	10.990	21	11.540	20	8.670			22	14.100
O	20	13.280			21	10.670	19	12.300	20	7.920			22	13.530
P	18	10.030			21	13.330	21	13.230	20	10.230			22	14.940
Q	22	10.780			21	8.040	20	11.210	21	9.020			22	12.850
R	21	15.610			21	7.100	22	6.920	20	10.740			23	12.120
S	22	6.290			21	6.890	21	12.750	20	7.940			22	12.480
T	19	16.130			22	12.880	20	10.090	20	8.720			22	13.870
Avg	20.10	11.0695	21.33	9.7100	21.20	9.9790	20.25	10.9670	20.45	9.3420	19.91	8.0918	22.00	13.2560
Std	1.210	3.161	0.707	2.424	0.894	2.082	1.070	2.712	0.510	1.147	0.831	2.546	0.459	0.814
n	20		9		20		20		20		11		20	

**Length and dry weight data for juvenile brown trout from each treatment
12699-001**

filename: length & wt.xls

LSV2C

	LSV2C 016 std length (mm)	LSV2C 016 dry wt (mg)	LSV2C 017 std length (mm)	LSV2C 017 dry wt (mg)	LSV2C 019 std length (mm)	LSV2C 019 dry wt (mg)	LSV2C 020 std length (mm)	LSV2C 020 dry wt (mg)	LSV2C 021 std length (mm)	LSV2C 021 dry wt (mg)
A	22	17.320	24	25.130	23	26.560	21	13.920	22	8.320
B	19	15.730	23	20.260	25	23.350	21	12.980	23	8.500
C	22	16.060	23	15.130	23	21.680	21	10.990	19	10.380
D	25	14.700	23	24.680	24	18.840	23	7.890	20	10.280
E	22	9.710	23	17.080	23	25.970	21	10.270	20	9.960
F	23	17.160	23	21.450	21	15.450	23	8.570	20	16.840
G	23	16.230	25	13.590	25	13.100	22	8.090	19	7.710
H	22	12.390	23	19.750	23	10.410	22	8.660	20	10.690
I	20	24.040	23	17.630	23	17.300	21	13.900	22	15.690
J	20	11.710	24	21.390	26	19.830	22	9.790	21	4.550
K	22	27.380	24	24.930	23	13.690	21	15.120	18	11.960
L	22	14.060	24	18.110	22	23.830	21	9.360	19	12.530
M	21	9.310	25	15.480	22	10.580	21	16.620	20	10.470
N	21	21.020	24	17.330	26	33.340	24	14.030	21	5.950
O	24	18.480	23	19.570	24	18.080	21	9.520	18	5.510
P	21	18.270	25	16.470	24	23.380	21	14.690	23	13.280
Q	23	12.900	24	9.940	24	17.950	22	12.750	19	9.940
R	23	7.430	21	17.680	26	21.150	22	14.230	21	18.350
S	20	20.420	25	23.820	22	14.510	21	9.250	19	16.000
T	21	12.390	25	18.130	24	17.390	24	10.840	20	6.210
Avg	21.80	15.8355	23.70	18.8775	23.65	19.3195	21.75	11.5735	20.20	10.6560
Std	1.473	4.974	1.031	3.966	1.424	5.764	1.020	2.701	1.473	3.901
n	20		20		20		20		20	

Average length and weight of brown trout (*Salmo trutta*) at test termination.**12699-001-300**

Note: each value represents the average of ~20 individual fish

filename: length & wt.xls

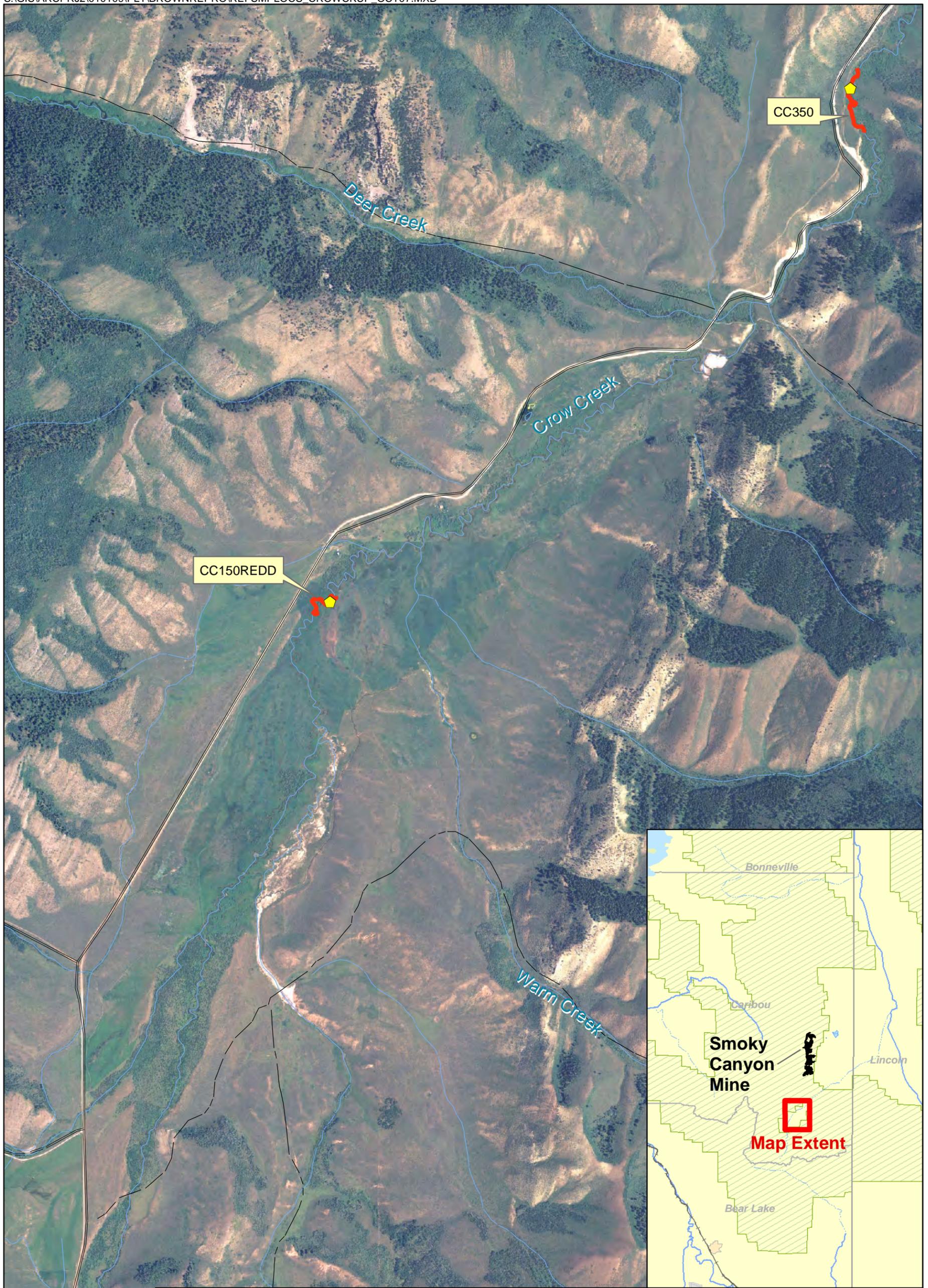
	avg std length (mm)	avg dry wt (mg)	avg std length before Isopropyl	avg wet wt (mg)	Grand Avg std length	Grand avg dry wt (mg)
Saratoga Hatchery						
SC-001	21.45	18.338				
SC-002	22.15	18.470				
SC-003	22.85	22.602				
SC-004	22.20	23.247				
SC-005	21.80	19.386				
SC-006	21.80	18.240				
SC-007	21.05	18.567				
SC-008	21.15	15.119			21.81	19.2459
SC-008 (2)	21.20	16.459	22.60	121.52		
Spring Creek Hatchery						
SPC-001	24.35	22.564				
SPC-002	22.85	20.852				
SPC-003	23.30	21.807				
SPC-004	23.45	21.564				
SPC-005	23.00	20.793				
SPC-006	23.15	22.385			23.35	21.6607
High						
LSV2C-002	20.10	11.069				
LSV2C-003	21.33	9.710				
LSV2C-004	21.20	9.979				
LSV2C-005	20.25	10.967				
LSV2C-006						
LSV2C-007						
LSV2C-008	20.45	9.342				
LSV2C-010	19.91	8.092				
LSV2C-012	22.00	13.256				
LSV2C-016	21.80	15.835				
LSV2C-017	23.70	18.878				
LSV2C-019	23.65	19.320				
LSV2C-020	21.75	11.574				
LSV2C-021	20.20	10.656			21.36	12.3898
Bkd						
CC-150-009	21.85	13.188				
CC-150-011	20.10	9.249				
CC-150-012	21.45	12.840				
CC-150-013	22.55	15.583				
CC-150-015	22.55	14.486				
CC-150-016	22.80	16.736				
CC-150-017	20.95	10.652				
CC-150-018	21.30	13.245				
CC-150-020	20.20	11.248			21.53	13.0250
Low						
CC-350-006	21.40	12.343				
CC-350-007	21.30	11.009				
CC-350-008	20.55	12.089			21.08	11.8135

(measured after storage in isopropyl)

(measured prior to storage in isopropyl)

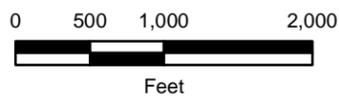
(all other fish measured prior to storage in isopropyl)

APPENDIX B
Detail Figures for Locations where Brown Trout were Collected



Legend

-  Surface Water Sampling Location
-  Fishing Reach



J.R. SIMPLOT COMPANY
SMOKY CANYON MINE

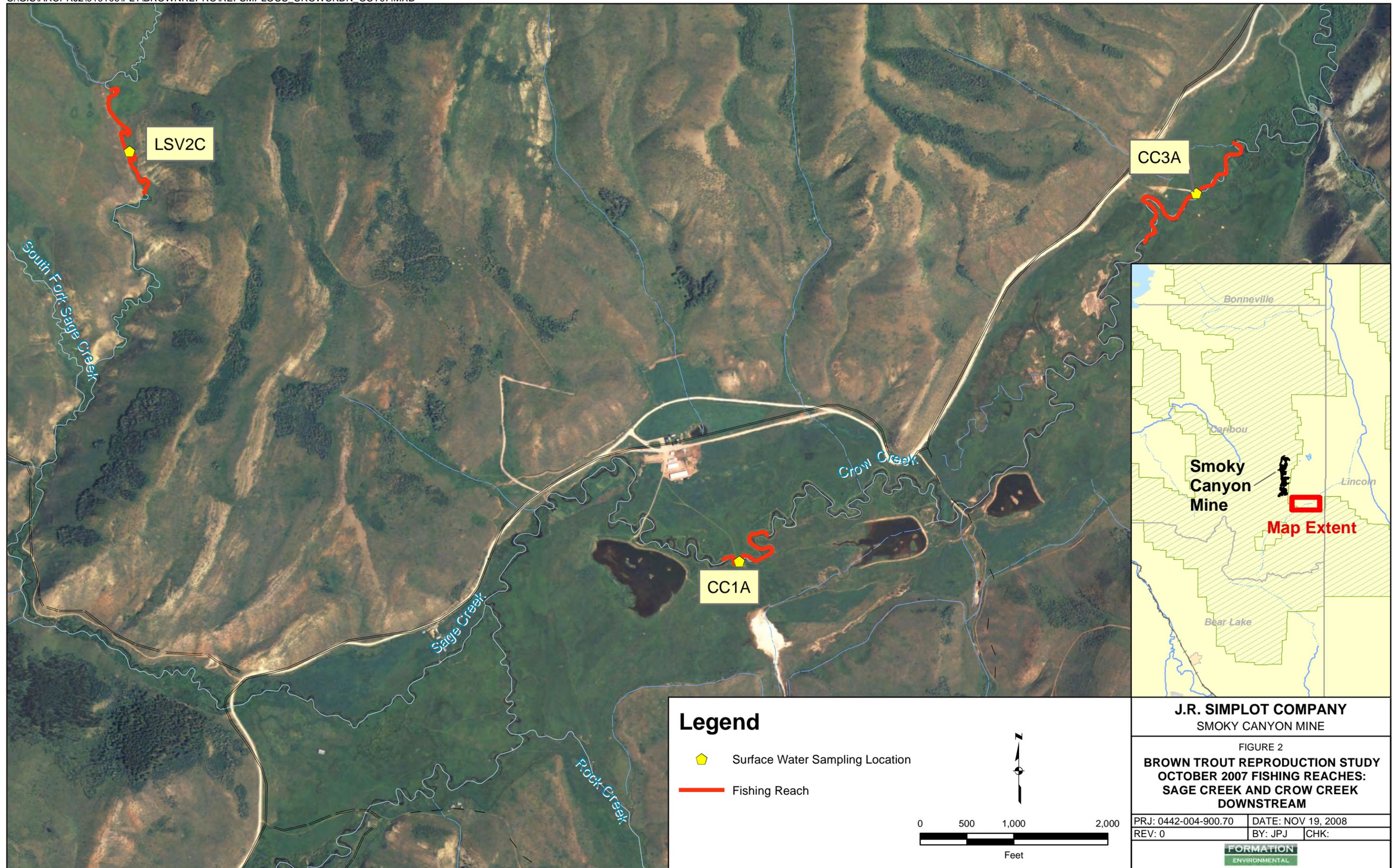
FIGURE 1

**BROWN TROUT REPRODUCTION STUDY
OCTOBER 2007 FISHING REACHES:
CROW CREEK UPSTREAM**

PRJ: 0442-004-900.70 DATE: NOV 19, 2008

REV: 0 BY: JPJ CHK:





Legend

-  Surface Water Sampling Location
-  Fishing Reach

0 500 1,000 2,000

 Feet

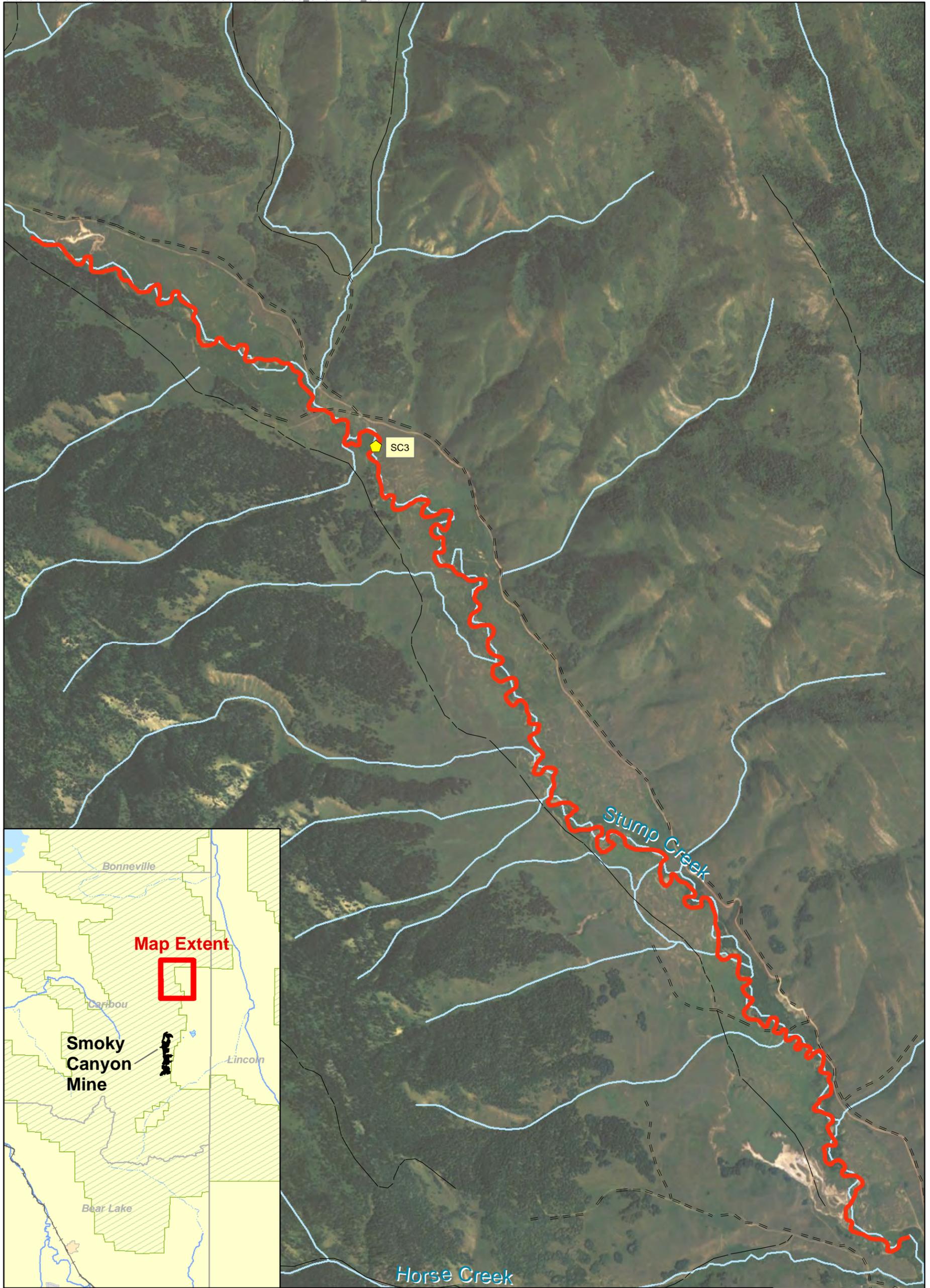

 N

J.R. SIMPLOT COMPANY
SMOKY CANYON MINE

FIGURE 2
BROWN TROUT REPRODUCTION STUDY
OCTOBER 2007 FISHING REACHES:
SAGE CREEK AND CROW CREEK
DOWNSTREAM

PRJ: 0442-004-900.70	DATE: NOV 19, 2008
REV: 0	BY: JPJ CHK:





J.R. SIMPLOT COMPANY
SMOKY CANYON MINE

FIGURE 3

BROWN TROUT REPRODUCTION STUDY
OCTOBER 2007 FISHING REACHES:
STUMP CREEK

PRJ: 0442-004-900.70	DATE: NOV 19, 2008
REV: 0	BY: JPJ CHK:

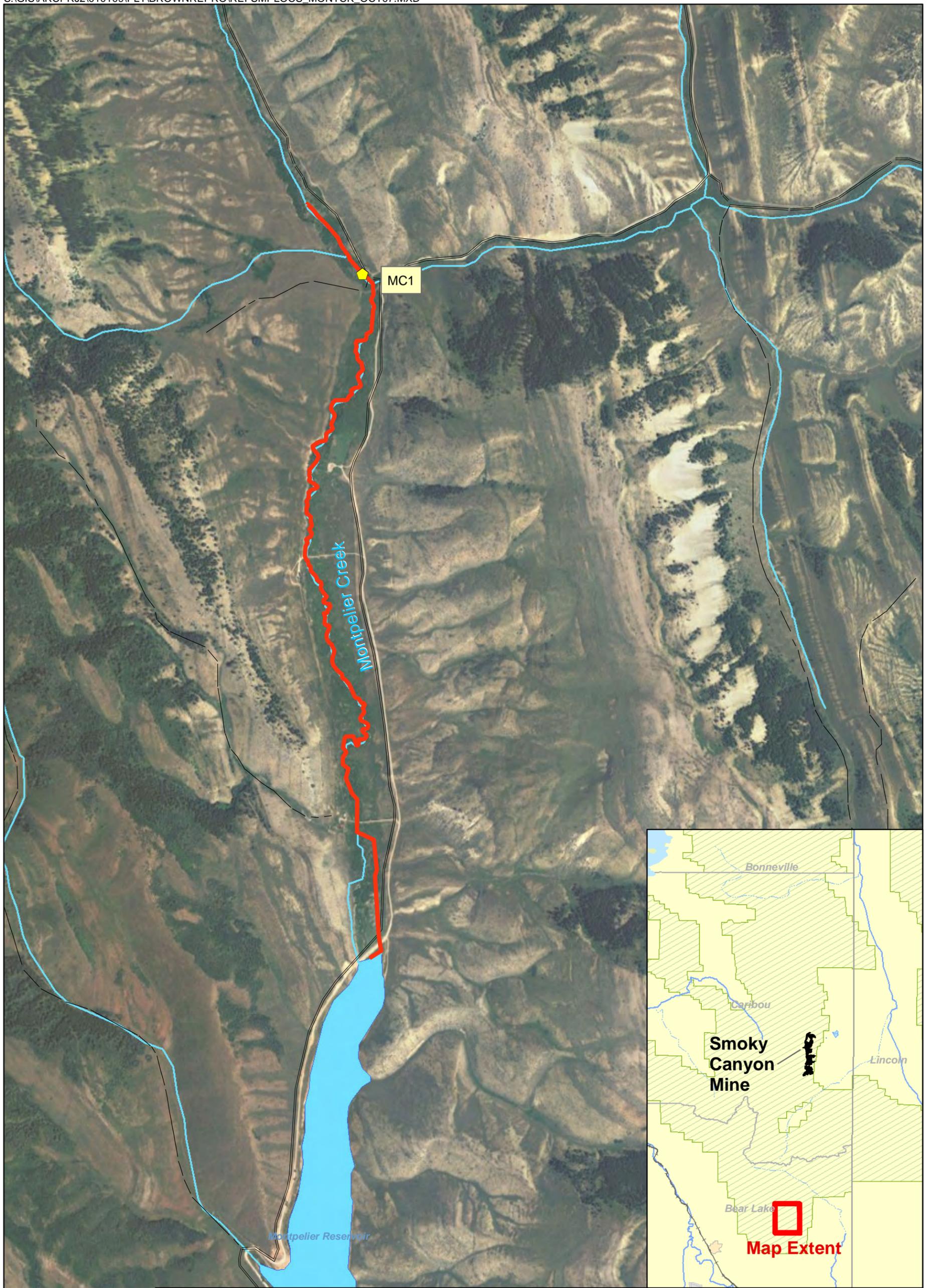
FORMATION
ENVIRONMENTAL

Legend

- Surface Water Sampling Location
- Fishing Reach

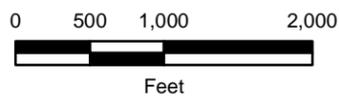
0 500 1,000 2,000
Feet





Legend

-  Surface Water Sampling Location
-  Fishing Reach



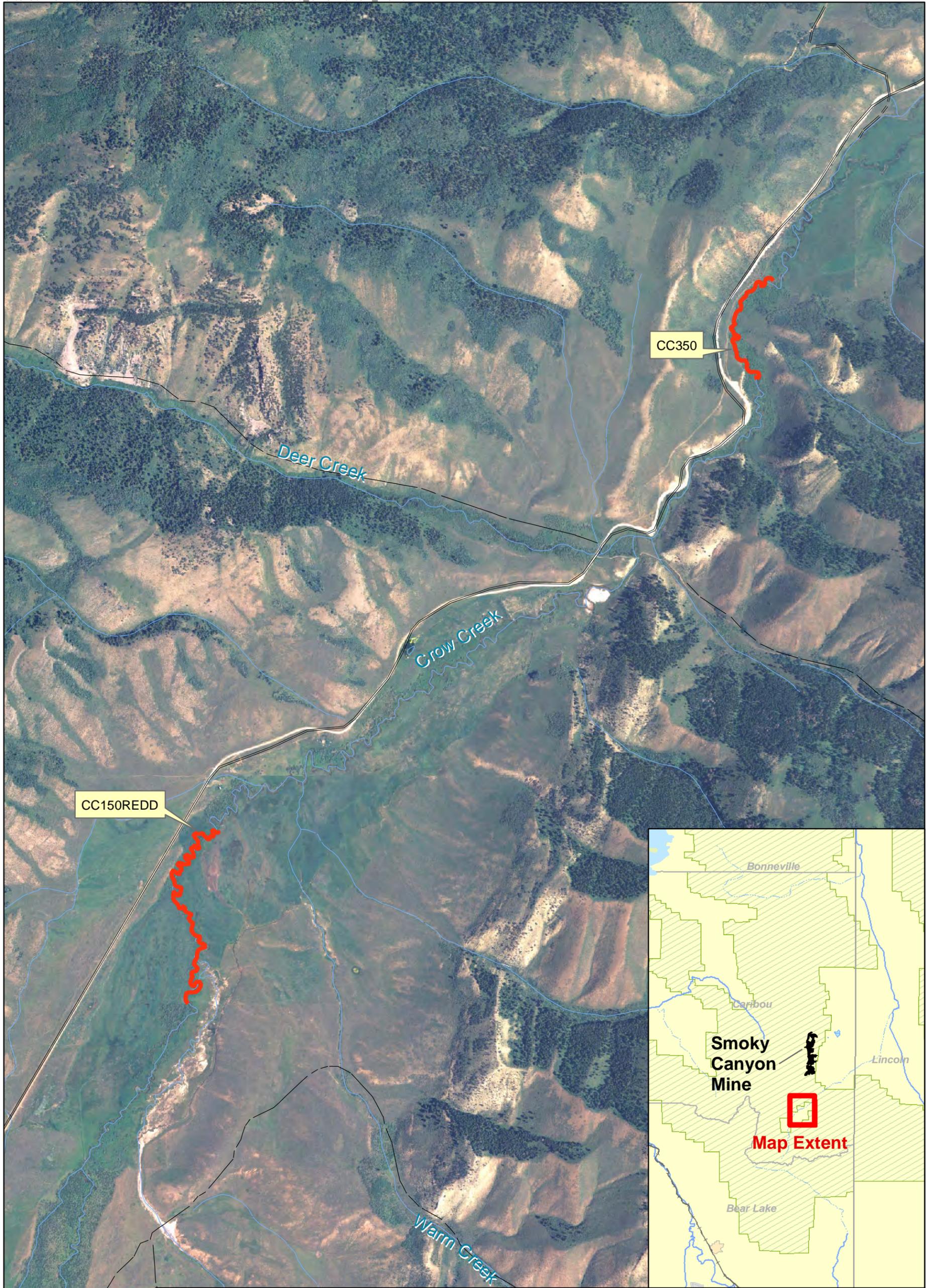
J.R. SIMPLOT COMPANY
SMOKY CANYON MINE

FIGURE 4

**BROWN TROUT REPRODUCTION STUDY
OCTOBER 2007 FISHING REACHES:
MONTPELIER CREEK**

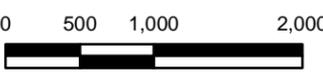
PRJ: 0442-004-900.70	DATE: NOV 19, 2008
REV: 0	BY: JPJ CHK:





Legend

— Fishing Reach



0 500 1,000 2,000
Feet

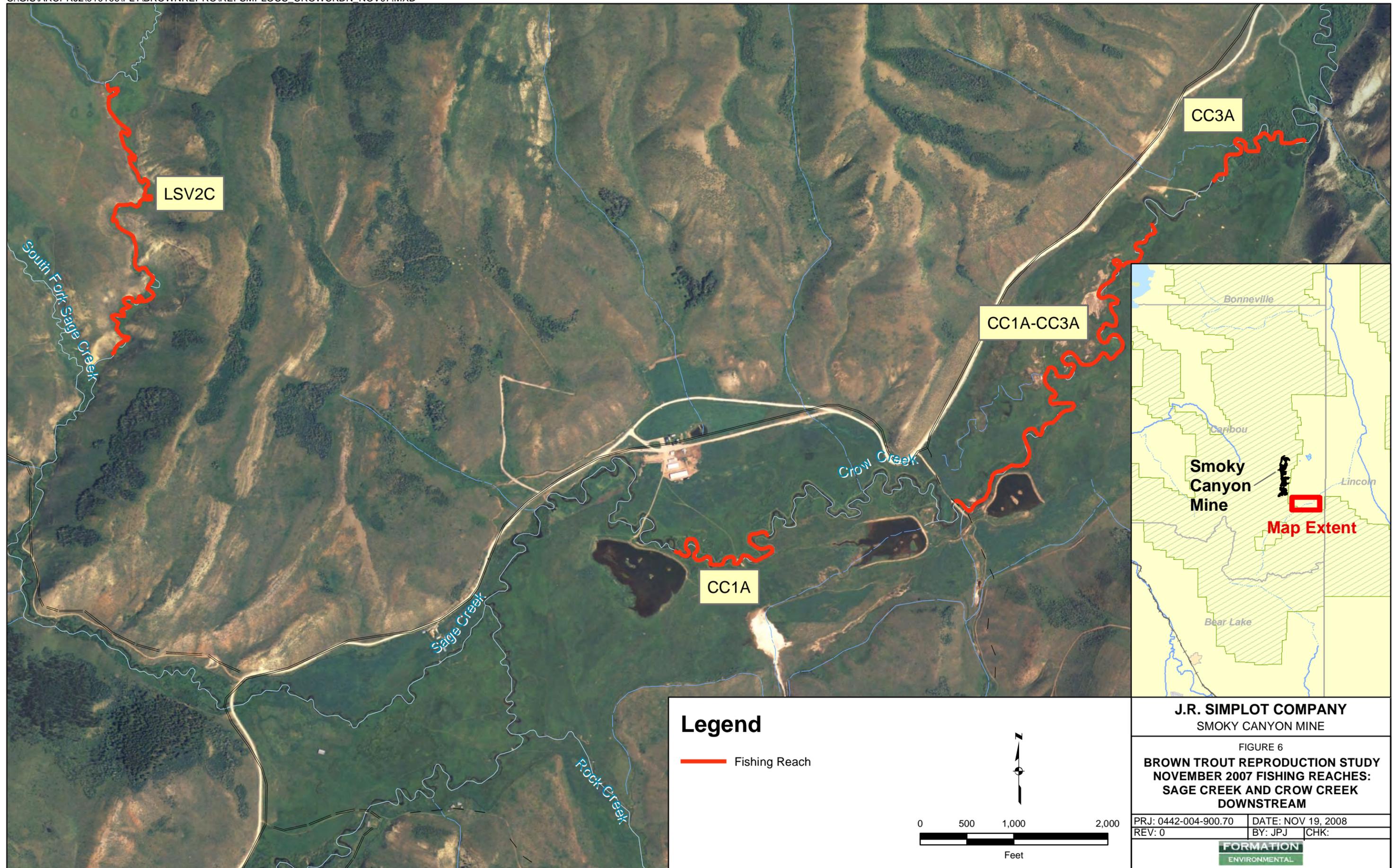
J.R. SIMPLOT COMPANY
SMOKY CANYON MINE

FIGURE 5

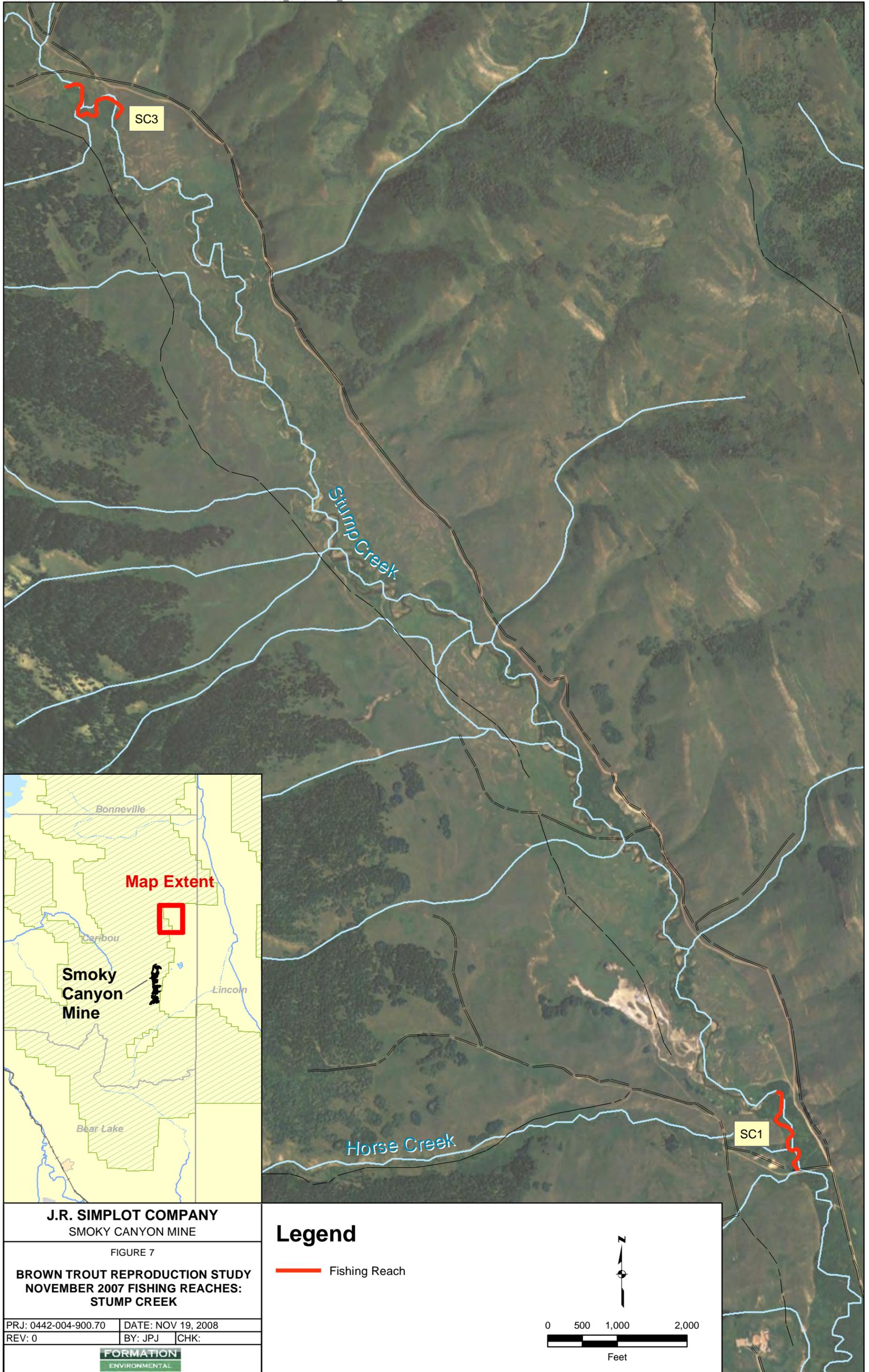
BROWN TROUT REPRODUCTION STUDY
NOVEMBER 2007 FISHING REACHES:
CROW CREEK UPSTREAM

PRJ: 0442-004-900.70	DATE: NOV 19, 2008	
REV: 0	BY: JPJ	CHK:

FORMATION
ENVIRONMENTAL



Legend		
— Fishing Reach		
N ↑		
0 500 1,000 2,000 Feet		
J.R. SIMPLOT COMPANY SMOKY CANYON MINE		
FIGURE 6 BROWN TROUT REPRODUCTION STUDY NOVEMBER 2007 FISHING REACHES: SAGE CREEK AND CROW CREEK DOWNSTREAM		
PRJ: 0442-004-900.70	DATE: NOV 19, 2008	
REV: 0	BY: JPJ	CHK:
FORMATION ENVIRONMENTAL		

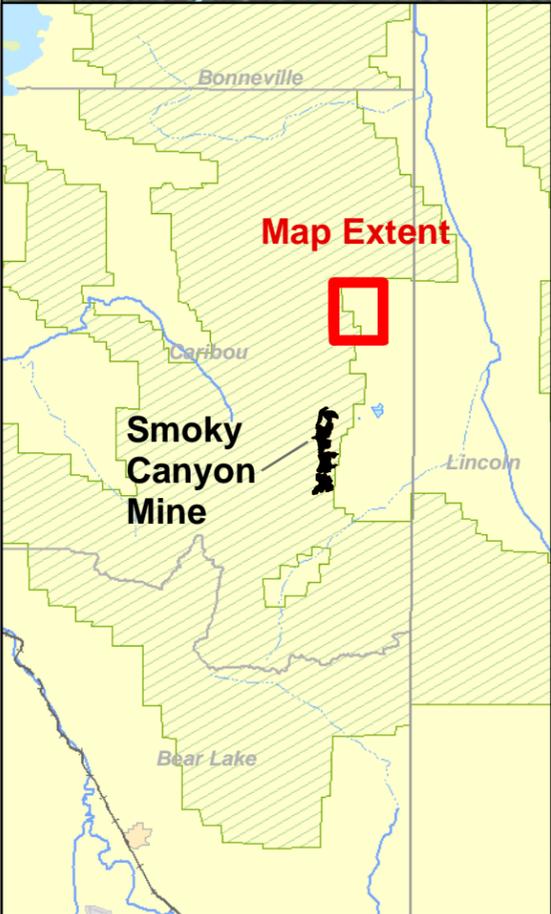


SC3

Stump Creek

SC1

Horse Creek



Map Extent

Smoky Canyon Mine

J.R. SIMPLOT COMPANY
SMOKY CANYON MINE

FIGURE 7

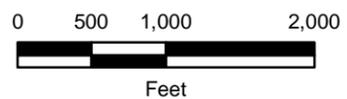
BROWN TROUT REPRODUCTION STUDY
NOVEMBER 2007 FISHING REACHES:
STUMP CREEK

PRJ: 0442-004-900.70	DATE: NOV 19, 2008
REV: 0	BY: JPJ CHK:



Legend

Fishing Reach



APPENDIX C
Photographs of example deformities and Summary of the Deformity Rankings for Each Sample-
CSU

Deformity Assessment

The general scoring criteria were adopted from Holm et al. (2003) and included assessments of craniofacial deformities, mostly of the head, eyes, and jaw, vertebral deformities, fin deformities, and edema. The original publication showed pictures of some deformities but others, particularly the intermediate categories were not illustrated or were poorly described. More specific definitions for each of the assessment categories were developed to give better repeatability and consistency across studies, and to aid others in learning the range of deformities possible.

Deformities in each of the categories described above were given a score from 0-3, with 0 being a normal condition and 3 being the most deformed. Some range finding was conducted over the first several samples to find background and severe levels of deformities in each category. Initial samples were rescored as necessary to bring them into compliance with the standards that were used throughout the assessment. In the second batch of fish analyzed (~100 from 5 LSV2C sites), it was not always possible to score each fish for each category due to the condition of the organism. Therefore, in several cases no scoring was possible.

The protocol for assessing damage was to place several fish, head to the left, in a Petri dish and examine them under a dissecting microscope and 10X magnification. The lateral side was examined for spinal deformities (lordosis, kyphosis), appearance of the eye, head and snout shape, edema, and fin deformities. The fish was turned ventrally to look for mouth deformities and further spinal deformities (scoliosis), turned laterally again for the same criteria as the other side, and then dorsally for issues associated with eyes, head size, spinal deformities.

Craniofacial deformities included shortening of the jaw, snout, and missing or poorly developed eye or eyes, and head shape abnormalities. A slightly shortened lower jaw (\leq 1 lip width) received a 1, a shortened jaw = 2 lip widths or a slightly shortened and slightly disfigured jaw = 2, and a flat lower jaw or much disfigured (non-functional) jaw = 3. An assessment of fish independent of this study revealed that other brown trout of the same size and developmental state did not have the slight deformity that was assessed as CF = 1 for the jaw (J). Thus, the CF = 1 score where the J was concerned were deemed real. A slightly blunted snout (about 50% eye diameter, usually is > than that) = 1, very blunt or flat = 2, deformed or bulbous = 3. Eye deformities were scored as one eye blind or poorly pigmented or poorly developed = 1, both poorly developed = 2, both blind = 3. Skulls that were slightly bulbous ($1/3 >$ normal) = 1, moderately bulbous ($2/3 >$ normal) = 2, and bulbous ($1x$ or > than normal) = 3. Usually factors occurred together so a combination of two "1" conditions = 2, three "1" conditions = 3, or a 1 and a 2 = 3, and so on. For example, a deformed jaw and a blind eye = 2, two blind eyes = 2, but a badly deformed jaw (= 2 alone) plus a blind eye (= 1 alone), = 3.

Skeletal deformities included any deformity of the vertebrae or spines. A slight bend of less than 45 degrees (but > than body width off of straight) or a minor body constriction (e.g. a tight rubber band about the body effect) was given a score of 1, 2 slight bends or constrictions anywhere, or bend of > 45-90 degrees was scored a 2, and multi-directional

bends > 90 degrees were given a 3. Bends caused by skeletal deformities were usually detectable from normal bending of the body during preservation (these fish were usually well preserved, very straight) by presence of a slight or greater bump below the surface of the epidermis on the outside of the bend. However, some fish with SD = 1 had just a very slight bend in the range the deformity described but could be due to preservation or the poor condition of the fish. This was sometimes especially true in larger fish, which may be more muscular and undergo stronger contraction during preservation and thus, bend slightly. A score "CF = 1" was a slight deformity, if at all. The scores of SD = 1 involving kyphosis or lordosis were deemed real because that is an unusual preservation deformity. Also, samples BKD 015 SU (i.e., extra fry from CC-150-015 at swim-up), LOW 008 SU (i.e., extra fry from CC-350-008 at swim-up), and SC 003 SU (i.e., extra fry from SC-003 at swim-up) were re-examined; most fish were very straight so some samples with higher SD scores (e.g., PSU samples) were determined accurate. Thin fish difficult to score, and often looked like they were underfed or starving.

Fin deformities included variation in fin or finfold morphology and a slightly smaller or missing fin (in thin fish, the adipose fin was often absent, indicating fat absorption, not uncommon and scored 1) or one with a bend or incomplete ray development (in older fish) was given a 1, 2 fins damaged or malformed = 2, and > 2 fins malformed or if fins were missing (except adipose) was = 3. Often fins were malformed associated with vertebral deformities that did not permit proper development. Folded finfolds as a result of preservation were not counted.

Edema was not originally scheduled for assessment because it was thought sometimes not a teratogenic effect and may be transitory as fish develop. However, it was assessed because it was common in one early sample and not others, and because it was considered a condition that could affect emergence, mobility, and other factors that may limit survival of fish in the wild. Edema was detected by an obvious swelling and fluid buildup, usually abdominally, and ventrally, which often displaced the gut, and was usually clear fluid that was slightly soft when touched with a blunt probe. The yolk, which was present in some quantity in some study specimens, also created some swelling but was typically yellowish, opaque, and small, and hard to the touch in preservation. Slight edema = 1 was for a fish with up to 1X swelling of the normal body width or depth, up to 2x = 2, and > 2x = 3.

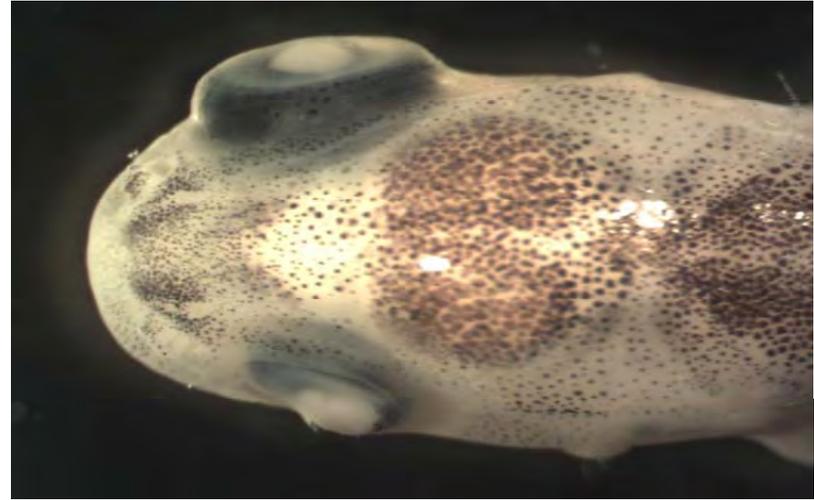
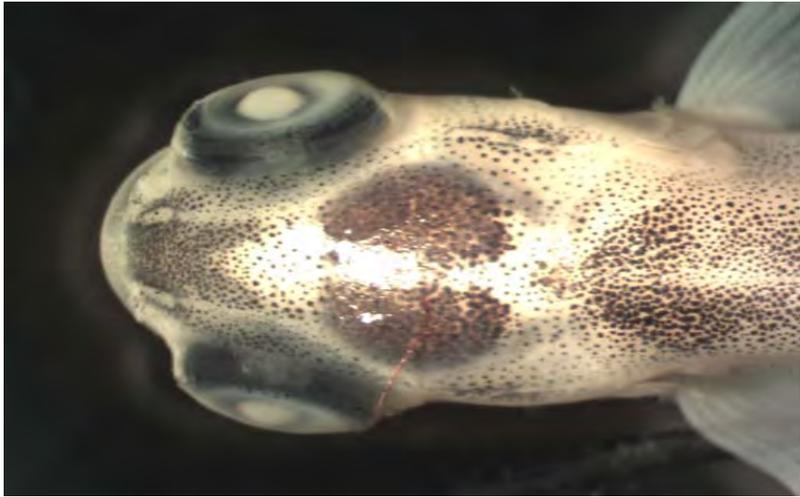
A sample of 50 fish and a sample of 30 fish were scored twice, the same fish for each batch but not necessarily the same order. This sample was characterized by a low incidence of fin deformities (slow development) and a high incidence of jaw deformities and blindness (SC 003 SU). Those cranio-facial traits are difficult to score because they are additive, and subjective as to severity. Thus, the results may be a conservative view of what score replicability should be like for other traits in other samples that are easier to score.

Replicability of frequency of cranio-facial abnormalities was high among assessments at 50 and 52% in the first sample of 50 fish, and identical frequencies of 46.7 % in each assessment for the sample of 30 fish. The cumulative sums of the scores were also quite

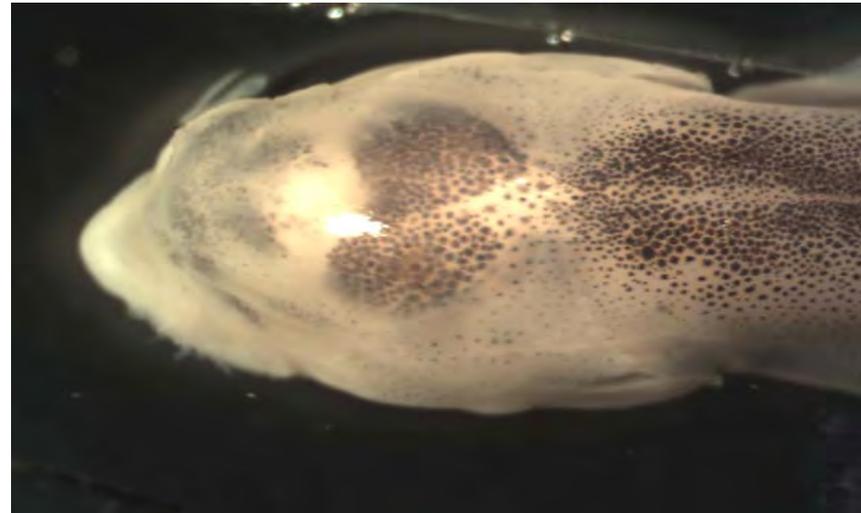
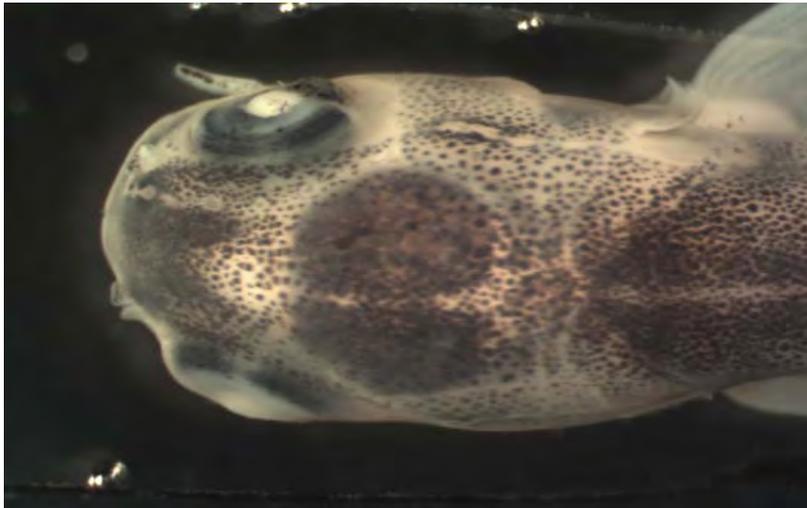
close, but reflecting variability in scoring for all three categories of severity in each sample. Replicability of fin ray development assessments for both frequency and the sum of the scores was identical in both samples.

Below we have included photographs of each of the deformities assessed described above, demonstrating scoring values of 0 – 3 for each of the deformities.

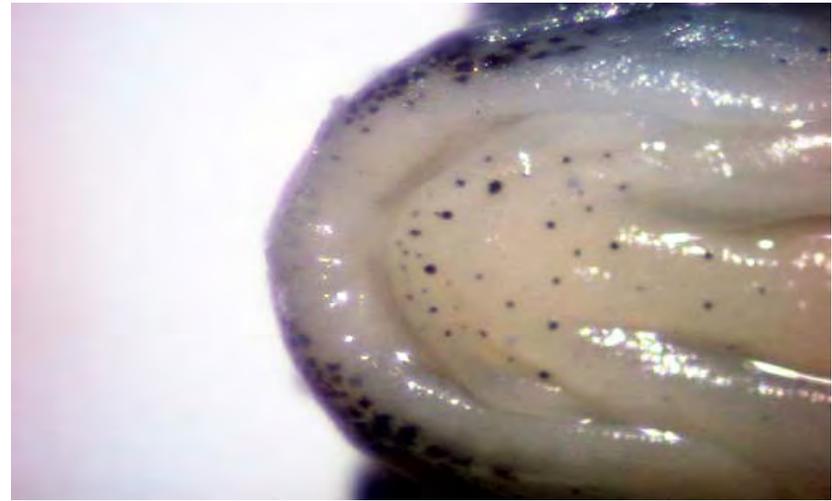
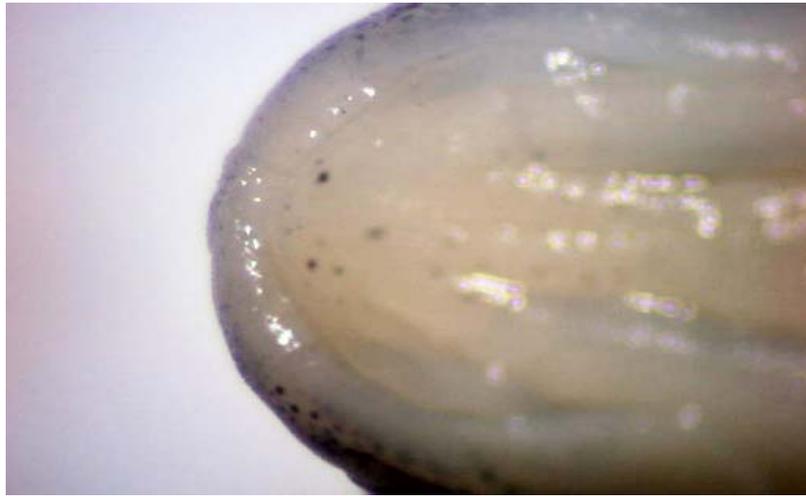
Photos 1 and 2: Example of normal brown trout eyes (left) and an example of a cranio-facial eye deformity with a score of 1 (right).



Photos 3 and 4: Examples of cranio-facial eye deformities with a score of 3 (both).



Photos 1 and 2: Example of a normal brown trout jaw (left) and an example of a cranio-facial jaw deformity with a score of 1 (right).



Photos 3 and 4: Example of a cranio-facial jaw deformity with a score of 2 (left) and 3 (right).



Photos 1 and 2: Example of a healthy brown trout fish (left) and an example of the spinal deformity constriction with a score of 1 (right).



Photo 3: Example of the spinal deformity constriction with a score of 1.



Photos 1 and 2: Example of a healthy brown trout fish (left) and an example of the skeletal deformity kyphosis with a score of 1 (right).



Photos 3 and 4: Example of the skeletal deformity kyphosis with a score of 2 (left) and 3 (right).



Photos 1 and 2: Example of a healthy brown trout fish (left) and an example of the skeletal deformity lordosis with a score of 1 (right).



Photos 3 and 4: Example of the skeletal deformity lordosis with a score of 2 (left) and 3 (right).



Photos 1 and 2: Example of a healthy brown trout fish (left) and an example of the spinal deformity scoliosis with a score of 1 (right).



Photos 3 and 4: Example of the spinal deformity scoliosis with a score of 2 (left) and 3 (right).



Photos 1 and 2: Example of a healthy brown trout fish (left) and an example of a fin deformity with a score of 1 (right).



Photos 3 and 4: Example of a fin deformity with a score of 2 (left) and 3 (right).



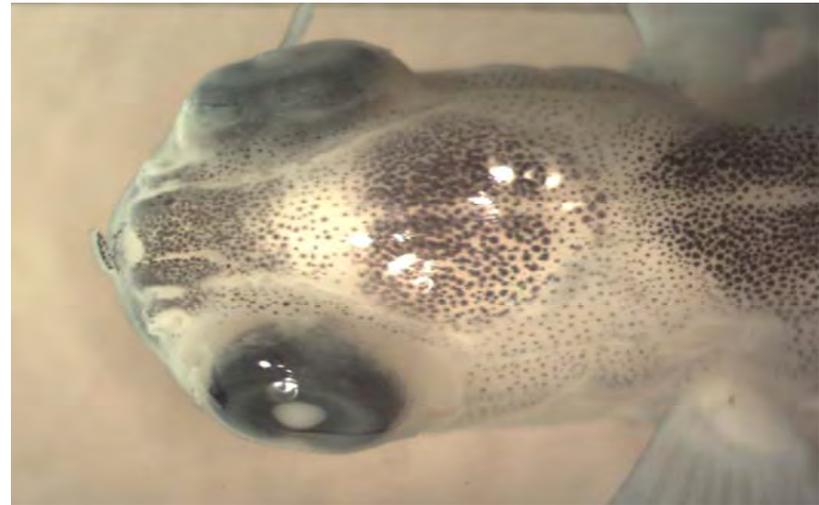
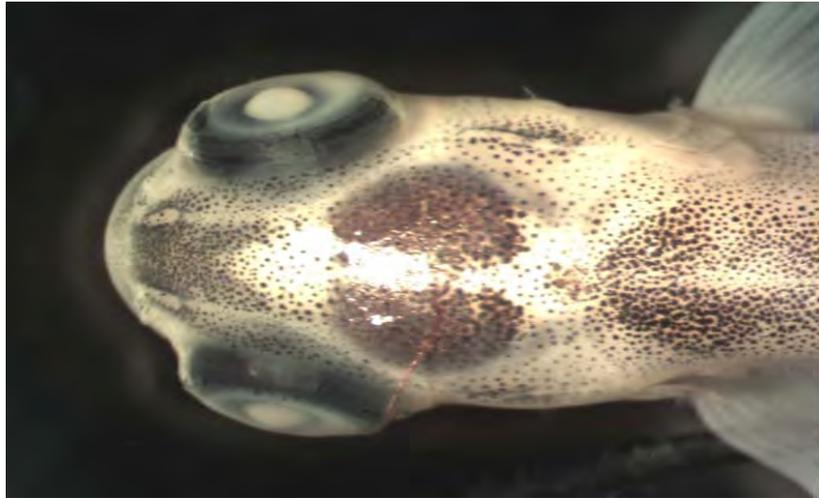
Photos 1 and 2: Example of a healthy brown trout fish (left) and an example of abdominal edema with a score of 1 (right).



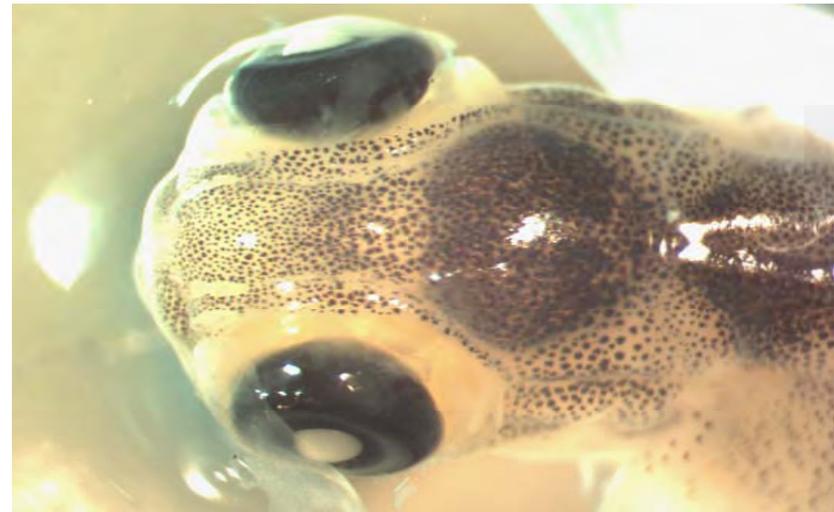
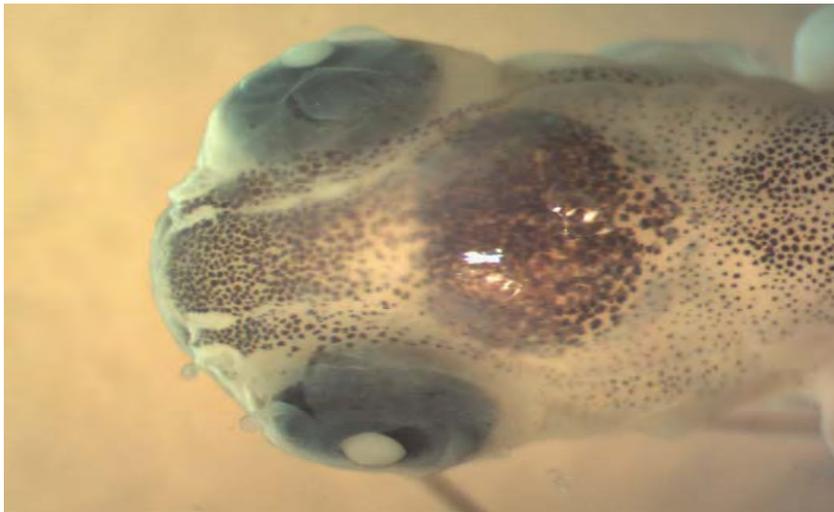
Photos 3 and 4: Examples of abdominal edema with a score of 2 (left) and 3 (right).



Photos 1 and 2: Example of a healthy brown trout fish (left) and an example of cranial edema with a score of 1 (right).



Photos 3 and 4: Example of cranial edema with a score of 2 (left) and 3 (right).



Photos 1 and 2: Examples of brown trout with unusual deformities (both having two heads).



Photos 3 and 4: Examples of unusual deformities.



Deformity assessment of fry preserved after death during the BT parental study.

filename: LSV2C def data.xls

0 (normal) CF = craniofacial deformities
 1 (slight/few) SD = vertebral deformities
 2 (mod/several) FD = fin deformities
 3 (severe/many) ED = edema

Counts

		CF				Grand Total	Total assessed
Location	Field Sample	0	1	2	3		
LSV2C	003	0	3	83	12	98	98
	004	0	6	80	15	101	101
	005	0	14	66	9	89	89
	010	0	16	55	0	71	71
	021	0	2	53	46	101	101

		CF				Grand Total
Location	Field Sample	0	1	2	3	
LSV2C	003	0.0%	3.1%	84.7%	12.24%	100%
	004		5.9%	79.2%	14.85%	100%
	005		15.7%	74.2%	10.11%	100%
	010		22.5%	77.5%	0.00%	100%
	021		2.0%	52.5%	45.54%	100%

		SD				Grand Total	Total assessed
Location	Field Sample	0	1	2	3		
LSV2C	003	0	56	20	6	82	82
	004	0	47	13	4	64	64
	005	0	28	16	40	84	84
	010	0	30	16	7	53	53
	021	0	35	19	8	62	62

		SD				Grand Total
Location	Field Sample	0	1	2	3	
LSV2C	003	0.0%	68.3%	24.4%	7.32%	100%
	004		73.4%	20.3%	6.25%	100%
	005		33.3%	19.0%	47.62%	100%
	010		56.6%	30.2%	13.21%	100%
	021		56.5%	30.6%	12.90%	100%

		FD				Grand Total	Total assessed
Location	Field Sample	0	1	2	3		
LSV2C	003					0	0
	004		1			1	1
	005	0	13	7	35	55	55
	010					0	0
	021	0	9	5	0	14	14

		FD				Grand Total
Location	Field Sample	0	1	2	3	
LSV2C	003	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!
	004	100.0%	0.0%	0.00%		100%
	005	23.6%	12.7%	63.64%		100%
	010	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!	#DIV/0!
	021	64.3%	35.7%	0.00%		100%

		ED				Grand Total	Total assessed
Location	Field Sample	0	1	2	3		
LSV2C	003	0	47	30	7	84	84
	004	0	57	28	6	91	91
	005	0	40	13	5	58	58
	010	0	16	19	10	45	45
	021	0	62	19	1	82	82

		ED				Grand Total
Location	Field Sample	0	1	2	3	
LSV2C	003		56.0%	35.7%	8.33%	100%
	004		62.6%	30.8%	6.59%	100%
	005		69.0%	22.4%	8.62%	100%
	010		35.6%	42.2%	22.22%	100%
	021		75.6%	23.2%	1.22%	100%

Note: scoring criteria were not possible for all organisms due to the poor physical condition of some samples. For these samples, no value was included.

No organisms scored a "0" on any of the different assessments (i.e., CF, SD, FD, ED)

Deformity assessment results for brown trout in reproductive success study

Vaues represent the number of fish (at swimup and at test termination) in each scoring criterion (i.e., 0 - 3).

See below for a definition of scoring criteria.

filename: deformity data.xls

Count of Fish #		Craniofacial Deformities (CF)				Grand Total
Location	Field Sample ID	0	1	2	3	
CC-150	009	136	1	2	3	142
	011	114	150	2		266
	012	191	86	4	1	282
	013	183	31	28	68	310
	015	231	207	5	2	445
	016	20	2		1	23
	017	108	54	1		163
	018	288	193	2	3	486
	020	506	52			558
CC-150 Total		1777	776	44	78	2675
CC-350	006	228	122	22	14	386
	007	102	12	11	6	131
	008	315	8	5	10	338
CC-350 Total		645	142	38	30	855
LSV2C	002	531	13			544
	003		3	83	12	98
	004	63	6	80	15	164
	005	27	27	75	9	138
	008	165	24	5		194
	010		16	55		71
	012	511	39	3	1	554
	016	495	34	1		530
	017	122	16	10	2	150
	019	302	79	8	1	390
	020	257	36	3		296
021	47	13	57	53	170	
LSV2C Total		2520	306	380	93	3299
SC	001	96	14	4	1	115
	002	104	6	1	2	113
	003	174	37	55	36	302
	004	69	26	26	19	140
	005	39	3			42
	006	519	2	6	8	535
	007	119	11	6	1	137
	008	339	12	3	5	359
	SC Total		1459	111	101	72
SPC	001	490	75	2	1	568
	003	448	91	6		545
	005	476	82	2	1	561
	006	475	77	3	1	556
SPC Total		1889	325	13	3	2230
Grand Total		8290	1619	239	194	10342

Craniofacial deformities included shortening of the jaw, snout, and missing or poorly developed eye or eyes, and head shape abnormalities. A slightly shortened lower jaw (≤ 1 lip width) received a 1, a shortened jaw = 2 lip widths or a slightly shortened and slightly disfigured jaw = 2, and a flat lower jaw or much disfigured (non-functional) jaw = 3. An assessment of fish independent of this study revealed that other brown trout of the same size and developmental state did not have the slight deformity that was assessed as CF = 1 for the jaw (J). Thus, the CF = 1 score where the J was concerned were deemed real. A slightly blunted snout (about 50% eye diameter, usually is > than that) = 1, very blunt or flat = 2, deformed or bulbous = 3. Eye deformities were scored as one eye blind or poorly pigmented or poorly developed = 1, both poorly developed = 2, both blind = 3. Skulls that were slightly bulbous ($1/3 >$ normal) = 1, moderately bulbous ($2/3 >$ normal) = 2, and bulbous (1x or > than normal) = 3. Usually factors occurred together so a combination of two "1" conditions = 2, three "1" conditions = 3, or a 1 and a 2 = 3, and so on. For example, a deformed jaw and a blind eye = 2, two blind eyes = 2, but a badly deformed jaw (= 2 alone) plus a blind eye (= 1 alone), = 3.

Deformity assessment results for brown trout in reproductive success study

Vaues represent the number of fish (at swimup and at test termination) in each scoring criterion (i.e., 0 - 3).

See below for a definition of scoring criteria.

filename: deformity data.xls

Count of Fish #		Skeletal Deformities (SD)				
Location	Field Sample ID	0	1	2	3	Grand Total
CC-150	009	109	28	3	2	142
	011	213	50	3		266
	012	237	42	3		282
	013	214	81	11	4	310
	015	402	33	8	2	445
	016	13	10			23
	017	150	11	2		163
	018	353	121	11	1	486
	020	499	44	15		558
CC-150 Total		2190	420	56	9	2675
CC-350	006	198	117	43	28	386
	007	83	22	20	6	131
	008	284	43	7	4	338
CC-350 Total		565	182	70	38	855
LSV2C	002	499	38	7		544
	003		56	20	6	82
	004	20	83	20	4	127
	005	17	44	29	43	133
	008	173	19	2		194
	010		30	16	7	53
	012	235	306	13		554
	016	486	41	3		530
	017	138	10		2	150
	019	341	46	2	1	390
	020	274	17	4	1	296
021	20	71	32	8	131	
LSV2C Total		2203	761	148	72	3184
SC	001	79	28	7	1	115
	002	75	32	3	3	113
	003	260	39	3		302
	004	99	28	6	7	140
	005	25	17			42
	006	486	42	6	1	535
	007	105	23	4	5	137
	008	291	47	8	13	359
	SC Total		1420	256	37	30
SPC	001	493	62	9	4	568
	003	457	64	21	3	545
	005	479	65	12	5	561
	006	488	41	22	5	556
SPC Total		1917	232	64	17	2230
Grand Total		8295	1655	291	101	10342

Skeletal deformities included any deformity of the vertebrae or spines. A slight bend of less than 45 degrees (but > than body width off of straight) or a minor body constriction (e.g. a tight rubberband about the body effect) was given a score of 1, 2 slight bends or constrictions anywhere, or bend of > 45-90 degrees was scored a 2, and multi-directional bends > 90 degrees were given a 3. Bends caused by skeletal deformities were usually detectable from normal bending of the body during preservation (these fish were usually well preserved, very straight) by presence of a slight or greater bump below the surface of the epidermis on the outside of the bend. However, some fish with SD = 1 had just a very slight bend in the range the deformity described but could be due to preservation or the poor condition of the fish. This was sometimes especially true in larger fish, which may be more muscular and undergo stronger contraction during preservation and thus, bend slightly. A score "CF = 1" was a slight deformity, if at all. The scores of SD = 1 involving kyphosis or lordosis were deemed real because that is an unusual preservation deformity. Some samples were re-examined; most fish were very straight so some samples with higher SD scores (e.g., PSU samples) were determined accurate.

Deformity assessment results for brown trout in reproductive success study

Vaues represent the number of fish (at swimup and at test termination) in each scoring criterion (i.e., 0 - 3).

See below for a definition of scoring criteria.

filename: deformity data.xls

Count of Fish #		Fin Deformities (FD)				
Location	Field Sample ID	0	1	2	3	Grand Total
CC-150	009	137	2	1	2	142
	011	266				266
	012	279	1		2	282
	013	287	17	4	2	310
	015	437	3	4	1	445
	016	23				23
	017	162	1			163
	018	483	3			486
	020	549	9			558
CC-150 Total		2623	36	9	7	2675
CC-350	006	325	16	16	29	386
	007	95	10	18	8	131
	008	303	25	7	3	338
CC-350 Total		723	51	41	40	855
LSV2C	002	528	15	1		544
	003					0
	004	48	15	1		64
	005	39	17	11	37	104
	008	194				194
	010					0
	012	544	9	1		554
	016	485	45			530
	017	144	4		2	150
	019	390				390
	020	292			1	296
021	27	51	5		83	
LSV2C Total		2691	156	20	42	2909
SC	001	102	7	5	1	115
	002	103	6	4		113
	003	280	21		1	302
	004	113	10	13	4	140
	005	42				42
	006	501	21	7	6	535
	007	114	11	7	5	137
	008	343	4	3	9	359
	SC Total		1598	80	39	26
SPC	001	542	11	10	5	568
	003	524	8	7	6	545
	005	533	16	4	8	561
	006	529	11	7	9	556
SPC Total		2128	46	28	28	2230
Grand Total		9763	346	125	108	10342

Fin deformities included variation in fin or finfold morphology and a slightly smaller or missing fin (in thin fish, the adipose fin was often absent, indicating fat absorption, not uncommon and scored 1) or one with a bend or incomplete ray development (in older fish) was given a 1, 2 fins damaged or malformed = 2, and > 2 fins malformed or if fins were missing (except adipose) was = 3. Often fins were malformed associated with vertebral deformities that did not permit proper development. Folded finfolds as a result of preservation were not counted.

Deformity assessment results for brown trout in reproductive success study

Vaues represent the number of fish (at swimup and at test termination) in each scoring criterion (i.e., 0 - 3).
See below for a definition of scoring criteria.

filename: deformity data.xls

Count of Fish #		Edema Deformities (ED)				Grand Total
Location	Field Sample ID	0	1	2	3	
CC-150	009	141	1			142
	011	266				266
	012	282				282
	013	308	2			310
	015	445				445
	016	23				23
	017	163				163
	018	485			1	486
	020	558				558
CC-150 Total		2671	3	1		2675
CC-350	006	382	3	1		386
	007	126	3	2		131
	008	337	1			338
CC-350 Total		845	7	3		855
LSV2C	002	541	3			544
	003		47	30	7	84
	004	63	57	28	6	154
	005	42	46	14	5	107
	008	180	6	8		194
	010		16	19	10	45
	012	554				554
	016	530				530
	017	135	9	5	1	150
	019	381	8	1		390
	020	296				296
021	69	62	19	1	151	
LSV2C Total		2791	254	124	30	3199
SC	001	114	1			115
	002	113				113
	003	302				302
	004	139	1			140
	005	42				42
	006	534	1			535
	007	137				137
	008	359				359
SC Total		1740	3			1743
SPC	001	565	3			568
	003	539	4	2		545
	005	558	3			561
	006	553	1	1	1	556
SPC Total		2215	11	3	1	2230
Grand Total		10262	56	22	2	10342

Edema was not originally scheduled for assessment because it was thought sometimes not a teratogenic effect and may be transitory as fish develop. However, it was assessed because it was common in one early sample and not others, and because it was thought a condition that could affect emergence, mobility, and other factors that may limit survival of fish in the wild. Edema was detected by an obvious swelling and fluid buildup, usually abdominally, and ventrally, which often displaced the gut, and was usually clear fluid that was slightly soft when touched with a blunt probe. The yolk, which was present in some quantity in some study specimens, also created some swelling but was typically yellowish, opaque, and small, and hard to the touch in preservation. Slight edema = 1 was for a fish with up to 1X swelling of the normal body width or depth, up to 2x = 2, and > 2x = 3.

Figure 1 Cranio-Facial Frequency

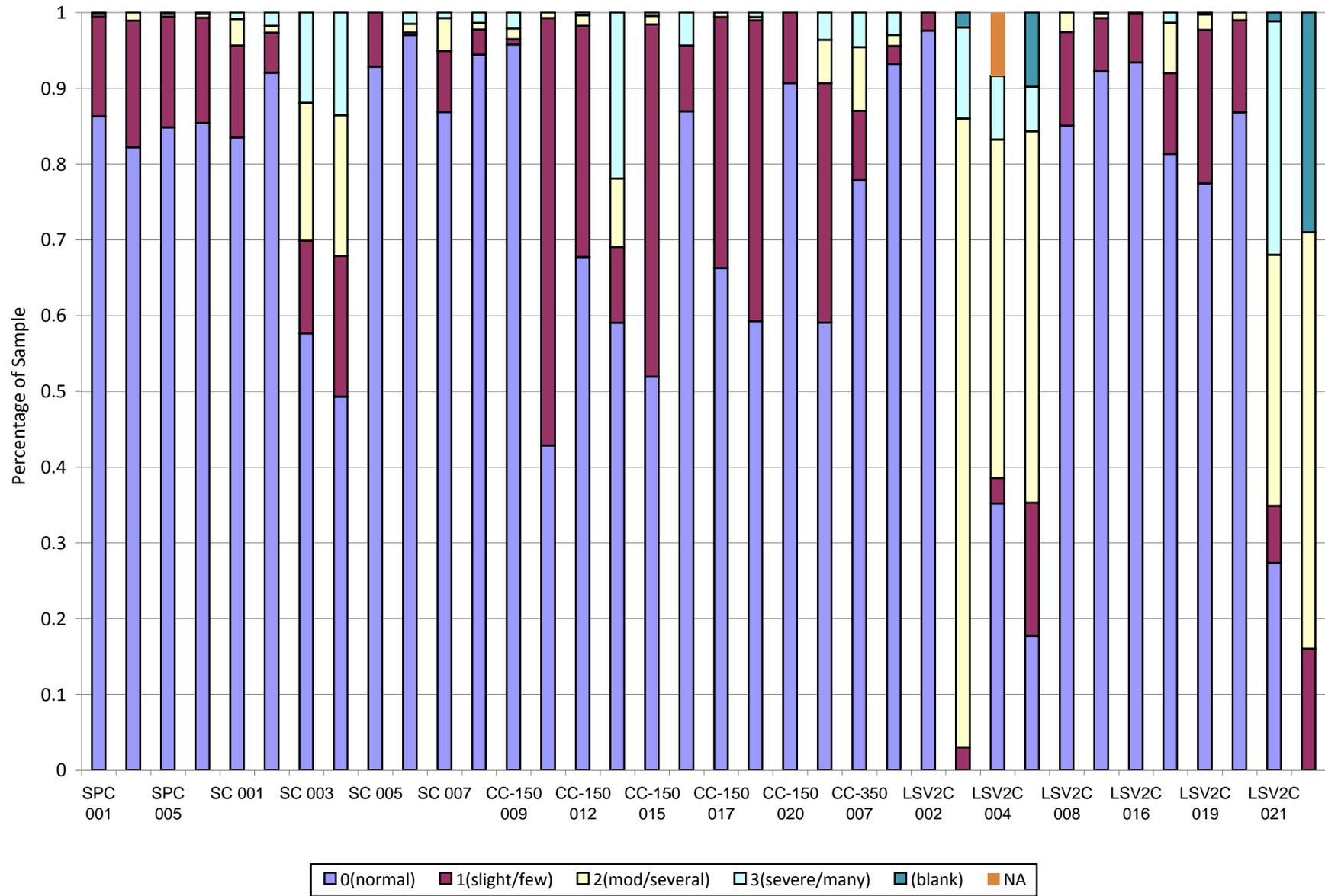


Figure 2 Skeletal Deformity Frequency

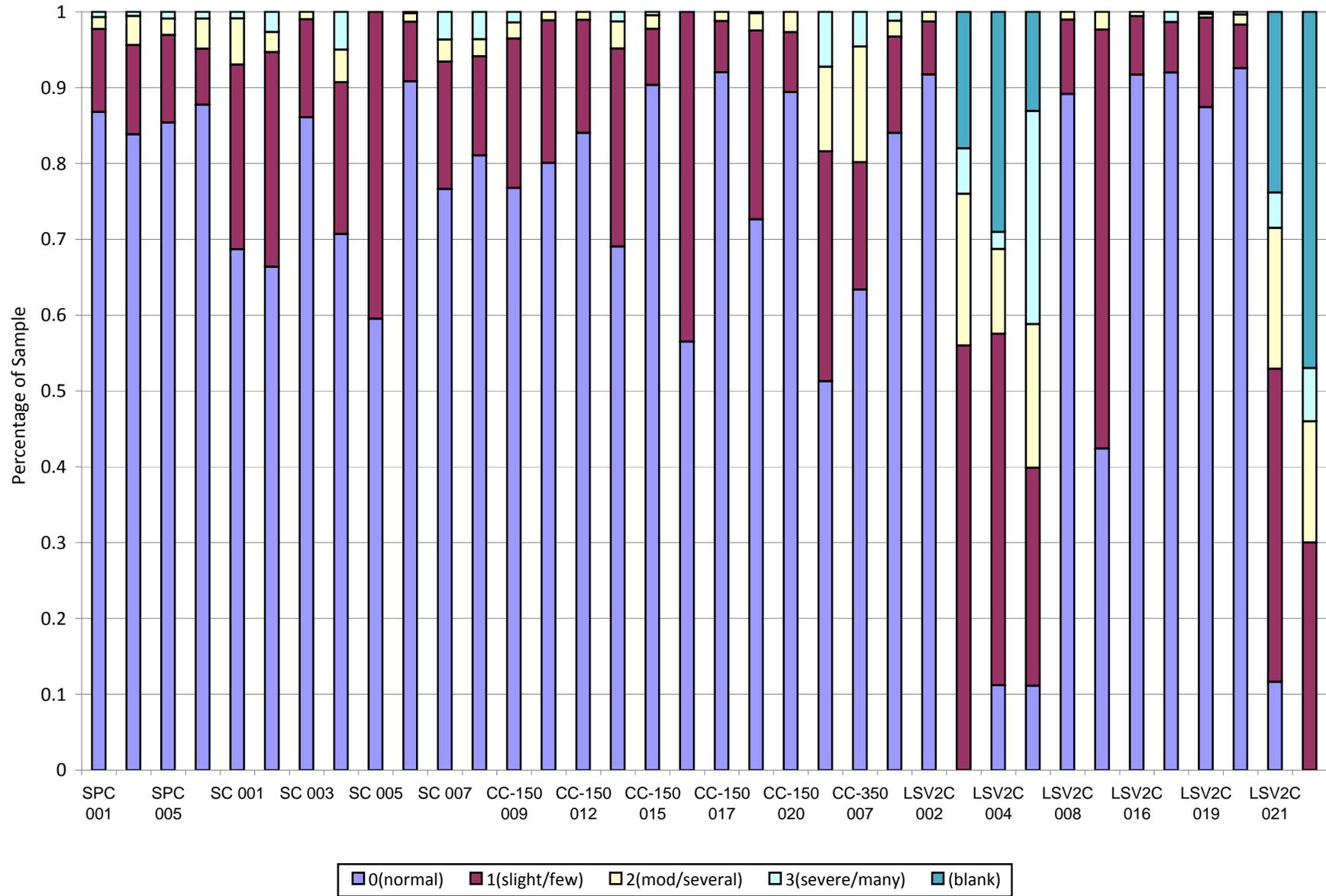


Figure 3 Fin or Finfold Deformity Frequency

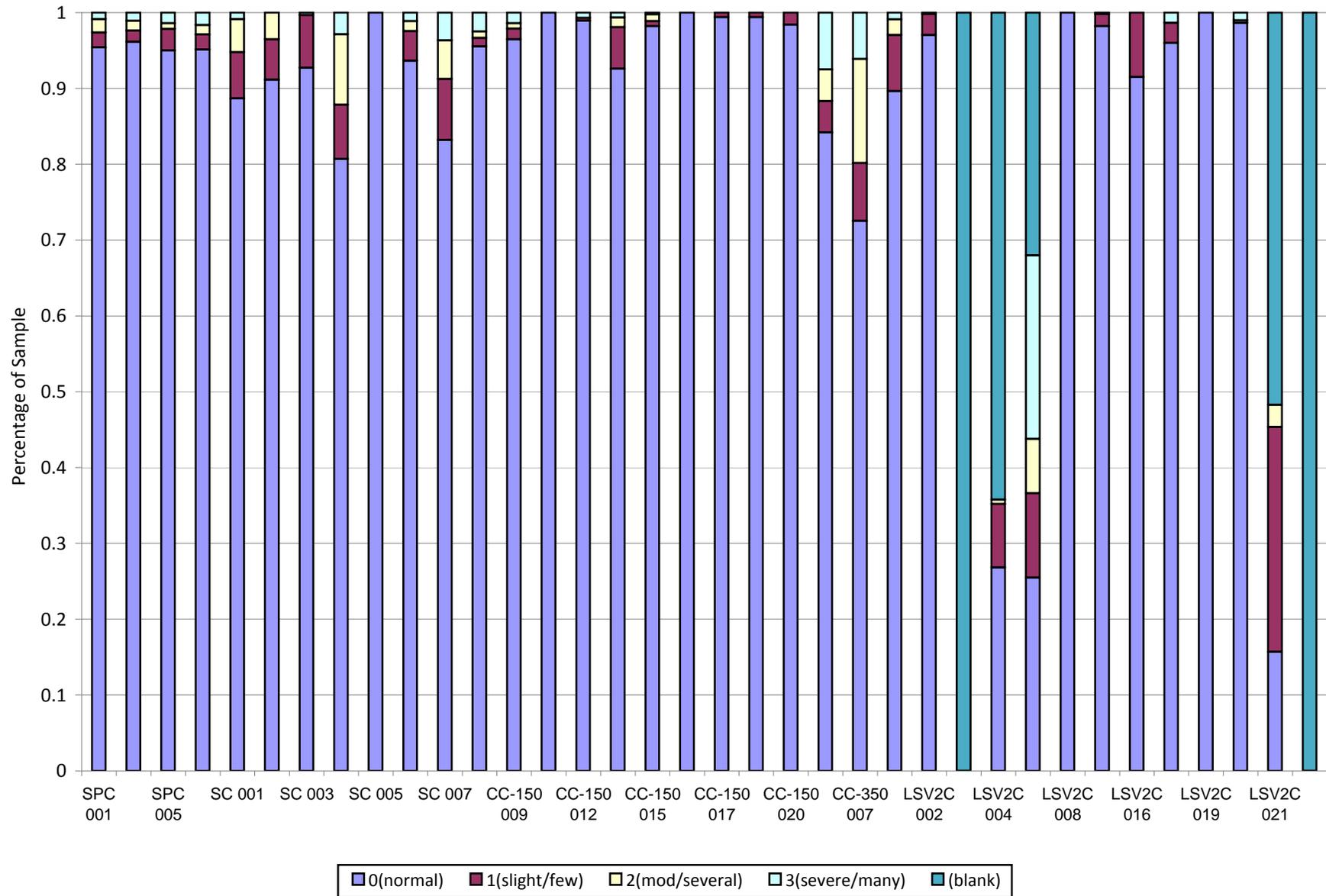
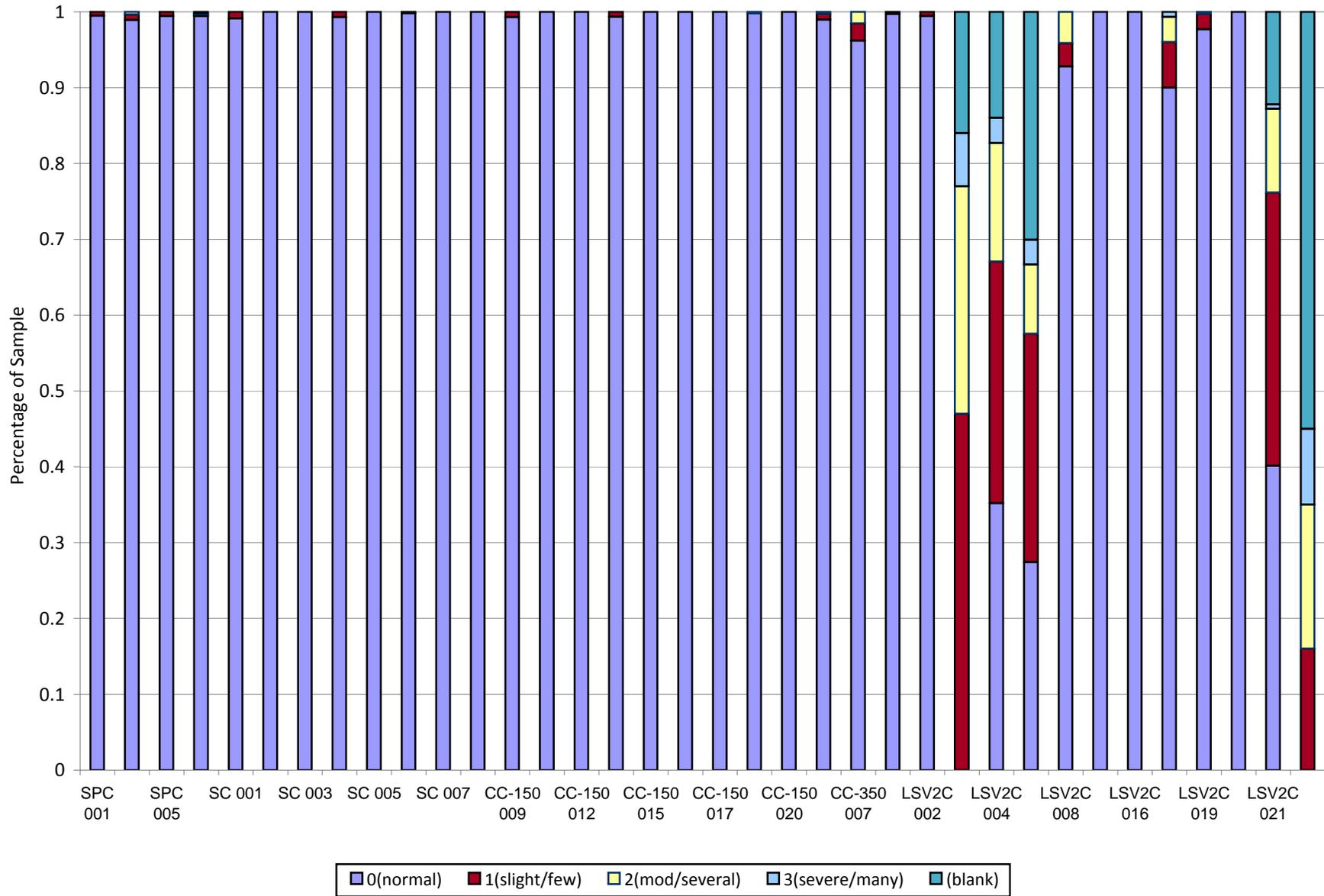


Figure 4 Edematous Tissue Frequency



APPENDIX D
Brown Trout Whole Body and Egg Tissue Selenium Analytical Data

January 23, 2008

Analytical Report for Service Request No: K0711481

Kathy Tegtmeyer
New Fields Environmental
4720 Walnut St., Suite 200
Boulder, CO 80301

RE: Se in Tissue

Dear Kathy:

Enclosed are the results of the samples submitted to our laboratory on December 07, 2007. For your reference, these analyses have been assigned our service request number K0711481.

All analyses were performed according to our laboratory's quality assurance program. Where applicable, the methods cited conform to the Methods Update Rule (effective 4/11/2007), which relates to the use of analytical methods for the drinking water and waste water programs. The test results meet requirements of the NELAC standards. Exceptions are noted in the case narrative report where applicable. All results are intended to be considered in their entirety, and Columbia Analytical Services, Inc. (CAS) is not responsible for use of less than the complete report. Results apply only to the items submitted to the laboratory for analysis and individual items (samples) analyzed, as listed in the report.

Please call if you have any questions. My extension is 3316. You may also contact me via Email at JChristian@caslab.com.

Respectfully submitted,

Columbia Analytical Services, Inc.



Jeff Christian
Laboratory Director

JC/lb

Page 1 of 124

Acronyms

ASTM	American Society for Testing and Materials
A2LA	American Association for Laboratory Accreditation
CARB	California Air Resources Board
CAS Number	Chemical Abstract Service registry Number
CFC	Chlorofluorocarbon
CFU	Colony-Forming Unit
DEC	Department of Environmental Conservation
DEQ	Department of Environmental Quality
DHS	Department of Health Services
DOE	Department of Ecology
DOH	Department of Health
EPA	U. S. Environmental Protection Agency
ELAP	Environmental Laboratory Accreditation Program
GC	Gas Chromatography
GC/MS	Gas Chromatography/Mass Spectrometry
LUFT	Leaking Underground Fuel Tank
M	Modified
MCL	Maximum Contaminant Level is the highest permissible concentration of a substance allowed in drinking water as established by the USEPA.
MDL	Method Detection Limit
MPN	Most Probable Number
MRL	Method Reporting Limit
NA	Not Applicable
NC	Not Calculated
NCASI	National Council of the Paper Industry for Air and Stream Improvement
ND	Not Detected
NIOSH	National Institute for Occupational Safety and Health
PQL	Practical Quantitation Limit
RCRA	Resource Conservation and Recovery Act
SIM	Selected Ion Monitoring
TPH	Total Petroleum Hydrocarbons
tr	Trace level is the concentration of an analyte that is less than the PQL but greater than or equal to the MDL.

Inorganic Data Qualifiers

- * The result is an outlier. See case narrative.
- # The control limit criteria is not applicable. See case narrative.
- B The analyte was found in the associated method blank at a level that is significant relative to the sample result.
- E The result is an estimate amount because the value exceeded the instrument calibration range.
- J The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
 - i The MRL/MDL has been elevated due to a matrix interference.
- X See case narrative.

Metals Data Qualifiers

- # The control limit criteria is not applicable. See case narrative.
- B The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- E The percent difference for the serial dilution was greater than 10%, indicating a possible matrix interference in the sample.
- M The duplicate injection precision was not met.
- N The Matrix Spike sample recovery is not within control limits. See case narrative.
- S The reported value was determined by the Method of Standard Additions (MSA).
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
- W The post-digestion spike for furnace AA analysis is out of control limits, while sample absorbance is less than 50% of spike absorbance.
 - i The MRL/MDL has been elevated due to a matrix interference.
- X See case narrative.
- * The duplicate analysis not within control limits. See case narrative.
- + The correlation coefficient for the MSA is less than 0.995.

Organic Data Qualifiers

- * The result is an outlier. See case narrative.
- # The control limit criteria is not applicable. See case narrative.
- A A tentatively identified compound, a suspected aldol-condensation product.
- B The analyte was found in the associated method blank at a level that is significant relative to the sample result.
- C The analyte was qualitatively confirmed using GC/MS techniques, pattern recognition, or by comparing to historical data.
- D The reported result is from a dilution.
- E The result is an estimate amount because the value exceeded the instrument calibration range.
- J The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- N The result is presumptive. The analyte was tentatively identified, but a confirmation analysis was not performed.
- P The GC or HPLC confirmation criteria was exceeded. The relative percent difference is greater than 40% between the two analytical results (25% for CLP Pesticides).
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
 - i The MRL/MDL has been elevated due to a chromatographic interference.
- X See case narrative.

Additional Petroleum Hydrocarbon Specific Qualifiers

- F The chromatographic fingerprint of the sample matches the elution pattern of the calibration standard.
- L The chromatographic fingerprint of the sample resembles a petroleum product, but the elution pattern indicates the presence of a greater amount of lighter molecular weight constituents than the calibration standard.
- H The chromatographic fingerprint of the sample resembles a petroleum product, but the elution pattern indicates the presence of a greater amount of heavier molecular weight constituents than the calibration standard.
- O The chromatographic fingerprint of the sample resembles an oil, but does not match the calibration standard.
- Y The chromatographic fingerprint of the sample resembles a petroleum product eluting in approximately the correct carbon range, but the elution pattern does not match the calibration standard.
- Z The chromatographic fingerprint does not resemble a petroleum product.

Columbia Analytical Services, Inc.
Kelso, WA
State Certifications, Accreditations, and Licenses

Program	Number
Alaska DEC UST	UST-040
Arizona DHS	AZ0339
Arkansas - DEQ	88-0637
California DHS	2286
Colorado DPHE	-
Florida DOH	E87412
Hawaii DOH	-
Idaho DHW	-
Indiana DOH	C-WA-01
Louisiana DEQ	3016
Louisiana DHH	LA050010
Maine DHS	WA0035
Michigan DEQ	9949
Minnesota DOH	053-999-368
Montana DPHHS	CERT0047
Nevada DEP	WA35
New Jersey DEP	WA005
New Mexico ED	-
North Carolina DWQ	605
Oklahoma DEQ	9801
Oregon - DHS	WA200001
South Carolina DHEC	61002
Utah DOH	COLU
Washington DOE	C1203
Wisconsin DNR	998386840
Wyoming (EPA Region 8)	-



Case Narrative

COLUMBIA ANALYTICAL SERVICES, INC.

Client: New Fields Environmental
Project: Tissue - Se
Sample Matrix: Tissue

Service Request No.: K0711481
Date Received: 12/7/07

CASE NARRATIVE

All analyses were performed consistent with the quality assurance program of Columbia Analytical Services, Inc. (CAS). This report contains analytical results for samples designated for Tier III validation deliverables including summary forms and all of the associated raw data for each of the analyses. When appropriate to the method, method blank results have been reported with each analytical test.

Sample Receipt

Tissue samples were received for analysis at Columbia Analytical Services on 12/7/07. The samples were received in good condition and consistent with the accompanying chain of custody form. The samples were stored frozen at -20°C upon receipt at the laboratory.

Total Metals

General Comments:

The samples were homogenized, then freeze-dried to determine moisture and to allow complete homogenization of the dry material. The dried material was milled to a fine meal, and then sub-sampled for digestion. A thorough digestion was performed prior to instrumental analysis to convert all Selenium species to Selenate. Prior to hydride formation, the valence was adjusted by reduction to Selenite.

No anomalies associated with the analysis of these samples were observed.

Approved by _____

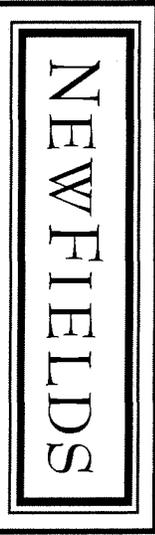


Date

1/22/08

**Chain of Custody
Documentation**

Chain of Custody



Project Contact
Courier/Airbill:

Sean Covington/Kathy Tegtmeyer
PO 0442-004-900.70

Shipped to:

Columbia Analytical Services, Inc.
1317 South 13th Ave
Kelso, WA 98626

Telephone:

(360) 430-7733

COC # 40006

COC #:

4720 Walnut St, Suite 200
Boulder, CO 80301
Phone: 303-442-0267
Fax: 303-442-3679

Sample ID	Sample Date	Sample Time	Matrix	Tox/ Diss	Analysis	Preservative	Lab QC	Comments
SM1007-cc1a-FT0012	10/30/2007		Fish Tissue		Selenium, % Solids	<i>PCB</i>		
SM1007-cc150-FT0013	10/30/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0014	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0015	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0016	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0017	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0018	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0019	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0020	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0021	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0022	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0023	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0024	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0025	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0026	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0027	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0028	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0029	11/14/2007		Fish Tissue		Selenium, % Solids			

Sampler Signature: *[Handwritten Signature]*

LAB USE ONLY - Sample condition on Receipt

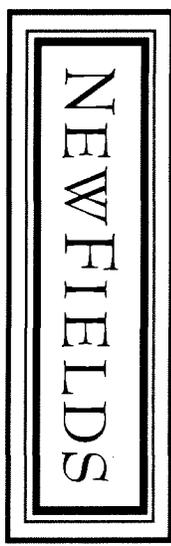
Relinquished by *[Handwritten Signature]* Date/Time *12/6/07 0900* Received by *[Handwritten Signature]* Date/Time *CAS 12/7/07 0920*

2810704

Chain of Custody Page 2 of 3

Project Contact: Sean Covington/Kathy Tegtmeyer PO 0442-004-900.70
 Courier/Airbill: Columbia Analytical Services, Inc.

Shipped to: 1317 South 13th Ave
 Kelso, WA 98626
 Telephone: (360) 430-7733



4720 Walnut St, Suite 200
 Boulder, CO 80301
 Phone: 303-442-0267
 Fax: 303-442-3679

COC #:

Sample ID	Sample Date	Sample Time	Matrix	Tox/Dis	Analysis	Preservative	Lab QC	Comments
SM1107-LSV2c-FT0030	11/14/2007		Fish Tissue		Selenium, % Solids	120		
SM1107-LSV2c-FT0031	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0032	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-LSV2c-FT0033	11/14/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC150-FT0034	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC150-FT0035	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC150-FT0036	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC150-FT0037	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC150-FT0038	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC150-FT0039	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC150-FT0040	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC150-FT0041	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC150-FT0042	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC350-FT0043	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC350-FT0044	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1107-CC350-FT0045	11/15/2007		Fish Tissue		Selenium, % Solids			
SM1007-cc1a-FT0046	10/30/2007		Eggs		Selenium, % Solids			
SM1007-cc150-FT0047	10/30/2007		Eggs		Selenium, % Solids			

Sampler Signature: *NSM*

LAB USE ONLY - Sample condition on Receipt

Relinquished by: <i>NSM</i>	Date/Time: <i>12/6/07 09:00</i>	Received by: <i>[Signature]</i>	Date/Time: <i>12/7/07 09:20</i>
-----------------------------	---------------------------------	---------------------------------	---------------------------------

Chain of Custody Page 3 of 3

Project Contact: Sean Covington/Kathy Tegtmeyer PO 0442-004-900.70
 Courier/Airbill: Columbia Analytical Services, Inc.

Shipped to: 1317 South 13th Ave
 Kelso, WA 98626
 Telephone: (360) 430-7733



4720 Walnut St, Suite 200
 Boulder, CO 80301
 Phone: 303-442-0267
 Fax: 303-442-3679

COC #:

Sample ID	Sample Date	Sample Time	Matrix	Tot/ Diss	Analysis	Preservative	Lab QC	Comments
SM1107-LSV2c-FT0048	11/14/2007		Eggs		Selenium, % Solids	None		
SM1107-LSV2c-FT0049	11/14/2007		Eggs		Selenium, % Solids	↓		

Total Number of Containers: 38 ^{Samples} Individual Lines Reflect Single Containers, Except for Aqueous Analyses Assigned as Laboratory QC

Sampler Signature:

Relinquished by: Date/Time: 12/6/07 1600
 Received by: Date/Time: CAS 12/7/07 0920

LAB USE ONLY - Sample condition on Receipt

**Columbia Analytical Services, Inc.
Cooler Receipt and Preservation Form**

PC JC

Client / Project: ENSR Service Request **K07** 11481

Received: 12/7/07 Opened: 12/7/07 By: M

1. Samples were received via? US Mail **Fed Ex** UPS DHL GH GS PDX Courier Hand Delivered
2. Samples were received in: (circle) **Cooler** Box Envelope Other 1 front NA
3. Were custody seals on coolers? NA Y N If yes, how many and where? _____
If present, were custody seals intact? Y N If present, were they signed and dated? Y N
4. Is shipper's air-bill filed? If not, record air-bill number: _____ NA Y N
5. Temperature of cooler(s) upon receipt (°C): -6.0 (Dry Ice)
Temperature Blank (°C): —
6. If applicable, list Chain of Custody Numbers: —
7. Were custody papers properly filled out (ink, signed, etc.)? NA Y N
8. Packing material used. Inserts Bubble Wrap Gel Packs Wet Ice Sleeves Other Baggies
9. Did all bottles arrive in good condition (unbroken)? Indicate in the table below. NA Y N
10. Were all sample labels complete (i.e analysis, preservation, etc.)? Y N
11. Did all sample labels and tags agree with custody papers? Indicate in the table below Y N
12. Were the correct types of bottles used for the tests indicated? NA Y N
13. Were all of the preserved bottles received at the lab with the appropriate pH? Indicate in the table below NA Y N
14. Were VOA vials and 1631 Mercury bottles checked for absence of air bubbles? Indicate in the table below. NA Y N
15. Are CWA Microbiology samples received with >1/2 the 24hr. hold time remaining from collection? NA Y N
16. Was C12/Res negative? NA Y N

Sample ID on Bottle	Sample ID on COC	Sample ID on Bottle	Sample ID on COC

Sample ID	Bottle Count	Bottle Type	Out of Temp	Head-space	Broken	pH	Reagent	Volume added	Reagent Lot Number	Initials

Additional Notes, Discrepancies, & Resolutions: _____

Total Solids

COLUMBIA ANALYTICAL SERVICES, INC.

Analytical Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0711481
Date Collected: 10/30-11/14/07
Date Received: 12/7/07

Solids, Total

Prep Method: NONE
 Analysis Method: Freeze Dry
 Test Notes:

Units: PERCENT
 Basis: Wet

Sample Name	Lab Code	Date Analyzed	Result	Result Notes
SM1007-cc1a-FT0012	K0711481-001	12/18/07	24.8	
SM1007-cc150-FT0013	K0711481-002	12/18/07	24.8	
SM1107-LSV2c-FT0014	K0711481-003	12/18/07	21.9	
SM1107-LSV2c-FT0015	K0711481-004	12/18/07	21.5	
SM1107-LSV2c-FT0016	K0711481-005	12/18/07	22.1	
SM1107-LSV2c-FT0017	K0711481-006	12/18/07	21.8	
SM1107-LSV2c-FT0018	K0711481-007	12/18/07	21.8	
SM1107-LSV2c-FT0019	K0711481-008	12/18/07	22.8	
SM1107-LSV2c-FT0020	K0711481-009	12/18/07	23.1	
SM1107-LSV2c-FT0021	K0711481-010	12/18/07	22.5	
SM1107-LSV2c-FT0022	K0711481-011	12/18/07	22.0	
SM1107-LSV2c-FT0023	K0711481-012	12/18/07	21.6	
SM1107-LSV2c-FT0024	K0711481-013	12/18/07	24.2	
SM1107-LSV2c-FT0025	K0711481-014	12/18/07	23.2	
SM1107-LSV2c-FT0026	K0711481-015	12/18/07	22.3	
SM1107-LSV2c-FT0027	K0711481-016	12/18/07	20.3	
SM1107-LSV2c-FT0028	K0711481-017	12/18/07	23.6	
SM1107-LSV2c-FT0029	K0711481-018	12/18/07	22.1	
SM1107-LSV2c-FT0030	K0711481-019	12/18/07	21.8	
SM1107-LSV2c-FT0031	K0711481-020	12/18/07	23.2	

COLUMBIA ANALYTICAL SERVICES, INC.

Analytical Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0711481
Date Collected: 10/30-11/15/07
Date Received: 12/07/07

Solids, Total

Prep Method: NONE
 Analysis Method: Freeze Dry
 Test Notes:

Units: PERCENT
 Basis: Wet

Sample Name	Lab Code	Date Analyzed	Result	Result Notes
SM1107-LSV2c-FT0032	K0711481-021	12/18/07	21.1	
SM1107-LSV2c-FT0033	K0711481-022	12/18/07	23.0	
SM1107-CC150-FT0034	K0711481-023	12/18/07	22.6	
SM1107-CC150-FT0035	K0711481-024	12/18/07	23.9	
SM1107-CC150-FT0036	K0711481-025	12/18/07	21.7	
SM1107-CC150-FT0037	K0711481-026	12/18/07	23.1	
SM1107-CC150-FT0038	K0711481-027	12/18/07	22.7	
SM1107-CC150-FT0039	K0711481-028	12/18/07	23.6	
SM1107-CC150-FT0040	K0711481-029	12/18/07	22.8	
SM1107-CC150-FT0041	K0711481-030	12/18/07	23.2	
SM1107-CC150-FT0042	K0711481-031	12/18/07	23.8	
SM1107-CC350-FT0043	K0711481-032	12/18/07	22.5	
SM1107-CC350-FT0044	K0711481-033	12/18/07	23.1	
SM1107-CC350-FT0045	K0711481-034	12/18/07	23.1	
SM1007-cc1a-FT0046	K0711481-035	12/18/07	37.6	
SM1007-cc150-FT0047	K0711481-036	12/18/07	37.4	
SM1107-LSV2c-FT0048	K0711481-037	12/18/07	35.7	
SM1107-LSV2c-FT0049	K0711481-038	12/18/07	36.6	

COLUMBIA ANALYTICAL SERVICES, INC.

QA/QC Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0711481
Date Collected: 10/30/07
Date Received: 12/7/07
Date Extracted: NA
Date Analyzed: 12/18/07

Duplicate Summary
Total Metals

Sample Name: SM1007-cc1a-FT0012
Lab Code: K0711481-001D
Test Notes:

Units: PERCENT
Basis: Wet

Analyte	Prep Method	Analysis Method	Sample Result	Duplicate Sample Result	Average	Relative Percent Difference	Result Notes
Solids, Total	NA	Freeze Dry	24.8	24.9	24.8	<1	

COLUMBIA ANALYTICAL SERVICES, INC.

QA/QC Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0711481
Date Collected: 11/14/07
Date Received: 12/7/07
Date Extracted: NA
Date Analyzed: 12/18/07

Duplicate Summary
Total Metals

Sample Name: SM1107-LSV2c-FT0032
Lab Code: K0711481-021D
Test Notes:

Units: PERCENT
Basis: Wet

Analyte	Prep Method	Analysis Method	Sample Result	Duplicate Sample Result	Average	Relative Percent Difference	Result Notes
Solids, Total	NA	Freeze Dry	21.1	21.2	21.2	<1	

Service Request #:

K0711481

Analytical Batch

Analysis For:

Freeze Dried Solids

KA0628221

Lab Code	Wet Weight (g)	Tare (g)	Tare + Dry Wt.(g)	Dry Weight (g)	% Total Solids
K0711481-01	30.186	78.713	86.206	7.493	24.8
- 01b	32.467	78.921	87.001	8.08	24.9
- 02	37.444	78.723	87.996	9.273	24.8
- 03	29.822	78.753	85.294	6.541	21.9
- 04	28.337	79.158	85.237	6.079	21.5
- 05	30.778	78.876	85.689	6.813	22.1
- 06	32.211	79.460	86.491	7.031	21.8
- 07	34.266	78.594	86.049	7.455	21.8
- 08	33.956	78.963	86.717	7.754	22.8
- 09	35.812	79.412	87.700	8.288	23.1
- 10	28.284	78.290	84.641	6.351	22.5
- 11	28.990	78.280	84.661	6.381	22.0
- 12	33.570	78.608	85.859	7.251	21.6
- 13	33.902	79.190	87.387	8.197	24.2
- 14	47.593	78.729	89.781	11.052	23.2
- 15	34.997	78.714	86.526	7.806	22.3
- 16	32.483	78.909	85.494	6.585	20.3
- 17	29.263	78.646	85.557	6.911	23.6
- 18	34.264	78.984	86.548	7.564	22.1
- 19	30.841	78.977	85.690	6.713	21.8
- 20	28.779	78.756	85.443	6.687	23.2
- 21	36.282	78.444	86.112	7.668	21.1
- 21b	33.294	78.652	85.695	7.043	21.2
- 22	37.247	78.545	87.101	8.556	23.0
- 23	34.495	79.450	87.254	7.804	22.6
- 24	30.931	79.094	86.481	7.387	23.9
- 25	31.641	78.946	85.798	6.857	21.7

Time In: 11:00a.m. 12/19/07 Time Out: 9:30a.m. 12/21/07

Comments:

Balance 21-B

\bar{x} =

RPD =

Analyst: <u>Angela Black</u>	Date: <u>12/18/07</u>
Reviewed By: <u>[Signature]</u>	Date: <u>1/2/07</u>

FREZ-DRY

Service Request #: K0711481

Analysis For: Freeze Dried Solids

Lab Code	Wet Weight (g)	Tare (g)	Tare + Dry Wt.(g)	Dry Weight (g)	% Total Solids
K0711481-26	31.278	79.200	86.435	7.235	23.1
- 27	28.103	79.015	85.383	6.368	22.7
- 28	55.995	79.197	92.380	13.189	23.4
- 29	29.973	78.387	85.224	6.837	22.8
- 30	34.664	79.229	87.263	8.034	23.2
- 31	35.317	79.119	87.535	8.416	23.8
- 32	34.918	78.697	86.559	7.862	22.5
- 33	37.535	78.483	87.144	8.661	23.1
- 34	30.933	79.299	86.434	7.135	23.1
- 35	19.819	78.872	80.321	7.449	37.6
- 36	26.681	79.117	89.107	9.99	37.4
- 37	31.659	79.083	90.401	11.318	35.7
- 38	25.621	79.332	88.705	9.373	36.6
<i>LAB 12/18/07</i>					

Time In: 11:00a.m. 12/19/07 Time Out: 9:30a.m. 12/21/07

Comments: Balanie 21-B

\bar{x} = RPD =

Analyst: <u>Angela Brack</u>	Date: <u>2/18/07</u>
Reviewed By: <u>[Signature]</u>	Date: <u>1/21/07</u>

Sample Number(s): As Listed Service Request Number(s): K0711481

ISSUE COMPOSITION DATA

Laboratory ID	Weight (g)	Tare (g)		Matrix	Length
K0711481-01	272.76	279.58		whole body FSM	
- 02	358.05	281.05			
- 03	308.05	276.46			
- 04	220.77	276.79			
- 05	217.35	282.46			
- 06	218.83	279.83			
- 07	216.64	278.64			
- 08	334.56	279.38			
- 09	243.40	276.88			
- 10	193.82	278.08			
- 11	278.27	276.75			
- 12	346.76	283.03			
- 13	340.97	276.85			
- 14	396.10	277.26			
- 15	376.69 276.79	278.48	278.57 AB 12/19/07		
- 16	250.09	277.02			
- 17	197.54	276.82			
- 18	274.87	278.39			
- 19	416.32	281.68			
- 20	282.09	276.40			
- 21	198.44	276.52			
- 22	246.07	276.76			
- 23	519.87	276.38			
- 24	303.07	276.76			
- 25	231.80	282.70			

car
1

Comments: K0711481-026 had a tag identifier attached, clipped it off and put back in bag. balance 23

Analyst: Angela Brack / Eric P. Witt Date: 12/18/07
 Reviewed: [Signature] Date: 1/2/07

Sample Number(s):

As Listed

Service Request Number(s):

TISSUE COMPOSITION DATA

Laboratory ID	Weight (g)	Tare (g)	Matrix	Length	
40711481-26	282.90	277.00	Whole body fish		
- 27	212.53	277.51			
- 28	467.62	282.51			
- 29	149.99	276.78			
- 30	223.62	277.04			
- 31	238.32	276.65			
- 32	373.09	276.53			
- 33	273.27 ^{331.04}	278.27			
- 34	253.68	276.35			
- 35	20.15	276.47			
- 36	92.68	276.22		eggs	
- 37	87.11	279.51			
- 38	107.89	276.87			
- 15	196.68	275.87			
(AB 12/18/07)					

Comments:

Balance - 23

Analyst: Angela Mack / Eric R. Nth

Reviewed: *[Signature]*

Date: 12/18/07

Date: 11/2/07

Metals

Columbia Analytical Services

- Cover Page -
INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental
Project Name: Se in Tissue
Project No.:

Service Request: K0711481

Sample Name:

Lab Code:

SM1007-cc1a-FT0012	K0711481-001
SM1007-cc1a-FT0012D	K0711481-001D
SM1007-cc1a-FT0012S	K0711481-001S
SM1007-cc150-FT0013	K0711481-002
SM1107-LSV2c-FT0014	K0711481-003
SM1107-LSV2c-FT0015	K0711481-004
SM1107-LSV2c-FT0016	K0711481-005
SM1107-LSV2c-FT0017	K0711481-006
SM1107-LSV2c-FT0018	K0711481-007
SM1107-LSV2c-FT0019	K0711481-008
SM1107-LSV2c-FT0020	K0711481-009
SM1107-LSV2c-FT0021	K0711481-010
SM1107-LSV2c-FT0022	K0711481-011
SM1107-LSV2c-FT0023	K0711481-012
SM1107-LSV2c-FT0024	K0711481-013
SM1107-LSV2c-FT0025	K0711481-014
SM1107-LSV2c-FT0026	K0711481-015
SM1107-LSV2c-FT0027	K0711481-016
SM1107-LSV2c-FT0028	K0711481-017
SM1107-LSV2c-FT0029	K0711481-018
SM1107-LSV2c-FT0030	K0711481-019
SM1107-LSV2c-FT0031	K0711481-020
SM1107-LSV2c-FT0032	K0711481-021
SM1107-LSV2c-FT0032D	K0711481-021D
SM1107-LSV2c-FT0032S	K0711481-021S
SM1107-LSV2c-FT0033	K0711481-022
SM1107-CC150-FT0034	K0711481-023
SM1107-CC150-FT0035	K0711481-024
SM1107-CC150-FT0036	K0711481-025
SM1107-CC150-FT0037	K0711481-026
SM1107-CC150-FT0038	K0711481-027
SM1107-CC150-FT0039	K0711481-028
SM1107-CC150-FT0040	K0711481-029
SM1107-CC150-FT0041	K0711481-030
SM1107-CC150-FT0042	K0711481-031
SM1107-CC350-FT0043	K0711481-032
SM1107-CC350-FT0044	K0711481-033
SM1107-CC350-FT0045	K0711481-034
SM1007-cc1a-FT0046	K0711481-035

Comments:

Approved By:



Date:

1/22/03

Columbia Analytical Services

- Cover Page -
INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental
Project Name: Se in Tissue
Project No.:

Service Request: K0711481

Sample Name:

SM1007-cc150-FT0047

SM1107-LSV2c-FT0048

SM1107-LSV2c-FT0049

Method Blank

Method Blank

Lab Code:

K0711481-036

K0711481-037

K0711481-038

K0711481-MB1

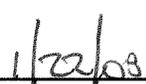
K0711481-MB2

Comments:

Approved By: _____



Date: _____



Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium	10.0	9.94	99	10.0	9.71	97	9.30	93	7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium				10.0	10.04	100	10.21	102	7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium				10.0	10.00	100			7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium	10.0	9.47	95	10.0	9.61	96	9.61	96	7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium				10.0	9.71	97	9.85	98	7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium				10.0	9.68	97			7742

Metals

- 2b -

CRDL STANDARD FOR AA AND ICP

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Concentration Units: ug/L

Analyte	CRDL Standard for AA			CRDL Standard for ICP				
	True	Found	%R	Initial		Final		
	True	Found	%R	True	Found	%R	Found	%R
Selenium	0.5	0.59	118.0					

Metals

- 2b -

CRDL STANDARD FOR AA AND ICP

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Concentration Units: ug/L

Analyte	CRDL Standard for AA			CRDL Standard for ICP				
	True	Found	%R	Initial		Final		
				True	Found	%R	Found	%R
Selenium	0.5	0.64	128.0					

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)		Continuing Calibration Blank (ug/L)						Method
	1	C	1	C	2	C	3	C	
Selenium	0.1	U	0.1	U	0.1	U	0.1	U	7742

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)		Continuing Calibration Blank (ug/L)						Method
		C	1	C	2	C	3	C	
Selenium			0.1	U	0.1	U			7742

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)		Continuing Calibration Blank (ug/L)						Method
		C	1	C	2	C	3	C	
Selenium	0.2	B	0.1	U	0.1	B	0.2	B	7742

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)		Continuing Calibration Blank (ug/L)						Method
		C	1	C	2	C	3	C	
Selenium			0.2	B	0.2	B			7742

Metals

- 5A -

SPIKE SAMPLE RECOVERY

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Units: MG/KG

Project Name: Se in Tissue

Basis: DRY

Matrix: TISSUE

Sample Name: SM1007-ccl1a-FT0012S

Lab Code: K0711481-001S

Analyte	Control Limit %R	Spike Result	C	Sample Result	C	Spike Added	%R	Q	Method
Selenium	65 - 124	29.7		8.7		24.27	86.5		7742

An empty field in the Control Limit column indicates the control limit is not applicable

Metals

- 5A -

SPIKE SAMPLE RECOVERY

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Units: MG/KG

Project Name: Se in Tissue

Basis: DRY

Matrix: TISSUE

Sample Name: SM1107-LSV2c-FT0032S

Lab Code: K0711481-021S

Analyte	Control Limit %R	Spike Result	C	Sample Result	C	Spike Added	%R	Q	Method
Selenium	65 - 124	39.4		11.3		25.00	112.4		7742

An empty field in the Control Limit column indicates the control limit is not applicable

Metals

- 6 -

DUPLICATES

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Units: MG/KG

Project Name: Se in Tissue

Basis: DRY

Matrix: TISSUE

Sample Name: SM1007-cc1a-FT0012D

Lab Code: K0711481-001D

Analyte	Control Limit	Sample (S)	C	Duplicate (D)	C	RPD	Q	Method
Selenium	30	8.7		8.1		7.1		7742

An empty field in the Control Limit column indicates the control limit is not applicable.

Metals

- 6 -

DUPLICATES

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Units: MG/KG

Project Name: Se in Tissue

Basis: DRY

Matrix: TISSUE

Sample Name: SM1107-LSV2c-FT0032D

Lab Code: K0711481-021D

Analyte	Control Limit	Sample (S)	C	Duplicate (D)	C	RPD	Q	Method
Selenium	30	11.3		11.4		0.9		7742

An empty field in the Control Limit column indicates the control limit is not applicable.

Metals

- 7 -

LABORATORY CONTROL SAMPLE

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Aqueous LCS Source:

Solid LCS Source: NRCC DOLT 3

Analyte	Aqueous (ug/L)			Solid (mg/kg)					
	True	Found	%R	True	Found	C	Limits	%R	
Selenium				7.06	8.1		5.26	9.05	114.7

Metals

- 7 -

LABORATORY CONTROL SAMPLE

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Aqueous LCS Source:

Solid LCS Source: NRCC DOLT 3

Analyte	Aqueous (ug/L)			Solid (mg/kg)				
	True	Found	%R	True	Found	C	Limits	%R
Selenium				7.06	6.6		5.26 9.05	93.5

Metals

- 10 -

DETECTION LIMITS

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

ICP/ICP-MS ID #:

GFAA ID #: K-FLAA-02

AA ID #:

Analyte	Wave-length (nm)	Back-ground	MRL ug/L	MDL ug/L	M
Selenium			0.5	0.1	F

Comments: _____

Metals
-13-
PREPARATION LOG

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Method: F

Sample ID	Preparation Date	Initial Volume	Final Volume (mL)
K0711481-001	12/28/07	0.2180	20.0
K0711481-001D	12/28/07	0.2080	20.0
K0711481-001S	12/28/07	0.2060	20.0
K0711481-002	12/28/07	0.2080	20.0
K0711481-003	12/28/07	0.2010	20.0
K0711481-004	12/28/07	0.2010	20.0
K0711481-005	12/28/07	0.2040	20.0
K0711481-006	12/28/07	0.2310	20.0
K0711481-007	12/28/07	0.2090	20.0
K0711481-008	12/28/07	0.2390	20.0
K0711481-009	12/28/07	0.2100	20.0
K0711481-010	12/28/07	0.2070	20.0
K0711481-011	12/28/07	0.2180	20.0
K0711481-012	12/28/07	0.2110	20.0
K0711481-013	12/28/07	0.2010	20.0
K0711481-014	12/28/07	0.2140	20.0
K0711481-015	12/28/07	0.2070	20.0
K0711481-016	12/28/07	0.2070	20.0
K0711481-017	12/28/07	0.2100	20.0
K0711481-018	12/28/07	0.2070	20.0
K0711481-019	12/28/07	0.2030	20.0
K0711481-020	12/28/07	0.2130	20.0
K0711481-MB1	12/28/07	0.2000	20.0
LCSS DOLT	12/28/07	0.2000	20.0

Metals
-13-
PREPARATION LOG

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Method: F

Sample ID	Preparation Date	Initial Volume	Final Volume (mL)
K0711481-021	12/28/07	0.2080	20.0
K0711481-021D	12/28/07	0.2090	20.0
K0711481-021S	12/28/07	0.2000	20.0
K0711481-022	12/28/07	0.2060	20.0
K0711481-023	12/28/07	0.2020	20.0
K0711481-024	12/28/07	0.2090	20.0
K0711481-025	12/28/07	0.2110	20.0
K0711481-026	12/28/07	0.2030	20.0
K0711481-027	12/28/07	0.2160	20.0
K0711481-028	12/28/07	0.2140	20.0
K0711481-029	12/28/07	0.2070	20.0
K0711481-030	12/28/07	0.2270	20.0
K0711481-031	12/28/07	0.2100	20.0
K0711481-032	12/28/07	0.2050	20.0
K0711481-033	12/28/07	0.2150	20.0
K0711481-034	12/28/07	0.2370	20.0
K0711481-035	12/28/07	0.2430	20.0
K0711481-036	12/28/07	0.2320	20.0
K0711481-037	12/28/07	0.2000	20.0
K0711481-038	12/28/07	0.2010	20.0
K0711481-MB2	12/28/07	0.2000	20.0
LCSS DOLT2	12/28/07	0.2070	20.0

Metals
- 14 -
ANALYSIS RUN LOG

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Instrument ID Number: K-FLAA-02

Method: F

Start Date: 1/7/08

End Date: 1/7/08

Sample No.	D/F	Time	% R	Analytes																									
				A L	S B	A S	B A	B E	C D	C A	C R	C O	C U	F E	P B	M G	M N	H G	N I	K	S E	A G	N A	T L	V	Z N	C N		
ZK0711481-016	10	10:10																											
CCV3	1	10:13																			X								
CCB3	1	10:15																			X								
K0711481-017	10	10:18																			X								
K0711481-018	10	10:20																			X								
ZK0711481-019	10	10:23																											
K0711481-020	10	10:26																			X								
K0711481-001A	10	10:28	1125.																		X								
CCV4	1	10:30																			X								
CCB4	1	10:33																			X								
ZLCSS DOLT	10	10:35																											
K0711481-003	20	10:38																			X								
K0711481-006	20	10:40																			X								
K0711481-008	20	10:43																			X								
K0711481-011	20	10:45																			X								
K0711481-014	20	10:48																			X								
K0711481-015	10	10:50																			X								
K0711481-016	20	10:53																			X								
K0711481-019	20	10:55																			X								
LCSS DOLT	40	10:58																			X								
CCV5	1	11:00																			X								
CCB5	1	11:03																			X								

* - Denotes additional elements (other than the standard CLP elements) are represented on another Form 14

Metals
- 14 -
ANALYSIS RUN LOG

Client: New Fields Environmental

Service Request: K0711481

Project No.: NA

Project Name: Se in Tissue

Instrument ID Number: K-FLAA-02

Method: F

Start Date: 1/21/08

End Date: 1/21/08

Sample No.	D/F	Time	% R	Analytes																									
				A L	S B	A S	B A	B E	C D	C A	C R	C O	C U	F E	P B	M G	M N	H G	N I	K	S E	A G	N A	T L	V	Z N	C N		
CAL BLK	1	09:44																			X								
STD 0.5	1	09:46																			X								
STD 1.0	1	09:48																			X								
STD 5.0	1	09:51																			X								
STD 10.0	1	09:53																			X								
STD 15.0	1	09:55																			X								
ICV2	1	09:58																			X								
ICB2	1	10:00																			X								
CRA2	1	10:03																			X								
CCV1	1	10:05																			X								
CCB1	1	10:07																			X								
K0711481-MB2	2	10:10																			X								
LCSS DOLT2	20	10:12																			X								
K0711481-021	10	10:14																			X								
K0711481-021D	10	10:16																			X								
K0711481-021S	40	10:19																			X								
ZK0711481-022	10	10:21																											
K0711481-023	10	10:24																			X								
K0711481-024	10	10:27																			X								
K0711481-025	10	10:29																			X								
K0711481-026	10	10:31																			X								
CCV2	1	10:34																			X								
CCB2	1	10:36																			X								
K0711481-027	10	10:38																			X								
K0711481-028	10	10:41																			X								
K0711481-029	10	10:43																			X								
K0711481-030	10	10:45																			X								
K0711481-031	10	10:48																			X								
K0711481-032	10	10:50																			X								
K0711481-033	10	10:52																			X								
K0711481-034	10	10:55																			X								
ZK0711481-035	10	10:57																											

* - Denotes additional elements (other than the standard CLP elements) are represented on another Form 14

Columbia Analytical Services
Metals Tissue Digestion Sheet

Service Request Number(s): K0711481 1-20

Star Lims Run No.: 100083 Analysis for: ICP ICP-MS GFAA
Method: Tissue other: Se hydride

Sample	Initial Weight (g)	freeze Dry	Wet	Final Volume (ml)	Matrix
PB			X	20mls	15L HNO3
Dolt	0.200		I		
K0711481-01	0.218	X			
- 01D	0.208				
- 015	0.206				
- 02	0.208				
- 03	0.201				
- 04	0.201				
- 05	0.204				
- 06	0.231				
- 07	0.209				
- 08	0.239				
- 09	0.210				
- 10	0.207				
- 11	0.218				
- 12	0.211				
- 13	0.201				
- 14	0.214				
- 15	0.207				
- 16	0.207				
- 17	0.210				
- 18	0.207				
- 19	0.203				
- 20	0.213				
		<u>AB 12/28/07</u>			

Time Digestion Started: 4:15pm 12/28/07 Oven Temp: 105°C
Lot # Acids Used: HNO3 MS10-81°C

Time Digestion Ended: 10:30am 1/2/08
Oven Temp: 108°C
Balance I.D.: 21B

LCS: Dorm-2, Dolt-3

QCP CICV-1, MET1-59-H, _____ mls. added
QCP CICV-2, MET1-59-I, _____ mls. added
QCP CICV-3, MET1-59-J, _____ mls. Added
SS6, MET1-62-A, _____ mls. Added

SPIKE INFO

SS1-MET1-61-R, 0.3 mls added
SS5-MET1-61-P, 0.05 mls added
SS6-MET1-62-A, 0.05 mls added

Additional spikes: _____

Comments: _____

Analyst <u>Angela Black</u>	Date <u>12/28/07</u>
Reviewer <u>[Signature]</u>	Date <u>1/2/08</u>

TissueDig.xls
12/13/2007

METALS SPIKE FORM

Service Request # K0711481 1-20
 Q.C. Sample # K0711481-1

Circle type of digest: GFAA ICP FAA ICP-MS Other: _____ Initials / Date: AB, 12/28/07
 Circle type of sample: Soil Water Misc. Sludge Oil Other: TISSUE

Solution Name	Element	mls of 1000ppm Solution	Final Volume	Source	Lot#	Exp. Date	Solution Conc. mg/L	Enter mls Added
SS1-MET1-61-R	HNO3	50.0	1000ml	JT Baker	E17044	-	-	0.3 Expires:3/7/08
	Al	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	200	
	Ag	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Ba	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	200	
	Be	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Cd	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Co	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Cr	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	20	
	Cu	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	25	
	Fe	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	100	
	Pb	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Mn	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Ni	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Sb	50*	1000ml	MET1-53-A	619312	3/7/2008	50	
V	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50		
Zn	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50		
SS4-MET1-62-J	HNO3	25.0	500ml	JT BAKER	A48046	-	-	Expires:2/1/08
	As	2.0	500ml	MET1-57-Z	Z-AS02032	7/1/2008	4	
	Cd	2.0	500ml	MET1-57-M	Z-CD02004	7/1/2008	4	
	Pb	2.0	500ml	MET1-57-R	A2-PB02135	7/1/2008	4	
	Se	2.0	500ml	MET1-57-K	Z-SE01120	7/1/2008	4	
	Tl	2.0	500ml	MET1-54-M	Z-TL01097	2/1/2008	4	
	Cu	2.0	500ml	MET1-55-O	Z-CU02084	5/1/2008	4	
SS5-MET1-61-P	HNO3	25.0	500ml	JT BAKER	E17044	-	-	0.05 Expires:2/1/08
	As	50.0	500ml	MET1-57-Z	Z-AS02032	7/1/2008	100	
	Se	50.0	500ml	MET1-59-C	Z-SE01120	8/1/2008	100	
	Tl	50.0	500ml	MET1-54-M	Z-TL01097	2/1/2008	100	
SS6-MET1-62-A	HNO3	25	500ml	JT BAKER	E27027	-	-	0.05 Expires: 2/1/08
	B	50	500ml	MET1-59-A	715006	1/2/2009	100	
	Mo	50	500ml	MET1-54-K	Z-MO02012	2/1/2008	100	

GFLCSW (MET1-60-A)	HNO3	10.0	1000ml	JT BAKER	E01042	-	-	Expires: 3/1/08
	As, Pb, Se, Tl	5.0	1000ml	QCP-CICV-3	Z-CICP19048	3/1/2008	2.5	
	Cd	-	-	QCP-CICV-3	Z-CICP19048	3/1/2008	1.25	
	Cu	2.5	1000ml	MET-55-O	Z-CU0284	5/1/2008	2.5	
QCP-CICV-1 (MET1-59-H)	Ca, Mg, Na, K	no dilution	-	IV	AZ-MEB236021	9/1/2008	2500	Expires:9/1/08
	Al, Ba	no dilution	-	IV	AZ-MEB236021	9/1/2008	1000	
	Fe	no dilution	-	IV	AZ-MEB236021	9/1/2008	500	
	Co, Mn, Ni, V, Zn	no dilution	-	IV	AZ-MEB236021	9/1/2008	250	
	Cu, Ag	no dilution	-	IV	AZ-MEB236021	9/1/2008	125	
	Cr	no dilution	-	IV	AZ-MEB236021	9/1/2008	100	
	Be	no dilution	-	IV	AZ-MEB236021	9/1/2008	25	
QCP-CICV-2 (MET1-59-I)	Sb	no dilution	-	IV	Z-CICP19033	9/1/2008	500	Expires: 9/1/08
QCP-CICV-3 (MET1-59-J)	As, Pb, Se, Tl	no dilution	-	IV	Z-CICP19048	9/1/2008	500	Expires: 9/1/08
	Cd	no dilution	-	IV	Z-CICP19048	9/1/2008	250	

* Denotes volume of 1000 ppm stock standard.

Element	mls of	ppm	Source	Lot# / Lab Code	Exp. Date

Columbia Analytical Services
Metals Tissue Digestion Sheet

Service Request Number(s) : K0711481 (21-38)

Star Lims Run No.: 000684 Analysis for: ICP ICP-MS GFAA
Method: Tissue other: Se hyande

Sample	Initial Weight (g)	freeze Dry	Wet	Final Volume (ml)	Matrix
DB			X	20mls	15/1 HNDZ
DDIF	0.207		↓		
K0711481-21	0.208	X			
- 21D	0.209				
- 21S	0.200				
- 22	0.206				
- 23	0.202				
- 24	0.209				
- 25	0.211				
- 26	0.203				
- 27	0.216				
- 28	0.214				
- 29	0.207				
- 30	0.227				
- 31	0.210				
- 32	0.205				
- 33	0.215				
- 34	0.237				
- 35	0.243				
- 36	0.232				
- 37	0.200				
- 38	0.201				
<u>AB 12/28/07</u>					

Time Digestion Started: 4:15p.m. 12/28/07 Oven Temp: 105°C
Lot # Acids Used: HNO3 4.810.81-C

Time Digestion Ended: 10:30a.m. 1/2/08
Oven Temp: 108°C

LCS: Dorm-2, Dolt-3

Balance I.D.: 21B

QCP CICV-1, MET1-59-H, _____ mls. added
QCP CICV-2, MET1-59-I, _____ mls. added
QCP CICV-3, MET1-59-J, _____ mls. Added
SS6, MET1-62-A, _____ mls. Added

SPIKE INFO

SS1-MET1-61-R, 0.3 mls added
SS5-MET1-61-P, 0.05 mls added
SS6-MET1-62-A, 0.05 mls added

Additional spikes: _____

Comments: _____

Analyst <u>Angela Black</u>	Date <u>12/28/07</u>
Reviewer <u>[Signature]</u>	Date <u>1/2/08</u>

TissueDig.xls
12/13/2007

METALS SPIKE FORM

Service Request # K0711481 21-38
 Q.C. Sample # K0711481-21

Circle type of digest: GFAA ICP **FAA** ICP-MS Other: _____ Initials / Date: AB, 12/28/07
 Circle type of sample: Soil Water Misc. Sludge Oil Other: TISSUE

Solution Name	Element	mls of 1000ppm Solution	Final Volume	Source	Lot#	Exp. Date	Solution Conc. mg/L	Enter mls Added
SS1-MET1-61-R	HNO3	50.0	1000ml	JT Baker	E17044	-	-	0.3
	Al	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	200	
	Ag	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Ba	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	200	
	Be	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Cd	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Co	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Cr	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	20	
	Cu	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	25	
	Fe	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	100	
	Pb	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Mn	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Ni	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Sb	50*	1000ml	MET1-53-A	619312	3/7/2008	50	
	V	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
Zn	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50		
SS4-MET1-62-J	HNO3	25.0	500ml	JT BAKER	A48046	-	-	Expires: 2/1/08
	As	2.0	500ml	MET1-57-Z	Z-AS02032	7/1/2008	4	
	Cd	2.0	500ml	MET1-57-M	Z-CD02004	7/1/2008	4	
	Pb	2.0	500ml	MET1-57-R	A2-PB02135	7/1/2008	4	
	Se	2.0	500ml	MET1-57-K	Z-SE01120	7/1/2008	4	
	Tl	2.0	500ml	MET1-54-M	Z-TL01097	2/1/2008	4	
	Cu	2.0	500ml	MET1-55-O	Z-CU02084	5/1/2008	4	
SS5-MET1-61-P	HNO3	25.0	500ml	JT BAKER	E17044	-	-	Expires: 2/1/08
	As	50.0	500ml	MET1-57-Z	Z-AS02032	7/1/2008	100	
	Se	50.0	500ml	MET1-59-C	Z-SE01120	8/1/2008	100	
	Tl	50.0	500ml	MET1-54-M	Z-TL01097	2/1/2008	100	
SS6-MET1-62-A	HNO3	25	500ml	JT BAKER	E27027	-	-	Expires: 2/1/08
	B	50	500ml	MET1-59-A	715006	1/2/2009	100	
	Mo	50	500ml	MET1-54-K	Z-MO02012	2/1/2008	100	

GFLCSW (MET1-60-A)	HNO3	10.0	1000ml	JT BAKER	E01042	-	-	Expires: 3/1/08
	As, Pb, Se, Tl	5.0	1000ml	QCP-CICV-3	Z-CICP19048	3/1/2008	2.5	
	Cd	-	-	QCP-CICV-3	Z-CICP19048	3/1/2008	1.25	
	Cu	2.5	1000ml	MET-55-Q	Z-CU0284	5/1/2008	2.5	
QCP-CICV-1 (MET1-59-H)	Ca, Mg, Na, K	no dilution	-	IV	AZ-MEB236021	9/1/2008	2500	Expires: 9/1/08
	Al, Ba	no dilution	-	IV	AZ-MEB236021	9/1/2008	1000	
	Fe	no dilution	-	IV	AZ-MEB236021	9/1/2008	500	
	Co, Mn, Ni, V, Zn	no dilution	-	IV	AZ-MEB236021	9/1/2008	250	
	Cu, Ag	no dilution	-	IV	AZ-MEB236021	9/1/2008	125	
	Cr	no dilution	-	IV	AZ-MEB236021	9/1/2008	100	
QCP-CICV-2 (MET1-59-I)	Be	no dilution	-	IV	AZ-MEB236021	9/1/2008	25	Expires: 9/1/08
	Sb	no dilution	-	IV	Z-CICP19033	9/1/2008	500	
QCP-CICV-3 (MET1-59-J)	As, Pb, Se, Tl	no dilution	-	IV	Z-CICP19048	9/1/2008	500	Expires: 9/1/08
	Cd	no dilution	-	IV	Z-CICP19048	9/1/2008	250	

* Denotes volume of 1000 ppm stock standard.

Element	mls of	ppm	Source	Lot# / Lab Code	Exp. Date

Element Analyzed Se Hydride Instrument K-FLAA-2
Service Request # K11481 # 1-20

Batch QC SR's # _____

Calibration Std. AA1-8-A

Starlims # 103600

Run # 010708-Se

Hydride Data Review Form

	Yes	No	NA
1. ICV within 10% of true Value	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
2. Calibration data included	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
3. CCV's in control	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
4. CCB's and/or ICB's below MRL	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
5. All reported Results within Cal. Range	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
6. All Calculations are Correct	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Comments

Primary Reviewed by A Date 1/7/08

Secondary Reviewed by JDB Date 1/7/08

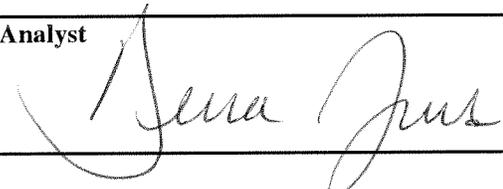
COLUMBIA ANALYTICAL SERVICES, INC.

GFAA Run Log

Method: (Circle Method Used) 7742 7062 Other: _____ Element: As Se	Service Request # :
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SAMPLE NUMBER	Dilution Factor	Measured (µg/L)	Recoveries (ICV, CCV, CRA, LCS, Matrix Spk.)	Comments
ICV	-	9.940	99%	
ICB	-	0.000		
CRA	-	0.592	118%	
CCV	-	9.711	97%	
CCB	-	-0.037		
K0711481-MB	1/2	0.068		
DOLT K0711481	1/2+1/10	1.403		Rerun
K0711481-001	1/2+1/5	9.429		
K0711481-001D	1/2+1/5	8.441		
K0711481-001S	1/2+1/20	7.650	85%	
K0711481-002	1/2+1/5	5.626		1/17/08
K0711481-003	1/2+1/5	16.616		Rerun
K0711481-004	1/2+1/5	8.952		
K0711481-005	1/2+1/5	14.026		
K0711481-006	1/2+1/5	16.242		Rerun
CCV	-	9.302	93%	
CCB	-	-0.003		
K0711481-007	1/2+1/5	14.193		
K0711481-008	1/2+1/5	15.685		Rerun
K0711481-009	1/2+1/5	7.051		
K0711481-010	1/2+1/5	9.902		
K0711481-011	1/2+1/5	18.565		Rerun
K0711481-012	1/2+1/5	13.904		1/17/08
K0711481-013	1/2+1/5	7.194		
K0711481-014	1/2+1/5	19.219		Rerun
K0711481-015	1/2+1/5	8.995		Rerun
K0711481-016	1/2+1/5	17.827		Rerun
CCV	-	10.039	100%	
CCB	-	0.002		
K0711481-017	1/2+1/5	9.638		

True Values/QC Limits:	LCSW	Water Spike	LCSS (ERA D045540)	Soil Spike
Arsenic:	2500ppb (80-120%)	1000ppb (80-120%)	146.0mg/kg (80-120%)	1000ppb (80-120%)
Selenium	2500ppb (71-122%)	1000ppb (80-120%)	73.0mg/kg (61-142%)	1000ppb (80-120%)

Analyst 	Date: 1/17/08	Page Number: 1
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Sample ID: Std 5.0
Analyst:

Date Collected: 1/7/2008 8:59:16 AM
Data Type: Original

Replicate Data: Std 5.0

Repl #	Sample Conc ug/L	Stnd Conc ug/L	Blk Corr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[5.0]	0.075	0.382	0.079			08:59:38	Yes
2		[5.0]	0.075	0.382	0.080			09:00:13	Yes
3		[5.0]	0.075	0.376	0.080			09:00:47	Yes
Mean:		[5.0]	0.075						
SD:		0.0	0.0003						
%RSD:		0.0	0.44						
Standard number 3 applied. [5.0]									
Correlation Coef.: 0.999642 Slope: 0.01504 Intercept: 0.00000									

Sequence No.: 5
Sample ID: Std 10.0
Analyst:

Autosampler Location: 5
Date Collected: 1/7/2008 9:01:38 AM
Data Type: Original

Replicate Data: Std 10.0

Repl #	Sample Conc ug/L	Stnd Conc ug/L	Blk Corr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[10.0]	0.143	0.732	0.148			09:02:01	Yes
2		[10.0]	0.142	0.702	0.147			09:02:36	Yes
3		[10.0]	0.141	0.731	0.145			09:03:10	Yes
Mean:		[10.0]	0.142						
SD:		0.0	0.0014						
%RSD:		0.0	0.98						
Standard number 4 applied. [10.0]									
Correlation Coef.: 0.999341 Slope: 0.01439 Intercept: 0.00000									

Sequence No.: 6
Sample ID: Std 15.0
Analyst:

Autosampler Location: 6
Date Collected: 1/7/2008 9:04:01 AM
Data Type: Original

Replicate Data: Std 15.0

Repl #	Sample Conc ug/L	Stnd Conc ug/L	Blk Corr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[15.0]	0.226	1.120	0.230			09:04:25	Yes
2		[15.0]	0.226	1.103	0.230			09:04:59	Yes
3		[15.0]	0.226	1.101	0.231			09:05:33	Yes
Mean:		[15.0]	0.226						
SD:		0.0	0.0004						
%RSD:		0.0	0.16						
Standard number 5 applied. [15.0]									
Correlation Coef.: 0.999215 Slope: 0.01483 Intercept: 0.00000									
The calibration curve may not be linear.									

Calibration data for Se 196.03

Equation: Linear Through Zero

ID	Mean Signal (Abs)	Entered Conc. ug/L	Calculated Conc. ug/L	Standard Deviation	%RSD
Cal Blk	0.0000	0	0.000	0.00	21.8
Std 0.5	0.0089	0.5	0.601	0.00	10.9
Std 1.0	0.0149	1.0	1.008	0.00	5.2
Std 5.0	0.0750	5.0	5.061	0.00	0.4
Std 10.0	0.1421	10.0	9.587	0.00	1.0
Std 15.0	0.2259	15.0	15.235	0.00	0.2

Correlation Coef.: 0.999215 Slope: 0.01483 Intercept: 0.00000

Sequence No.: 7
 Sample ID: ICV
 Analyst:

Autosampler Location: 7
 Date Collected: 1/7/2008 9:06:25 AM
 Data Type: Original

Replicate Data: ICV

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.912	9.912	0.147	0.745	0.151			09:06:49	Yes
2	10.01	10.01	0.148	0.721	0.153			09:07:24	Yes
3	9.896	9.896	0.147	0.729	0.151			09:07:58	Yes
Mean:	9.940	9.940	0.147						
SD:	0.064	0.064	0.0009						
%RSD:	0.644	0.644	0.64						

QC value within limits for Se 196.03 Recovery = 99.40%
 All analyte(s) passed QC.

Sequence No.: 8
 Sample ID: ICB
 Analyst:

Autosampler Location: 1
 Date Collected: 1/7/2008 9:08:50 AM
 Data Type: Original

Replicate Data: ICB

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.069	0.069	0.001	0.026	0.006			09:09:11	Yes
2	0.012	0.012	0.000	0.025	0.005			09:09:45	Yes
3	-0.081	-0.081	-0.001	0.006	0.003			09:10:19	Yes
Mean:	0.000	0.000	-0.000						
SD:	0.076	0.076	0.0011						
%RSD:	>999.9%	>999.9%	>999.9%						

QC value within limits for Se 196.03 Recovery = Not calculated
 All analyte(s) passed QC.

Sequence No.: 9
 Sample ID: CRA
 Analyst:

Autosampler Location: 2
 Date Collected: 1/7/2008 9:11:07 AM
 Data Type: Original

Replicate Data: CRA

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.704	0.704	0.010	0.067	0.015			09:11:29	Yes
2	0.565	0.565	0.008	0.046	0.013			09:12:03	Yes
3	0.506	0.506	0.008	0.063	0.012			09:12:37	Yes
Mean:	0.592	0.592	0.009						
SD:	0.101	0.101	0.0015						
%RSD:	17.15	17.15	17.15						

QC value within limits for Se 196.03 Recovery = 118.38%
 All analyte(s) passed QC.

Sequence No.: 10
 Sample ID: CCV
 Analyst:

Autosampler Location: 5
 Date Collected: 1/7/2008 9:13:27 AM
 Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.774	9.774	0.145	0.728	0.149			09:13:50	Yes
2	9.543	9.543	0.141	0.730	0.146			09:14:25	Yes
3	9.816	9.816	0.146	0.740	0.150			09:14:59	Yes
Mean:	9.711	9.711	0.144						

SD: 0.148 0.148 0.0022
%RSD: 1.519 1.519 1.52

QC value within limits for Se 196.03 Recovery = 97.11%
All analyte(s) passed QC.

Sequence No.: 11
Sample ID: CCB
Analyst:

Autosampler Location: 1
Date Collected: 1/7/2008 9:15:50 AM
Data Type: Original

Replicate Data: CCB

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Sequence No.: 12
Sample ID: K0711481-MB
Analyst:

Autosampler Location: 9
Date Collected: 1/7/2008 9:18:09 AM
Data Type: Original

Replicate Data: K0711481-MB

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 13
Sample ID: DOLT K0711481
Analyst:

Autosampler Location: 10
Date Collected: 1/7/2008 9:20:28 AM
Data Type: Original

Replicate Data: DOLT K0711481

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 14
Sample ID: K0711481-001
Analyst:

Autosampler Location: 11
Date Collected: 1/7/2008 9:22:50 AM
Data Type: Original

Replicate Data: K0711481-001

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Mean: 9.429 9.429 0.140
 SD: 0.101 0.101 0.0015
 %RSD: 1.075 1.075 1.07

Sequence No.: 15
 Sample ID: K0711481-001D
 Analyst:

Autosampler Location: 12
 Date Collected: 1/7/2008 9:25:10 AM
 Data Type: Original

Replicate Data: K0711481-001D

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	8.415	8.415	0.125	0.627	0.129			09:25:32	Yes
2	8.543	8.543	0.127	0.626	0.131			09:26:06	Yes
3	8.367	8.367	0.124	0.621	0.129			09:26:41	Yes
Mean:	8.441	8.441	0.125						
SD:	0.091	0.091	0.0014						
%RSD:	1.079	1.079	1.08						

Sequence No.: 16
 Sample ID: K0711481-001S
 Analyst:

Autosampler Location: 13
 Date Collected: 1/7/2008 9:27:31 AM
 Data Type: Original

Replicate Data: K0711481-001S

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	7.477	7.477	0.111	0.536	0.115			09:27:54	Yes
2	7.611	7.611	0.113	0.549	0.117			09:28:29	Yes
3	7.861	7.861	0.117	0.564	0.121			09:29:03	Yes
Mean:	7.650	7.650	0.113						
SD:	0.195	0.195	0.0029						
%RSD:	2.553	2.553	2.55						

Sequence No.: 17
 Sample ID: K0711481-002
 Analyst:

Autosampler Location: 14
 Date Collected: 1/7/2008 9:29:54 AM
 Data Type: Original

Replicate Data: K0711481-002

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	5.671	5.671	0.084	0.431	0.089			09:30:18	Yes
2	5.567	5.567	0.083	0.414	0.087			09:30:53	Yes
3	5.639	5.639	0.084	0.417	0.088			09:31:26	Yes
Mean:	5.626	5.626	0.083						
SD:	0.053	0.053	0.0008						
%RSD:	0.943	0.943	0.94						

Sequence No.: 18
 Sample ID: K0711481-003
 Analyst:

Autosampler Location: 15
 Date Collected: 1/7/2008 9:32:18 AM
 Data Type: Original

Replicate Data: K0711481-003

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	16.47	16.47	0.244	1.238	0.249			09:32:42	Yes
2	16.61	16.61	0.246	1.213	0.251			09:33:18	Yes
Sample concentration is greater than that of the highest standard.									
3	16.76	16.76	0.249	1.243	0.253			09:33:53	Yes
Sample concentration is greater than that of the highest standard.									
Mean:	16.62	16.62	0.246						

SD: 0.146 0.146 0.0022
%RSD: 0.877 0.877 0.88

Sample concentration is greater than that of the highest standard.

Sequence No.: 19
Sample ID: K0711481-004
Analyst:

Autosampler Location: 16
Date Collected: 1/7/2008 9:35:18 AM
Data Type: Original

Replicate Data: K0711481-004

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 20
Sample ID: K0711481-005
Analyst:

Autosampler Location: 17
Date Collected: 1/7/2008 9:37:43 AM
Data Type: Original

Replicate Data: K0711481-005

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 21
Sample ID: K0711481-006
Analyst:

Autosampler Location: 18
Date Collected: 1/7/2008 9:40:03 AM
Data Type: Original

Handwritten note: 1/17/08

Replicate Data: K0711481-006

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 22
Sample ID: CCV
Analyst:

Autosampler Location: 5
Date Collected: 1/7/2008 9:42:56 AM
Data Type: Original

Replicate Data: CCV

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

%RSD: 1.321 1.321 1.32

Sequence No.: 27
Sample ID: K0711481-010
Analyst:

Autosampler Location: 22
Date Collected: 1/7/2008 9:55:13 AM
Data Type: Original

Replicate Data: K0711481-010

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.900	9.900	0.147	0.745	0.151			09:55:35	Yes
2	9.902	9.902	0.147	0.738	0.151			09:56:09	Yes
3	9.905	9.905	0.147	0.729	0.151			09:56:44	Yes
Mean:	9.902	9.902	0.147						
SD:	0.002	0.002	0.0000						
%RSD:	0.024	0.024	0.02						

Sequence No.: 28
Sample ID: K0711481-011
Analyst:

Autosampler Location: 23
Date Collected: 1/7/2008 9:57:34 AM
Data Type: Original

1/7/08

Replicate Data: K0711481-011

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	18.79	18.79	0.279	1.373	0.283			09:57:55	Yes
2	18.38	18.38	0.273	1.371	0.277			09:58:30	Yes
3	18.52	18.52	0.275	1.355	0.279			09:59:04	Yes
Mean:	18.57	18.57	0.275						
SD:	0.210	0.210	0.0031						
%RSD:	1.130	1.130	1.13						

Sample concentration is greater than that of the highest standard.

Sample concentration is greater than that of the highest standard.

Sample concentration is greater than that of the highest standard.

Sample concentration is greater than that of the highest standard.

Sequence No.: 29
Sample ID: K0711481-012
Analyst:

Autosampler Location: 24
Date Collected: 1/7/2008 10:00:28 AM
Data Type: Original

Replicate Data: K0711481-012

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	14.05	14.05	0.208	1.044	0.213			10:00:50	Yes
2	13.76	13.76	0.204	1.021	0.209			10:01:24	Yes
3	13.91	13.91	0.206	1.021	0.211			10:01:58	Yes
Mean:	13.90	13.90	0.206						
SD:	0.143	0.143	0.0021						
%RSD:	1.032	1.032	1.03						

Sequence No.: 30
Sample ID: K0711481-013
Analyst:

Autosampler Location: 25
Date Collected: 1/7/2008 10:02:48 AM
Data Type: Original

Replicate Data: K0711481-013

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	7.166	7.166	0.106	0.515	0.111			10:03:12	Yes
2	7.162	7.162	0.106	0.519	0.111			10:03:47	Yes
3	7.253	7.253	0.108	0.527	0.112			10:04:21	Yes
Mean:	7.194	7.194	0.107						

SD: 0.051 0.051 0.0008
 %RSD: 0.712 0.712 0.71

Sequence No.: 31
 Sample ID: K0711481-014
 Analyst:

Autosampler Location: 26
 Date Collected: 1/7/2008 10:05:12 AM
 Data Type: Original

Replicate Data: K0711481-014

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	19.21	19.21	0.285	1.441	0.289			10:05:34	Yes
Sample concentration is greater than that of the highest standard.									
2	19.48	19.48	0.289	1.434	0.293			10:06:08	Yes
Sample concentration is greater than that of the highest standard.									
3	18.97	18.97	0.281	1.420	0.286			10:06:42	Yes
Sample concentration is greater than that of the highest standard.									
Mean:	19.22	19.22	0.285						
SD:	0.253	0.253	0.0037						
%RSD:	1.315	1.315	1.32						
Sample concentration is greater than that of the highest standard.									

Sequence No.: 32
 Sample ID: K0711481-015
 Analyst:

Autosampler Location: 27
 Date Collected: 1/7/2008 10:08:06 AM
 Data Type: Original

Replicate Data: K0711481-015

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	5.646	5.646	0.084	0.406	0.088			10:08:29	Yes
2	5.570	5.570	0.083	0.412	0.087			10:09:04	Yes
3	15.77	15.77	0.234	2.499	0.238			10:09:38	Yes
Mean:	8.995	8.995	0.133						
SD:	5.866	5.866	0.0870						
%RSD:	65.22	65.22	65.22						

Sequence No.: 33
 Sample ID: K0711481-016
 Analyst:

Autosampler Location: 28
 Date Collected: 1/7/2008 10:10:29 AM
 Data Type: Original

Replicate Data: K0711481-016

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	16.55	16.55	0.245	1.196	0.250			10:10:52	Yes
Sample concentration is greater than that of the highest standard.									
2	18.24	18.24	0.270	1.312	0.275			10:11:26	Yes
Changing BOC Sample concentration is greater than that of the highest standard.									
3	18.69	18.69	0.277	1.390	0.282			10:12:00	Yes
Sample concentration is greater than that of the highest standard.									
Mean:	17.83	17.83	0.264						
SD:	1.128	1.128	0.0167						
%RSD:	6.327	6.327	6.33						
Changing BOC Sample concentration is greater than that of the highest standard.									

Sequence No.: 34
 Sample ID: CCV
 Analyst:

Autosampler Location: 5
 Date Collected: 1/7/2008 10:13:26 AM
 Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.07	10.07	0.149	0.759	0.154			10:13:49	Yes
2	10.06	10.06	0.149	0.722	0.154			10:14:23	Yes
3	9.994	9.994	0.148	0.678	0.153			10:14:58	Yes
Mean:	10.04	10.04	0.149						
SD:	0.040	0.040	0.0006						
%RSD:	0.396	0.396	0.40						

QC value within limits for Se 196.03 Recovery = 100.39%
All analyte(s) passed QC.

Sequence No.: 35
Sample ID: CCB
Analyst:

Autosampler Location: 1
Date Collected: 1/7/2008 10:15:51 AM
Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.089	0.089	0.001	0.015	0.006			10:16:12	Yes
2	-0.005	-0.005	-0.000	0.017	0.004			10:16:47	Yes
3	-0.078	-0.078	-0.001	-0.001	0.003			10:17:23	Yes
Mean:	0.002	0.002	0.000						
SD:	0.084	0.084	0.0012						
%RSD:	>999.9%	>999.9%	>999.9%						

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Sequence No.: 36
Sample ID: K0711481-017
Analyst:

Autosampler Location: 29
Date Collected: 1/7/2008 10:18:13 AM
Data Type: Original

Replicate Data: K0711481-017

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.714	9.714	0.144	0.715	0.149			10:18:37	Yes
2	9.637	9.637	0.143	0.713	0.147			10:19:12	Yes
3	9.564	9.564	0.142	0.701	0.146			10:19:47	Yes
Mean:	9.638	9.638	0.143						
SD:	0.075	0.075	0.0011						
%RSD:	0.778	0.778	0.78						

Sequence No.: 37
Sample ID: K0711481-018
Analyst:

Autosampler Location: 30
Date Collected: 1/7/2008 10:20:40 AM
Data Type: Original

Replicate Data: K0711481-018

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	13.62	13.62	0.202	0.987	0.207			10:21:04	Yes
2	13.50	13.50	0.200	0.997	0.205			10:21:38	Yes
3	13.96	13.96	0.207	0.992	0.212			10:22:12	Yes
Mean:	13.69	13.69	0.203						
SD:	0.237	0.237	0.0035						
%RSD:	1.733	1.733	1.73						

Sequence No.: 38
Sample ID: K0711481-019
Analyst:

Autosampler Location: 31
Date Collected: 1/7/2008 10:23:05 AM
Data Type: Original

Replicate Data: K0711481-019

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	16.40	16.40	0.243	1.219	0.248			10:23:29	Yes
2	16.44	16.44	0.244	1.209	0.248			10:24:04	Yes
3	16.42	16.42	0.243	1.198	0.248			10:24:38	Yes
Mean:	16.42	16.42	0.244						
SD:	0.019	0.019	0.0003						
%RSD:	0.116	0.116	0.12						

1/7/08

Sequence No.: 39
Sample ID: K0711481-020
Analyst:

Autosampler Location: 32
Date Collected: 1/7/2008 10:26:05 AM
Data Type: Original

Replicate Data: K0711481-020

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.247	9.247	0.137	0.669	0.142			10:26:26	Yes
2	9.192	9.192	0.136	0.643	0.141			10:27:00	Yes
3	9.120	9.120	0.135	0.656	0.140			10:27:35	Yes
Mean:	9.186	9.186	0.136						
SD:	0.063	0.063	0.0009						
%RSD:	0.690	0.690	0.69						

Sequence No.: 40
Sample ID: K0711481-001A
Analyst:

Autosampler Location: 33
Date Collected: 1/7/2008 10:28:23 AM
Data Type: Original

Replicate Data: K0711481-001A

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	12.80	12.80	0.190	0.966	0.194			10:28:43	Yes
2	12.78	12.78	0.189	0.914	0.194			10:29:18	Yes
3	12.83	12.83	0.190	1.150	0.195			10:29:53	Yes
Mean:	12.80	12.80	0.190						
SD:	0.025	0.025	0.0004						
%RSD:	0.196	0.196	0.20						

Sequence No.: 41
Sample ID: CCV
Analyst:

Autosampler Location: 5
Date Collected: 1/7/2008 10:30:42 AM
Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.18	10.18	0.151	0.742	0.155			10:31:05	Yes
2	10.35	10.35	0.154	0.775	0.158			10:31:40	Yes
3	10.10	10.10	0.150	0.687	0.154			10:32:14	Yes
Mean:	10.21	10.21	0.151						
SD:	0.129	0.129	0.0019						
%RSD:	1.260	1.260	1.26						

QC value within limits for Se 196.03 Recovery = 102.11%
All analyte(s) passed QC.

Sequence No.: 42
Sample ID: CCB
Analyst:

Autosampler Location: 1
Date Collected: 1/7/2008 10:33:06 AM
Data Type: Original

Mean: 10.35 10.35 0.153
SD: 0.115 0.115 0.0017
%RSD: 1.116 1.116 1.12

Sequence No.: 46

Sample ID: K0711481-008 1/10

Analyst:

Autosampler Location:

Date Collected: 1/7/2008 10:43:09 AM

Data Type: Original

Replicate Data: K0711481-008 1/10

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.806	9.806	0.145	0.750	0.150			10:43:28	Yes
2	10.83	10.83	0.161	0.733	0.165			10:44:02	Yes
3	10.15	10.15	0.150	0.733	0.155			10:44:37	Yes
Mean:	10.26	10.26	0.152						
SD:	0.522	0.522	0.0077						
%RSD:	5.086	5.086	5.09						

Sequence No.: 47

Sample ID: K0711481-0011 1/10

Analyst:

Autosampler Location:

Date Collected: 1/7/2008 10:45:38 AM

Data Type: Original

Replicate Data: K0711481-0011 1/10

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	12.53	12.53	0.186	0.889	0.190			10:45:54	Yes
2	12.07	12.07	0.179	0.801	0.184			10:46:28	Yes
3	12.33	12.33	0.183	0.876	0.187			10:47:02	Yes
Mean:	12.31	12.31	0.183						
SD:	0.228	0.228	0.0034						
%RSD:	1.856	1.856	1.86						

Sequence No.: 48

Sample ID: K0711481-014 1/10

Analyst:

Autosampler Location:

Date Collected: 1/7/2008 10:48:03 AM

Data Type: Original

Replicate Data: K0711481-014 1/10

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	12.69	12.69	0.188	0.926	0.193			10:48:20	Yes
2	13.03	13.03	0.193	0.928	0.198			10:48:55	Yes
3	12.52	12.52	0.186	0.903	0.190			10:49:29	Yes
Mean:	12.75	12.75	0.189						
SD:	0.263	0.263	0.0039						
%RSD:	2.064	2.064	2.06						

Sequence No.: 49

Sample ID: K0711481-015 1/5

Analyst:

Autosampler Location:

Date Collected: 1/7/2008 10:50:26 AM

Data Type: Original

Replicate Data: K0711481-015 1/5

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	6.414	6.414	0.095	0.485	0.100			10:50:42	Yes
2	6.389	6.389	0.095	0.476	0.099			10:51:17	Yes
3	6.436	6.436	0.095	0.468	0.100			10:51:52	Yes
Mean:	6.413	6.413	0.095						
SD:	0.024	0.024	0.0004						
%RSD:	0.369	0.369	0.37						

Sequence No.: 50
 Sample ID: K0711481-016 1/10
 Analyst:

Autosampler Location:
 Date Collected: 1/7/2008 10:53:00 AM
 Data Type: Original

Replicate Data: K0711481-016 1/10

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.716	9.716	0.144	0.711	0.149			10:53:16	Yes
2	9.886	9.886	0.147	0.710	0.151			10:53:51	Yes
3	9.693	9.693	0.144	0.693	0.148			10:54:25	Yes
Mean:	9.765	9.765	0.145						
SD:	0.105	0.105	0.0016						
%RSD:	1.079	1.079	1.08						

Sequence No.: 51
 Sample ID: K0711481-019 1/10
 Analyst:

Autosampler Location:
 Date Collected: 1/7/2008 10:55:25 AM
 Data Type: Original

Replicate Data: K0711481-019 1/10

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	8.911	8.911	0.132	0.647	0.137			10:55:42	Yes
2	8.667	8.667	0.129	0.615	0.133			10:56:17	Yes
3	8.723	8.723	0.129	0.621	0.134			10:56:51	Yes
Mean:	8.767	8.767	0.130						
SD:	0.128	0.128	0.0019						
%RSD:	1.461	1.461	1.46						

Sequence No.: 52
 Sample ID: LCSS DOLT
 Analyst:

Autosampler Location:
 Date Collected: 1/7/2008 10:58:20 AM
 Data Type: Original

Replicate Data: LCSS DOLT

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.040	2.040	0.030	0.174	0.035			10:58:36	Yes
2	1.954	1.954	0.029	0.144	0.034			10:59:11	Yes
3	2.048	2.048	0.030	0.152	0.035			10:59:45	Yes
Mean:	2.014	2.014	0.030						
SD:	0.052	0.052	0.0008						
%RSD:	2.601	2.601	2.60						

Sequence No.: 53
 Sample ID: CCV
 Analyst:

Autosampler Location:
 Date Collected: 1/7/2008 11:00:45 AM
 Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.961	9.961	0.148	0.732	0.152			11:01:01	Yes
2	10.01	10.01	0.148	0.730	0.153			11:01:36	Yes
3	10.04	10.04	0.149	0.724	0.153			11:02:10	Yes
Mean:	10.00	10.00	0.148						
SD:	0.038	0.038	0.0006						
%RSD:	0.382	0.382	0.38						

Sequence No.: 54

Autosampler Location:

Sample ID: CCB
Analyst:

Date Collected: 1/7/2008 11:03:05 AM
Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.026	0.026	0.000	0.018	0.005			11:03:21	Yes
2	-0.053	-0.053	-0.001	0.014	0.004			11:03:56	Yes
3	-0.014	-0.014	-0.000	0.017	0.004			11:04:30	Yes
Mean:	-0.014	-0.014	-0.000						
SD:	0.040	0.040	0.0006						
%RSD:	286.7	286.7	286.74						

Sequence No.: 55
Sample ID: LCSS DOLT 1/10
Analyst:

Autosampler Location:
Date Collected: 1/7/2008 11:05:29 AM
Data Type: Original

1/7/08

Replicate Data: LCSS DOLT 1/10

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.971	2.971	0.044	0.247	0.049			11:05:45	Yes
2	2.881	2.881	0.043	0.218	0.047			11:06:19	Yes
3	3.028	3.028	0.045	0.239	0.049			11:06:53	Yes
Mean:	2.960	2.960	0.044						
SD:	0.074	0.074	0.0011						
%RSD:	2.503	2.503	2.50						

Element Analyzed Se Hydride Instrument K-FLAA-2
Service Request # K11481

Batch QC SR's # _____

Calibration Std. AA1-8-A

Starlims # 104630

Run # 012108-Se

Hydride Data Review Form

	Yes	No	NA
1. ICV within 10% of true Value	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
2. Calibration data included	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
3. CCV's in control	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
4. CCB's and/or ICB's below MRL	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
5. All reported Results within Cal. Range	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
6. All Calculations are Correct	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Comments

Primary Reviewed by d Date 1/21/08

Secondary Reviewed by JPB Date 1/21/08

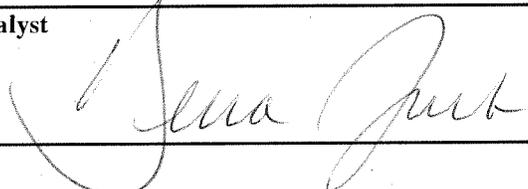
COLUMBIA ANALYTICAL SERVICES, INC.

GFAA Run Log

Method: (Circle Method Used) 7742 7062 Other: _____ Element: <u>As Se</u>	Service Request # : _____
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SAMPLE NUMBER	Dilution Factor	Measured (µg/L)	Recoveries (ICV, CCV, CRA, LCS, Matrix Spk.)	Comments
ICV	-	9.466	95%	
ICB	-	0.171		
CRA	-	0.638	128%	
CCV	-	9.609	96%	
CCB	-	0.072		
K0711481-MB	1/2	-0.079		
LCSS DOLT	1/2+1/10	3.393	96%	
K0711481-021	1/2+1/5	11.708		
K0711481-021D	1/2+1/5	11.880		
K0711481-021S	1/2+1/20	9.848	111%	
K0711481-022	1/2+1/5	17.901		1/21/08 Rerun
K0711481-023	1/2+1/5	8.439		
K0711481-024	1/2+1/5	5.894		
K0711481-025	1/2+1/5	7.036		
K0711481-026	1/2+1/5	6.022		
CCV	-	9.613	96%	
CCB	-	0.107		
K0711481-027	1/2+1/5	6.511		
K0711481-028	1/2+1/5	7.509		
K0711481-029	1/2+1/5	5.800		
K0711481-030	1/2+1/5	5.329		
K0711481-031	1/2+1/5	7.542		
K0711481-032	1/2+1/5	9.471		
K0711481-033	1/2+1/5	5.860		
K0711481-034	1/2+1/5	10.014		
K0711481-035	1/2+1/5	20.014		1/21/08 Rerun
K0711481-036	1/2+1/5	12.483		
CCV	-	9.709	97%	
CCB	-	0.181		
K0711481-037	1/2+1/5	31.523		1/21/08 Rerun

True Values/QC Limits:	LCSW	Water Spike	LCSS (ERA D045540)	Soil Spike
Arsenic:	2500ppb (80-120%)	1000ppb (80-120%)	146.0mg/kg (80-120%)	1000ppb (80-120%)
Selenium	2500ppb (71-122%)	1000ppb (65-122%)	73.0mg/kg (61-142%)	1000ppb (65-124%)

Analyst 	Date: 1/21/08	Page Number: 7
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Sample ID: Std 5.0
Analyst:

Date Collected: 1/21/2008 9:51:04 AM
Data Type: Original

Replicate Data: Std 5.0

Repl #	Sample Conc ug/L	Std Conc ug/L	Blk Corr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[5.0]	0.062	0.352	0.067			09:51:26	Yes
2		[5.0]	0.062	0.331	0.067			09:52:04	Yes
3		[5.0]	0.064	0.360	0.069			09:52:38	Yes
Mean:		[5.0]	0.062						
SD:		0.0	0.0016						
%RSD:		0.0	2.60						

Standard number 3 applied. [5.0]
Correlation Coef.: 0.999135 Slope: 0.01257 Intercept: 0.00000

Sequence No.: 5
Sample ID: Std 10.0
Analyst:

Autosampler Location: 5
Date Collected: 1/21/2008 9:53:29 AM
Data Type: Original

Replicate Data: Std 10.0

Repl #	Sample Conc ug/L	Std Conc ug/L	Blk Corr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[10.0]	0.147	0.936	0.152			09:53:52	Yes
2		[10.0]	0.123	0.657	0.128			09:54:25	Yes
3		[10.0]	0.124	0.676	0.129			09:55:00	Yes
Mean:		[10.0]	0.132						
SD:		0.0	0.0137						
%RSD:		0.0	10.43						

Standard number 4 applied. [10.0]
Correlation Coef.: 0.999467 Slope: 0.01304 Intercept: 0.00000

Sequence No.: 6
Sample ID: Std 15.0
Analyst:

Autosampler Location: 6
Date Collected: 1/21/2008 9:55:51 AM
Data Type: Original

Replicate Data: Std 15.0

Repl #	Sample Conc ug/L	Std Conc ug/L	Blk Corr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[15.0]	0.206	1.094	0.211			09:56:14	Yes
2		[15.0]	0.200	1.039	0.205			09:56:48	Yes
3		[15.0]	0.199	1.037	0.204			09:57:23	Yes
Mean:		[15.0]	0.202						
SD:		0.0	0.0041						
%RSD:		0.0	2.04						

Standard number 5 applied. [15.0]
Correlation Coef.: 0.999548 Slope: 0.01330 Intercept: 0.00000
The calibration curve may not be linear.

Calibration data for Se 196.03

Equation: Linear Through Zero

ID	Mean Signal (Abs)	Entered Conc. ug/L	Calculated Conc. ug/L	Standard Deviation	%RSD
Cal Blk	0.0000	0	0.000	0.00	4.9
Std 0.5	0.0078	0.5	0.586	0.00	2.6
Std 1.0	0.0135	1.0	1.017	0.00	2.6
Std 5.0	0.0625	5.0	4.697	0.00	2.6
Std 10.0	0.1315	10.0	9.889	0.01	10.4
Std 15.0	0.2016	15.0	15.162	0.00	2.0

Correlation Coef.: 0.999548 Slope: 0.01330 Intercept: 0.00000

Sequence No.: 7
 Sample ID: ICV
 Analyst:

Autosampler Location: 7
 Date Collected: 1/21/2008 9:58:14 AM
 Data Type: Original

Replicate Data: ICV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.637	9.637	0.128	0.714	0.133			09:58:38	Yes
2	9.305	9.305	0.124	0.664	0.129			09:59:13	Yes
3	9.457	9.457	0.126	0.676	0.131			09:59:47	Yes
Mean:	9.466	9.466	0.126						
SD:	0.166	0.166	0.0022						
%RSD:	1.757	1.757	1.76						

QC value within limits for Se 196.03 Recovery = 94.66%
 All analyte(s) passed QC.

Sequence No.: 8
 Sample ID: ICB
 Analyst:

Autosampler Location: 1
 Date Collected: 1/21/2008 10:00:39 AM
 Data Type: Original

Replicate Data: ICB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.210	0.210	0.003	0.028	0.008			10:01:00	Yes
2	0.203	0.203	0.003	0.025	0.008			10:01:36	Yes
3	0.099	0.099	0.001	0.026	0.006			10:02:11	Yes
Mean:	0.171	0.171	0.002						
SD:	0.062	0.062	0.0008						
%RSD:	36.37	36.37	36.37						

QC value within limits for Se 196.03 Recovery = Not calculated
 All analyte(s) passed QC.

Sequence No.: 9
 Sample ID: CRA
 Analyst:

Autosampler Location: 2
 Date Collected: 1/21/2008 10:03:00 AM
 Data Type: Original

Replicate Data: CRA

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.628	0.628	0.008	0.060	0.013			10:03:22	Yes
2	0.590	0.590	0.008	0.054	0.013			10:03:56	Yes
3	0.696	0.696	0.009	0.063	0.014			10:04:30	Yes
Mean:	0.638	0.638	0.008						
SD:	0.054	0.054	0.0007						
%RSD:	8.423	8.423	8.42						

QC value within limits for Se 196.03 Recovery = 127.67%
 All analyte(s) passed QC.

Sequence No.: 10
 Sample ID: CCV
 Analyst:

Autosampler Location: 5
 Date Collected: 1/21/2008 10:05:19 AM
 Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.838	9.838	0.131	0.721	0.136			10:05:42	Yes
2	9.571	9.571	0.127	0.672	0.132			10:06:16	Yes
3	9.417	9.417	0.125	0.690	0.130			10:06:51	Yes
Mean:	9.609	9.609	0.128						

SD: 0.213 0.213 0.0028
 %RSD: 2.220 2.220 2.22

QC value within limits for Se 196.03 Recovery = 96.09%
 All analyte(s) passed QC.

Sequence No.: 11

Sample ID: CCB

Analyst:

Autosampler Location: 1

Date Collected: 1/21/2008 10:07:41 AM

Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.227	0.227	0.003	0.033	0.008			10:08:02	Yes
2	-0.010	-0.010	-0.000	-0.002	0.005			10:08:37	Yes
3	0.001	0.001	0.000	0.005	0.005			10:09:11	Yes
Mean:	0.072	0.072	0.001						
SD:	0.134	0.134	0.0018						
%RSD:	184.7	184.7	184.70						

QC value within limits for Se 196.03 Recovery = Not calculated
 All analyte(s) passed QC.

Sequence No.: 12

Sample ID: K0711481-MB

Analyst:

Autosampler Location: 9

Date Collected: 1/21/2008 10:10:00 AM

Data Type: Original

Replicate Data: K0711481-MB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	-0.085	-0.085	-0.001	0.013	0.004			10:10:21	Yes
2	-0.030	-0.030	-0.000	0.017	0.005			10:10:55	Yes
3	-0.122	-0.122	-0.002	0.008	0.003			10:11:29	Yes
Mean:	-0.079	-0.079	-0.001						
SD:	0.046	0.046	0.0006						
%RSD:	58.45	58.45	58.45						

Sequence No.: 13

Sample ID: LCSS DOLT

Analyst:

Autosampler Location: 10

Date Collected: 1/21/2008 10:12:18 AM

Data Type: Original

Replicate Data: LCSS DOLT

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.475	3.475	0.046	0.254	0.051			10:12:40	Yes
2	3.319	3.319	0.044	0.239	0.049			10:13:15	Yes
3	3.385	3.385	0.045	0.239	0.050			10:13:49	Yes
Mean:	3.393	3.393	0.045						
SD:	0.078	0.078	0.0010						
%RSD:	2.308	2.308	2.31						

Sequence No.: 14

Sample ID: K0711481-021

Analyst:

Autosampler Location: 11

Date Collected: 1/21/2008 10:14:38 AM

Data Type: Original

Replicate Data: K0711481-021

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	11.85	11.85	0.158	0.834	0.163			10:15:00	Yes
2	11.66	11.66	0.155	0.804	0.160			10:15:35	Yes
3	11.62	11.62	0.155	0.806	0.160			10:16:09	Yes

Mean: 11.71 11.71 0.156
SD: 0.122 0.122 0.0016
%RSD: 1.039 1.039 1.04

Sequence No.: 15
Sample ID: K0711481-021D
Analyst:

Autosampler Location: 12
Date Collected: 1/21/2008 10:16:59 AM
Data Type: Original

Replicate Data: K0711481-021D

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 16
Sample ID: K0711481-021S
Analyst:

Autosampler Location: 13
Date Collected: 1/21/2008 10:19:20 AM
Data Type: Original

Replicate Data: K0711481-021S

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 17
Sample ID: K0711481-022
Analyst:

Autosampler Location: 14
Date Collected: 1/21/2008 10:21:42 AM
Data Type: Original

Replicate Data: K0711481-022

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Includes handwritten note '1/21/08' and 'Sample concentration is greater than that of the highest standard.' for replicates 1, 2, and 3.

Sequence No.: 18
Sample ID: K0711481-023
Analyst:

Autosampler Location: 15
Date Collected: 1/21/2008 10:24:42 AM
Data Type: Original

Replicate Data: K0711481-023

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 2 replicate rows and summary statistics.

3	8.321	8.321	0.111	0.586	0.116			10:26:15	Yes
Mean:	8.439	8.439	0.112						
SD:	0.146	0.146	0.0019						
%RSD:	1.730	1.730	1.73						

Sequence No.: 19
 Sample ID: K0711481-024
 Analyst:

Autosampler Location: 16
 Date Collected: 1/21/2008 10:27:08 AM
 Data Type: Original

Replicate Data: K0711481-024

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	5.947	5.947	0.079	0.428	0.084			10:27:32	Yes
2	5.877	5.877	0.078	0.424	0.083			10:28:06	Yes
3	5.859	5.859	0.078	0.405	0.083			10:28:41	Yes
Mean:	5.894	5.894	0.078						
SD:	0.046	0.046	0.0006						
%RSD:	0.788	0.788	0.79						

Sequence No.: 20
 Sample ID: K0711481-025
 Analyst:

Autosampler Location: 17
 Date Collected: 1/21/2008 10:29:32 AM
 Data Type: Original

Replicate Data: K0711481-025

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	6.328	6.328	0.084	0.603	0.089			10:29:53	Yes
2	7.454	7.454	0.099	0.519	0.104			10:30:28	Yes
3	7.325	7.325	0.097	0.509	0.102			10:31:03	Yes
Mean:	7.036	7.036	0.094						
SD:	0.616	0.616	0.0082						
%RSD:	8.762	8.762	8.76						

Sequence No.: 21
 Sample ID: K0711481-026
 Analyst:

Autosampler Location: 18
 Date Collected: 1/21/2008 10:31:54 AM
 Data Type: Original

Replicate Data: K0711481-026

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	6.039	6.039	0.080	0.421	0.085			10:32:15	Yes
2	6.190	6.190	0.082	0.449	0.087			10:32:49	Yes
3	5.838	5.838	0.078	0.378	0.083			10:33:24	Yes
Mean:	6.022	6.022	0.080						
SD:	0.177	0.177	0.0024						
%RSD:	2.935	2.935	2.94						

Sequence No.: 22
 Sample ID: CCV
 Analyst:

Autosampler Location: 5
 Date Collected: 1/21/2008 10:34:13 AM
 Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.770	9.770	0.130	0.724	0.135			10:34:36	Yes
2	9.453	9.453	0.126	0.694	0.131			10:35:10	Yes
3	9.614	9.614	0.128	0.691	0.133			10:35:44	Yes
Mean:	9.613	9.613	0.128						
SD:	0.158	0.158	0.0021						

%RSD: 1.649 1.649 1.65

QC value within limits for Se 196.03 Recovery = 96.13%
All analyte(s) passed QC.

Sequence No.: 23

Sample ID: CCB

Analyst:

Autosampler Location: 1

Date Collected: 1/21/2008 10:36:36 AM

Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.173	0.173	0.002	0.043	0.007			10:36:57	Yes
2	0.124	0.124	0.002	0.032	0.007			10:37:31	Yes
3	0.023	0.023	0.000	0.014	0.005			10:38:05	Yes
Mean:	0.107	0.107	0.001						
SD:	0.076	0.076	0.0010						
%RSD:	71.56	71.56	71.56						

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Sequence No.: 24

Sample ID: K0711481-027

Analyst:

Autosampler Location: 19

Date Collected: 1/21/2008 10:38:54 AM

Data Type: Original

Replicate Data: K0711481-027

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	6.551	6.551	0.087	0.460	0.092			10:39:16	Yes
2	6.476	6.476	0.086	0.450	0.091			10:39:50	Yes
3	6.504	6.504	0.087	0.452	0.092			10:40:24	Yes
Mean:	6.511	6.511	0.087						
SD:	0.038	0.038	0.0005						
%RSD:	0.579	0.579	0.58						

Sequence No.: 25

Sample ID: K0711481-028

Analyst:

Autosampler Location: 20

Date Collected: 1/21/2008 10:41:14 AM

Data Type: Original

Replicate Data: K0711481-028

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	7.436	7.436	0.099	0.529	0.104			10:41:35	Yes
2	7.784	7.784	0.104	0.537	0.109			10:42:09	Yes
3	7.306	7.306	0.097	0.503	0.102			10:42:43	Yes
Mean:	7.509	7.509	0.100						
SD:	0.247	0.247	0.0033						
%RSD:	3.296	3.296	3.30						

Sequence No.: 26

Sample ID: K0711481-029

Analyst:

Autosampler Location: 21

Date Collected: 1/21/2008 10:43:33 AM

Data Type: Original

Replicate Data: K0711481-029

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	5.967	5.967	0.079	0.424	0.084			10:43:54	Yes
2	5.664	5.664	0.075	0.404	0.080			10:44:28	Yes
3	5.769	5.769	0.077	0.415	0.082			10:45:02	Yes
Mean:	5.800	5.800	0.077						

SD: 0.154 0.154 0.0020
 %RSD: 2.651 2.651 2.65

Sequence No.: 27
 Sample ID: K0711481-030
 Analyst:

Autosampler Location: 22
 Date Collected: 1/21/2008 10:45:52 AM
 Data Type: Original

Replicate Data: K0711481-030

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	5.495	5.495	0.073	0.387	0.078			10:46:14	Yes
2	5.324	5.324	0.071	0.378	0.076			10:46:48	Yes
3	5.168	5.168	0.069	0.368	0.074			10:47:23	Yes
Mean:	5.329	5.329	0.071						
SD:	0.164	0.164	0.0022						
%RSD:	3.068	3.068	3.07						

Sequence No.: 28
 Sample ID: K0711481-031
 Analyst:

Autosampler Location: 23
 Date Collected: 1/21/2008 10:48:12 AM
 Data Type: Original

Replicate Data: K0711481-031

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	8.535	8.535	0.114	0.634	0.119			10:48:34	Yes
2	7.162	7.162	0.095	0.480	0.100			10:49:09	Yes
3	6.928	6.928	0.092	0.485	0.097			10:49:43	Yes
Mean:	7.542	7.542	0.100						
SD:	0.868	0.868	0.0115						
%RSD:	11.51	11.51	11.51						

Sequence No.: 29
 Sample ID: K0711481-032
 Analyst:

Autosampler Location: 24
 Date Collected: 1/21/2008 10:50:33 AM
 Data Type: Original

Replicate Data: K0711481-032

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.716	9.716	0.129	0.687	0.134			10:50:55	Yes
2	8.675	8.675	0.115	0.659	0.120			10:51:30	Yes
3	10.02	10.02	0.133	0.687	0.138			10:52:05	Yes
Mean:	9.471	9.471	0.126						
SD:	0.706	0.706	0.0094						
%RSD:	7.453	7.453	7.45						

Sequence No.: 30
 Sample ID: K0711481-033
 Analyst:

Autosampler Location: 25
 Date Collected: 1/21/2008 10:52:55 AM
 Data Type: Original

Replicate Data: K0711481-033

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	5.767	5.767	0.077	0.415	0.082			10:53:18	Yes
2	6.430	6.430	0.086	0.402	0.091			10:53:52	Yes
3	5.382	5.382	0.072	0.373	0.077			10:54:26	Yes
Mean:	5.860	5.860	0.078						
SD:	0.530	0.530	0.0070						
%RSD:	9.045	9.045	9.05						

Sequence No.: 31
Sample ID: K0711481-034
Analyst:

Autosampler Location: 26
Date Collected: 1/21/2008 10:55:17 AM
Data Type: Original

Replicate Data: K0711481-034

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.11	10.11	0.134	0.708	0.140			10:55:40	Yes
2	10.08	10.08	0.134	0.696	0.139			10:56:14	Yes
3	9.846	9.846	0.131	0.677	0.136			10:56:49	Yes
Mean:	10.01	10.01	0.133						
SD:	0.147	0.147	0.0019						
%RSD:	1.463	1.463	1.46						

Sequence No.: 32
Sample ID: K0711481-035
Analyst:

Autosampler Location: 27
Date Collected: 1/21/2008 10:57:40 AM
Data Type: Original

1/21/08

Replicate Data: K0711481-035

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	20.12	20.12	0.268	1.445	0.273			10:58:03	Yes
2	19.75	19.75	0.263	1.415	0.268			10:58:38	Yes
3	20.18	20.18	0.268	1.410	0.273			10:59:12	Yes
Mean:	20.01	20.01	0.266						
SD:	0.231	0.231	0.0031						
%RSD:	1.156	1.156	1.16						

Sequence No.: 33
Sample ID: K0711481-036
Analyst:

Autosampler Location: 28
Date Collected: 1/21/2008 11:00:36 AM
Data Type: Original

Replicate Data: K0711481-036

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	12.58	12.58	0.167	0.914	0.172			11:00:59	Yes
2	12.41	12.41	0.165	0.881	0.170			11:01:33	Yes
3	12.46	12.46	0.166	0.887	0.171			11:02:07	Yes
Mean:	12.48	12.48	0.166						
SD:	0.090	0.090	0.0012						
%RSD:	0.719	0.719	0.72						

Sequence No.: 34
Sample ID: CCV
Analyst:

Autosampler Location: 5
Date Collected: 1/21/2008 11:02:59 AM
Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.916	9.916	0.132	0.728	0.137			11:03:22	Yes
2	9.667	9.667	0.129	0.694	0.134			11:03:56	Yes
3	9.543	9.543	0.127	0.683	0.132			11:04:30	Yes
Mean:	9.709	9.709	0.129						
SD:	0.190	0.190	0.0025						
%RSD:	1.960	1.960	1.96						

QC value within limits for Se 196.03 Recovery = 97.09%
All analyte(s) passed QC.

Sequence No.: 35
Sample ID: CCB
Analyst:

Autosampler Location: 1
Date Collected: 1/21/2008 11:05:22 AM
Data Type: Original

Replicate Data: CCB

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicates and summary statistics.

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Sequence No.: 36
Sample ID: K0711481-037
Analyst:

Autosampler Location: 29
Date Collected: 1/21/2008 11:07:42 AM
Data Type: Original

Replicate Data: K0711481-037

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicates with concentration warnings and summary statistics.

Sample concentration is greater than that of the highest standard.

Sequence No.: 37
Sample ID: K0711481-038
Analyst:

Handwritten signature: A 1/21/08

Autosampler Location: 30
Date Collected: 1/21/2008 11:10:39 AM
Data Type: Original

Replicate Data: K0711481-038

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicates with concentration warnings and summary statistics.

Sample concentration is greater than that of the highest standard.

Sequence No.: 38
Sample ID: K0711481-021A
Analyst:

Autosampler Location: 31
Date Collected: 1/21/2008 11:13:37 AM
Data Type: Original

Replicate Data: K0711481-021A

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	15.05	15.05	0.200	1.052	0.205			11:14:01	Yes
2	14.81	14.81	0.197	1.004	0.202			11:14:36	Yes
3	14.55	14.55	0.193	1.008	0.199			11:15:10	Yes
Mean:	14.80	14.80	0.197						
SD:	0.249	0.249	0.0033						
%RSD:	1.681	1.681	1.68						

Sequence No.: 39

Sample ID: CCV

Analyst:

Autosampler Location: 5

Date Collected: 1/21/2008 11:16:02 AM

Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.05	10.05	0.134	0.730	0.139			11:16:25	Yes
2	9.833	9.833	0.131	0.696	0.136			11:16:59	Yes
3	9.652	9.652	0.128	0.692	0.133			11:17:34	Yes
Mean:	9.845	9.845	0.131						
SD:	0.199	0.199	0.0027						
%RSD:	2.026	2.026	2.03						

QC value within limits for Se 196.03 Recovery = 98.45%

All analyte(s) passed QC.

Sequence No.: 40

Sample ID: CCB

Analyst:

Autosampler Location: 1

Date Collected: 1/21/2008 11:18:26 AM

Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.303	0.303	0.004	0.051	0.009			11:18:47	Yes
2	0.273	0.273	0.004	0.034	0.009			11:19:21	Yes
3	0.057	0.057	0.001	0.025	0.006			11:19:55	Yes
Mean:	0.211	0.211	0.003						
SD:	0.134	0.134	0.0018						
%RSD:	63.72	63.72	63.72						

QC value within limits for Se 196.03 Recovery = Not calculated

All analyte(s) passed QC.

Analysis Begun

Logged In Analyst: ACOMET10

Technique: AA FIAS-Flame

Spectrometer Model: AAnalyst 200, S/N 200S5061701

Autosampler Model: AS-90

Sample Information File: C:\data-AA\ACOMET10\Sample Information\012108-Se.sif

Batch ID: 012108-Se

Results Data Set: 012108-Se

Results Library: R:\ICP\WIP\DATA\K-FLAA-02\Results.mdb

Sequence No.: 41

Sample ID: K0711481-022 1/10

Analyst:

Autosampler Location:

Date Collected: 1/21/2008 11:21:07 AM

Data Type: Original

Replicate Data: K0711481-022 1/10

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.53	10.53	0.140	0.715	0.145			11:21:23	Yes

2	10.23	10.23	0.136	0.691	0.141			11:21:57	Yes
3	10.08	10.08	0.134	0.691	0.139			11:22:31	Yes
Mean:	10.28	10.28	0.137						
SD:	0.230	0.230	0.0031						
%RSD:	2.240	2.240	2.24						

Sequence No.: 42

Sample ID: K0711481-035 1/10

Analyst:

Autosampler Location:

Date Collected: 1/21/2008 11:23:33 AM

Data Type: Original

Replicate Data: K0711481-035 1/10

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	11.74	11.74	0.156	0.828	0.161			11:23:49	Yes
2	11.55	11.55	0.154	0.798	0.159			11:24:24	Yes
3	11.41	11.41	0.152	0.797	0.157			11:24:59	Yes
Mean:	11.56	11.56	0.154						
SD:	0.166	0.166	0.0022						
%RSD:	1.436	1.436	1.44						

Sequence No.: 43

Sample ID: K0711481-037 1/20

Analyst:

Autosampler Location:

Date Collected: 1/21/2008 11:25:59 AM

Data Type: Original

Replicate Data: K0711481-037 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	11.39	11.39	0.151	0.804	0.157			11:26:15	Yes
2	11.05	11.05	0.147	0.768	0.152			11:26:50	Yes
3	11.10	11.10	0.148	0.756	0.153			11:27:24	Yes
Mean:	11.18	11.18	0.149						
SD:	0.181	0.181	0.0024						
%RSD:	1.615	1.615	1.61						

Sequence No.: 44

Sample ID: K0711481-038 1/20

Analyst:

Autosampler Location:

Date Collected: 1/21/2008 11:28:26 AM

Data Type: Original

Replicate Data: K0711481-038 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.24	10.24	0.136	0.722	0.141			11:28:43	Yes
2	10.16	10.16	0.135	0.689	0.140			11:29:17	Yes
3	10.66	10.66	0.142	0.687	0.147			11:29:51	Yes
Mean:	10.35	10.35	0.138						
SD:	0.267	0.267	0.0035						
%RSD:	2.579	2.579	2.58						

Sequence No.: 45

Sample ID: CCV

Analyst:

Autosampler Location:

Date Collected: 1/21/2008 11:30:52 AM

Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.861	9.861	0.131	0.727	0.136			11:31:08	Yes
2	9.529	9.529	0.127	0.691	0.132			11:31:42	Yes
3	9.660	9.660	0.128	0.684	0.134			11:32:17	Yes
Mean:	9.683	9.683	0.129						

SD: 0.168 0.168 0.0022
%RSD: 1.730 1.730 1.73

Sequence No.: 46

Sample ID: CCB

Analyst:

Autosampler Location:

Date Collected: 1/21/2008 11:33:18 AM

Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.145	0.145	0.002	0.028	0.007			11:33:35	Yes
2	0.146	0.146	0.002	0.030	0.007			11:34:09	Yes
3	0.163	0.163	0.002	0.025	0.007			11:34:43	Yes
Mean:	0.151	0.151	0.002						
SD:	0.010	0.010	0.0001						
%RSD:	6.722	6.722	6.72						

January 20, 2008

Analytical Report for Service Request No: K0711649

Kathy Tegtmeyer
New Fields Environmental
4720 Walnut St., Suite 200
Boulder, CO 80301

RE: Se in Tissue

Dear Kathy:

Enclosed are the results of the samples submitted to our laboratory on December 12, 2007. For your reference, these analyses have been assigned our service request number K0711649.

All analyses were performed according to our laboratory's quality assurance program. Where applicable, the methods cited conform to the Methods Update Rule (effective 4/11/2007), which relates to the use of analytical methods for the drinking water and waste water programs. The test results meet requirements of the NELAC standards. Exceptions are noted in the case narrative report where applicable. All results are intended to be considered in their entirety, and Columbia Analytical Services, Inc. (CAS) is not responsible for use of less than the complete report. Results apply only to the items submitted to the laboratory for analysis and individual items (samples) analyzed, as listed in the report.

Please call if you have any questions. My extension is 3316. You may also contact me via Email at JChristian@caslab.com.

Respectfully submitted,

Columbia Analytical Services, Inc.



Jeff Christian
Laboratory Director

JC/lb

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Acronyms

ASTM	American Society for Testing and Materials
A2LA	American Association for Laboratory Accreditation
CARB	California Air Resources Board
CAS Number	Chemical Abstract Service registry Number
CFC	Chlorofluorocarbon
CFU	Colony-Forming Unit
DEC	Department of Environmental Conservation
DEQ	Department of Environmental Quality
DHS	Department of Health Services
DOE	Department of Ecology
DOH	Department of Health
EPA	U. S. Environmental Protection Agency
ELAP	Environmental Laboratory Accreditation Program
GC	Gas Chromatography
GC/MS	Gas Chromatography/Mass Spectrometry
LUFT	Leaking Underground Fuel Tank
M	Modified
MCL	Maximum Contaminant Level is the highest permissible concentration of a substance allowed in drinking water as established by the USEPA.
MDL	Method Detection Limit
MPN	Most Probable Number
MRL	Method Reporting Limit
NA	Not Applicable
NC	Not Calculated
NCASI	National Council of the Paper Industry for Air and Stream Improvement
ND	Not Detected
NIOSH	National Institute for Occupational Safety and Health
PQL	Practical Quantitation Limit
RCRA	Resource Conservation and Recovery Act
SIM	Selected Ion Monitoring
TPH	Total Petroleum Hydrocarbons
tr	Trace level is the concentration of an analyte that is less than the PQL but greater than or equal to the MDL.

Inorganic Data Qualifiers

- * The result is an outlier. See case narrative.
- # The control limit criteria is not applicable. See case narrative.
- B The analyte was found in the associated method blank at a level that is significant relative to the sample result.
- E The result is an estimate amount because the value exceeded the instrument calibration range.
- J The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
 - i The MRL/MDL has been elevated due to a matrix interference.
- X See case narrative.

Metals Data Qualifiers

- # The control limit criteria is not applicable. See case narrative.
- B The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- E The percent difference for the serial dilution was greater than 10%, indicating a possible matrix interference in the sample.
- M The duplicate injection precision was not met.
- N The Matrix Spike sample recovery is not within control limits. See case narrative.
- S The reported value was determined by the Method of Standard Additions (MSA).
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
- W The post-digestion spike for furnace AA analysis is out of control limits, while sample absorbance is less than 50% of spike absorbance.
 - i The MRL/MDL has been elevated due to a matrix interference.
- X See case narrative.
- * The duplicate analysis not within control limits. See case narrative.
- + The correlation coefficient for the MSA is less than 0.995.

Organic Data Qualifiers

- * The result is an outlier. See case narrative.
- # The control limit criteria is not applicable. See case narrative.
- A A tentatively identified compound, a suspected aldol-condensation product.
- B The analyte was found in the associated method blank at a level that is significant relative to the sample result.
- C The analyte was qualitatively confirmed using GC/MS techniques, pattern recognition, or by comparing to historical data.
- D The reported result is from a dilution.
- E The result is an estimate amount because the value exceeded the instrument calibration range.
- J The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- N The result is presumptive. The analyte was tentatively identified, but a confirmation analysis was not performed.
- P The GC or HPLC confirmation criteria was exceeded. The relative percent difference is greater than 40% between the two analytical results (25% for CLP Pesticides).
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
 - i The MRL/MDL has been elevated due to a chromatographic interference.
- X See case narrative.

Additional Petroleum Hydrocarbon Specific Qualifiers

- F The chromatographic fingerprint of the sample matches the elution pattern of the calibration standard.
- L The chromatographic fingerprint of the sample resembles a petroleum product, but the elution pattern indicates the presence of a greater amount of lighter molecular weight constituents than the calibration standard.
- H The chromatographic fingerprint of the sample resembles a petroleum product, but the elution pattern indicates the presence of a greater amount of heavier molecular weight constituents than the calibration standard.
- O The chromatographic fingerprint of the sample resembles an oil, but does not match the calibration standard.
- Y The chromatographic fingerprint of the sample resembles a petroleum product eluting in approximately the correct carbon range, but the elution pattern does not match the calibration standard.
- Z The chromatographic fingerprint does not resemble a petroleum product.

Columbia Analytical Services, Inc.
Kelso, WA
State Certifications, Accreditations, and Licenses

Program	Number
Alaska DEC UST	UST-040
Arizona DHS	AZ0339
Arkansas - DEQ	88-0637
California DHS	2286
Colorado DPHE	-
Florida DOH	E87412
Hawaii DOH	-
Idaho DHW	-
Indiana DOH	C-WA-01
Louisiana DEQ	3016
Louisiana DHH	LA050010
Maine DHS	WA0035
Michigan DEQ	9949
Minnesota DOH	053-999-368
Montana DPHHS	CERT0047
Nevada DEP	WA35
New Jersey DEP	WA005
New Mexico ED	-
North Carolina DWQ	605
Oklahoma DEQ	9801
Oregon - DHS	WA200001
South Carolina DHEC	61002
Utah DOH	COLU
Washington DOE	C1203
Wisconsin DNR	998386840
Wyoming (EPA Region 8)	-



Case Narrative

COLUMBIA ANALYTICAL SERVICES, INC.

Client: New Fields Environmental
Project: Tissue - Se
Sample Matrix: Tissue

Service Request No.: K0711649
Date Received: 12/12/07

CASE NARRATIVE

All analyses were performed consistent with the quality assurance program of Columbia Analytical Services, Inc. (CAS). This report contains analytical results for samples designated for Tier III validation deliverables including summary forms and all of the associated raw data for each of the analyses. When appropriate to the method, method blank results have been reported with each analytical test.

Sample Receipt

Tissue samples were received for analysis at Columbia Analytical Services on 12/12/07. The samples were received in good condition and consistent with the accompanying chain of custody form. The samples were stored frozen at -20°C upon receipt at the laboratory.

Total Metals

General Comments:

The samples were freeze-dried to determine moisture and to allow complete homogenization of the dry material. The dried material was milled to a fine meal, and then sub-sampled for digestion. A thorough digestion was performed prior to instrumental analysis to convert all Selenium species to Selenate. Prior to hydride formation, the valence was adjusted by reduction to Selenite.

No anomalies associated with the analysis of these samples were observed.

Approved by _____



Date

1/18/08

Chain of Custody Documentation

Chain of Custody

Page of



Project Contact: Sean Covington/Kathy Tegtmeyer
 Courier/Airbill: PO 0442-004-900.70

Shipped to: Columbia Analytical Services, Inc.

1317 South 13th Ave
 Kelso, WA 98626
 Telephone: (360) 430-7733

*COCTAPE # 40066
 Initial: Y/N*

COC #:

4720 Walnut St, Suite 200
 Boulder, CO 80301
 Phone: 303-442-0267
 Fax: 303-442-3679

Sample ID	Sample Date	Sample Time	Matrix	Tot/Diss	Analysis	Preservative	Lab QC	Comments
SM1007-SNFH-FT0050	10/25/2007		Fish Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNFH-FT0051	10/25/2007		Fish Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNFH-FT0052	10/25/2007		Fish Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNFH-FT0053	10/25/2007		Fish Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNFH-FT0054	10/25/2007		Fish Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNFH-FT0055	10/25/2007		Fish Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNFH-FT0056	10/25/2007		Fish Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNFH-FT0057	10/25/2007		Fish Tissue	Tot	Selenium, % Solids	Dry Ice		

Total Number of Containers: 8 Individual Lines Reflect Single Containers, Except for Aqueous Analyses Assigned as Laboratory QC

Relinquished by: *MSM*
 Date/Time: *12/11/07 15:55*

Received by: *Kary Black*
 Date/Time: *12/11/07 10:30*

LAB USE ONLY - Sample condition on Receipt

**Columbia Analytical Services, Inc.
Cooler Receipt and Preservation Form**

PC JC

Client / Project: ENS12 Service Request **K07** 11649

Received: 12-12-07 Opened: 12-12-07 By: PD

1. Samples were received via? US Mail **Fed Ex** UPS DHL GH GS PDX Courier Hand Delivered
2. Samples were received in: (circle) **Cooler** Box Envelope Other _____ NA
3. Were custody seals on coolers? NA Y N If yes, how many and where? Front
If present, were custody seals intact? Y N If present, were they signed and dated? Y N
4. Is shipper's air-bill filed? If not, record air-bill number: 7924 7001 7231 NA Y N
5. Temperature of cooler(s) upon receipt (°C): -17.3
Temperature Blank (°C): _____
6. If applicable, list Chain of Custody Numbers: _____
7. Were custody papers properly filled out (ink, signed, etc.)? NA Y N
8. Packing material used. Inserts Bubble Wrap Gel Packs Wet Ice Sleeves **Other** Dry Ice
9. Did all bottles arrive in good condition (unbroken)? Indicate in the table below. NA Y N
10. Were all sample labels complete (i.e analysis, preservation, etc.)? Y N
11. Did all sample labels and tags agree with custody papers? Indicate in the table below Y N
12. Were the correct types of bottles used for the tests indicated? NA Y N
13. Were all of the preserved bottles received at the lab with the appropriate pH? Indicate in the table below NA Y N
14. Were VOA vials and 1631 Mercury bottles checked for absence of air bubbles? Indicate in the table below. NA Y N
15. Are CWA Microbiology samples received with >1/2 the 24hr. hold time remaining from collection? NA Y N
16. Was C12/Res negative? NA Y N

Sample ID on Bottle	Sample ID on COC	Sample ID on Bottle	Sample ID on COC

Sample ID	Bottle Count	Bottle Type	Out of Temp	Head-space	Broken	pH	Reagent	Volume added	Reagent Lot Number	Initials

Additional Notes, Discrepancies, & Resolutions: _____

Total Solids

COLUMBIA ANALYTICAL SERVICES, INC.

Analytical Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0711649
Date Collected: 10/25/07
Date Received: 12/12/07

Solids, Total

Prep Method: NONE
Analysis Method: Freeze Dry
Test Notes:

Units: PERCENT
Basis: Wet

Sample Name	Lab Code	Date Analyzed	Result	Result Notes
SM1007-SNFH-FT0050	K0711649-001	12/27/08	29.7	
SM1007-SNFH-FT0051	K0711649-002	12/27/08	26.0	
SM1007-SNFH-FT0052	K0711649-003	12/27/08	31.7	
SM1007-SNFH-FT0053	K0711649-004	12/27/08	32.3	
SM1007-SNFH-FT0054	K0711649-005	12/27/08	32.1	
SM1007-SNFH-FT0055	K0711649-006	12/27/08	27.6	
SM1007-SNFH-FT0056	K0711649-007	12/27/08	32.0	
SM1007-SNFH-FT0057	K0711649-008	12/27/08	32.8	

COLUMBIA ANALYTICAL SERVICES, INC.

QA/QC Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0711649
Date Collected: 10/25/07
Date Received: 12/12/07
Date Extracted: NA
Date Analyzed: 12/27/08

Duplicate Summary
Total Metals

Sample Name: SM1007-SNFH-FT0050
Lab Code: K0711649-001D
Test Notes:

Units: PERCENT
Basis: Wet

Analyte	Prep Method	Analysis Method	Sample Result	Duplicate Sample Result	Average	Relative Percent Difference	Result Notes
Solids, Total	NA	Freeze Dry	29.7	30.3	30.0	2	

Service Request #: K0711649
 Analysis For: Freeze Dried Solids

Extr/Prep Batch
 KP0613321

Lab Code	Wet Weight (g)	Tare (g)	Tare + Dry Wt.(g)	Dry Weight (g)	% Total Solids
K0711649-01	25.150	78.810	86.268	7.458	29.7
-01D	23.630	79.400	86.555	7.155	30.3
-02	24.378	78.938	85.286	6.348	26.0
-03	23.331	78.509	85.895	7.386	31.7
-04	23.314	78.618	86.155	7.537	32.3
-05	24.616	78.407	86.306	7.899	32.1
-06	22.450	79.103	85.295	6.192	27.6
-07	23.480	79.136	86.652	7.516	32.0
-08	23.573	79.143	86.884	7.741	32.8
LAB 12/27/07					

Time In: 4:00p.m. 12/31/07 Time Out: 9:00a.m. 1/2/08
 Comments: balance, 21B

\bar{x} = RPD =

Analyst: Eric R. R. / Angela Black Date: 12-27-07
 Reviewed By: [Signature] Date: 1/9/08

Sample Number(s): As Listed	Service Request Number(s): K0711649
--------------------------------	--

TISSUE COMPOSITION DATA

Laboratory ID	Weight (g)	Tare (g)			Matrix	Length
K0711649-01(1)	820.34	470.98			Whole body fish	
-01(2)	573.07	468.88				
-02	826.00	470.75				
-03(1)	814.27	479.24				
-03(2)	759.17	477.41				
-04(1)	888.06	478.65				
-04(2)	822.69	477.74				
-04(3)	789.00	485.36				
-05(1)	824.07	471.08				
-05(2)	834.73	486.42				
-05(3)	528.17	486.71				
-06	842.38	470.53				
-07(1)	713.20	479.15				
-07(2)	462.17	485.80				
-08(1)	828.11	479.25				
-08(2)	618.22	484.78				
LAB 12/27/07						

Comments: _____

Analyst: <i>Angela Brack</i>	Date: 12-27-07
Reviewed: <i>[Signature]</i>	Date: 1/9/08

Metals

Columbia Analytical Services

- Cover Page -
INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental
Project Name: Se in Tissue
Project No.:

Service Request: K0711649

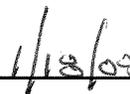
<u>Sample Name:</u>	<u>Lab Code:</u>
<u>SM1007-SNFH-FT0050</u>	<u>K0711649-001</u>
<u>SM1007-SNFH-FT0050D</u>	<u>K0711649-001D</u>
<u>SM1007-SNFH-FT0050S</u>	<u>K0711649-001S</u>
<u>SM1007-SNFH-FT0051</u>	<u>K0711649-002</u>
<u>SM1007-SNFH-FT0052</u>	<u>K0711649-003</u>
<u>SM1007-SNFH-FT0053</u>	<u>K0711649-004</u>
<u>SM1007-SNFH-FT0054</u>	<u>K0711649-005</u>
<u>SM1007-SNFH-FT0055</u>	<u>K0711649-006</u>
<u>SM1007-SNFH-FT0056</u>	<u>K0711649-007</u>
<u>SM1007-SNFH-FT0057</u>	<u>K0711649-008</u>
<u>Method Blank</u>	<u>K0711649-MB</u>

Comments:

Approved By: _____



Date: _____



Metals

- 1 -

INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental Service Request: K0711649
Project No.: NA Date Collected:
Project Name: Se in Tissue Date Received:
Matrix: TISSUE Units: mg/Kg
Basis: DRY

Sample Name: Method Blank Lab Code: K0711649-MB

Analyte	Analysis Method	MRL	MDL	Dil. Factor	Date Extracted	Date Analyzed	Result	C	Q
Selenium	7742	0.10	0.02	2.0	01/07/08	01/18/08	0.06	B	

% Solids: 100.0

Comments:

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium	10.0	9.52	95	10.0	9.83	98	9.58	96	7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium				10.0	9.16	92	9.37	94	7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium				10.0	9.46	95			7742

Metals

- 2b -

CRDL STANDARD FOR AA AND ICP

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Project Name: Se in Tissue

Concentration Units: ug/L

Analyte	CRDL Standard for AA			CRDL Standard for ICP				
	True	Found	%R	Initial		Final		
				True	Found	%R	Found	%R
Selenium	0.5	0.67	134.0					

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Project Name: Se in Tissue

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)		Continuing Calibration Blank (ug/L)						Method
		C	1	C	2	C	3	C	
Selenium	0.3	B	0.2	B	0.3	B	0.1	B	7742

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Project Name: Se in Tissue

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)	C	Continuing Calibration Blank (ug/L)						Method
			1	C	2	C	3	C	
Selenium			0.3	B	0.2	B			7742

Metals

- 5A -

SPIKE SAMPLE RECOVERY

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Units: MG/KG

Project Name: Se in Tissue

Basis: DRY

Matrix: TISSUE

Sample Name: SM1007-SNFH-FT0050S

Lab Code: K0711649-001S

Analyte	Control Limit %R	Spike Result	C	Sample Result	C	Spike Added	%R	Q	Method
Selenium	65 - 124	16.9		3.6		15.09	88.1		7742

An empty field in the Control Limit column indicates the control limit is not applicable

Metals

- 6 -

DUPLICATES

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Units: MG/KG

Project Name: Se in Tissue

Basis: DRY

Matrix: TISSUE

Sample Name: SM1007-SNEH-FT0050D

Lab Code: K0711649-001D

Analyte	Control Limit	Sample (S)	C	Duplicate (D)	C	RPD	Q	Method
Selenium	30	3.6		3.8		5.4		7742

An empty field in the Control Limit column indicates the control limit is not applicable.

Metals

- 7 -

LABORATORY CONTROL SAMPLE

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Project Name: Se in Tissue

Aqueous LCS Source:

Solid LCS Source:

Analyte	Aqueous (ug/L)			Solid (mg/kg)					
	True	Found	%R	True	Found	C	Limits	%R	
Selenium				7.06	6.00		75.5	128	85

Metals

- 10 -

DETECTION LIMITS

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Project Name: Se in Tissue

ICP/ICP-MS ID #:

GFAA ID #: K-FLAA-02

AA ID #:

Analyte	Wave-length (nm)	Back-ground	MRL ug/L	MDL ug/L	M
Selenium			0.5	0.1	F

Comments: _____

Metals
-13-
PREPARATION LOG

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Project Name: Se in Tissue

Method: F

Sample ID	Preparation Date	Initial Volume	Final Volume (mL)
K0711649-001	1/7/08	0.3150	30.0
K0711649-001D	1/7/08	0.3270	30.0
K0711649-001S	1/7/08	0.3320	30.0
K0711649-002	1/7/08	0.3180	30.0
K0711649-003	1/7/08	0.3040	30.0
K0711649-004	1/7/08	0.3440	30.0
K0711649-005	1/7/08	0.3830	30.0
K0711649-006	1/7/08	0.3160	30.0
K0711649-007	1/7/08	0.3110	30.0
K0711649-008	1/7/08	0.3080	30.0
K0711649-MB	1/7/08	0.3000	30.0
LCSS	1/7/08	0.31	30.0

Metals
- 14 -
ANALYSIS RUN LOG

Client: New Fields Environmental
Project No.: NA
Project Name: Se in Tissue

Service Request: K0711649

Instrument ID Number: K-FLAA-02

Method: F

Start Date: 01/18/08

End Date: 01/18/08

Sample No.	D/F	Time	% R	Analytes																							
				A L	S B	A S	B A	B E	C D	C A	C R	C O	C U	F E	P B	M G	M N	H G	N I	K	S E	A G	N A	T L	V	Z N	C N
ZZZZZZ	1	09:20																									
ZZZZZZ	1	09:22																									
ZZZZZZ	1	09:25																									
ZZZZZZ	1	09:27																									
ZZZZZZ	1	09:29																									
ZZZZZZ	1	09:32																									
ZZZZZZ	1	09:34																									
ZZZZZZ	1	09:37																									
ZZZZZZ	1	09:40																									
CAL BLK	1	09:42																			X						
STD 0.5	1	09:45																			X						
STD 1.0	1	09:47																			X						
STD 5.0	1	09:49																			X						
STD 10.0	1	09:52																			X						
STD 15.0	1	09:54																			X						
ICV1	1	09:56																			X						
ICB1	1	09:59																			X						
CRA	1	10:01																			X						
CCV1	1	10:04																			X						
CCB1	1	10:06																			X						
ZZZZZZ	1	10:08																									
ZZZZZZ	1	10:11																									
CCV2	1	10:13																			X						
CCB2	1	10:15																			X						
K0711649-MB	2	10:23																			X						
LCSS	20	10:25																			X						
K0711649-001	8	10:28																			X						
K0711649-001D	8	10:31																			X						
K0711649-001S	40	10:33																			X						
K0711649-002	8	10:36																			X						
K0711649-003	8	10:39																			X						
K0711649-004	8	10:41																			X						

* - Denotes additional elements (other than the standard CLP elements) are represented on another Form 14

Metals
- 14 -
ANALYSIS RUN LOG

Client: New Fields Environmental

Service Request: K0711649

Project No.: NA

Project Name: Se in Tissue

Instrument ID Number: K-FLAA-02

Method: F

Start Date: 1/18/08

End Date: 1/18/08

Sample No.	D/F	Time	% R	Analytes																						
				A L	S B	A S	B A	B E	C D	C A	C R	C O	C U	F E	P B	M G	M N	H G	N I	K	S E	A G	N A	T L	V	Z N
K0711649-005	8	10:44																	X							
K0711649-006	8	10:46																	X							
CCV3	1	10:49																	X							
CCB3	1	10:51																	X							
K0711649-007	8	10:54																	X							
K0711649-008	8	10:57																	X							
ZZZZZZ	8	10:59																								
ZZZZZZ	8	12:15																								
ZZZZZZ	8	12:17																								
ZZZZZZ	8	12:20																								
ZZZZZZ	8	12:22																								
ZZZZZZ	8	12:25																								
ZZZZZZ	8	12:28																								
ZZZZZZ	8	12:30																								
CCV4	1	12:33																	X							
CCB4	1	12:35																	X							
ZZZZZZ	8	12:37																								
ZZZZZZ	8	12:40																								
ZZZZZZ	8	12:42																								
CCV5	1	12:45																	X							
CCB5	1	12:47																	X							

* - Denotes additional elements (other than the standard CLP elements) are represented on another Form 14

Columbia Analytical Services
Metals Tissue Digestion Sheet

Service Request Number(s) : K0711649 redigest

Star Lims Run No.: _____ Analysis for: ICP ICP-MS GFAA
 Method : Tissue other: Se hydride

Sample	Initial Weight (g)	freeze Dry	Wet	Final Volume (ml)	Matrix
<u>KB</u>			<u>X</u>	<u>30mls</u>	<u>157-HND3</u>
<u>DD1+</u>	<u>0.311</u>		<u>I</u>		
<u>K0711649-01</u>	<u>0.315</u>	<u>X</u>			
- 018	<u>0.327</u>				
- 019	<u>0.332</u>				
- 02	<u>0.318</u>				
- 03	<u>0.304</u>				
- 04	<u>0.344</u>				
- 05	<u>0.383</u>				
- 06	<u>0.314</u>				
- 07	<u>0.311</u>				
- 08	<u>0.308</u>				
<u>K0711649/08</u>					

Time Digestion Started: 4:20p.m. 1/16/08 Oven Temp: 102°C Time Digestion Ended: 7:00a.m. 1/18/08

Lot # Acids Used: HNO3 MS11-64-C

Oven Temp: 103°C

LCS: Dorm-2, Dolt-3

Balance I.D.: 10aner

- QCP CICV-1, MET1-63-A, _____ mls. added
- QCP CICV-2, MET1-63-B, _____ mls. added
- QCP CICV-3, MET1-59-J, _____ mls. Added
- SS6, MET1-62-A, _____ mls. Added

SPIKE INFO

- SS1-MET1-61-R, 0.3 mls added
- SS5-MET1-61-P, 0.05 mls added
- SS6-MET1-62-A, 0.05 mls added

Additional spikes: _____

Comments: _____

Analyst <u>Angela Brack</u>	Date <u>1/16/08</u>
Reviewer <u>SMA</u>	Date <u>1/18/08</u>

TissueDig.xls
1/7/2008

METALS SPIKE FORM

Service Request # K0711049 redigxst
 Q.C. Sample # K0711049-1

Circle type of digest: GFAA ICP FAA ICP-MS Other: _____ Initials / Date: AB 1/11/08
 Circle type of sample: Soil Water Misc. Sludge Oil Other: Tissue

Solution Name	Element	mls of 1000ppm Solution	Final Volume	Source	Lot#	Exp. Date	Solution Conc. mg/L	Enter mls Added
SS1-MET1-61-R	HNO3	50.0	1000ml	JT Baker	E17044	-	-	0.3
	Al	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	200	
	Ag	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Ba	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	200	
	Be	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Cd	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Co	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Cr	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	20	
	Cu	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	25	
	Fe	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	100	
	Pb	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Mn	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Ni	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Sb	50*	1000ml	MET1-53-A	619312	3/7/2008	50	
V	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50		
Zn	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50		
SS4-MET1-62-J	HNO3	25.0	500ml	JT BAKER	A48046	-	-	Expires:2/1/08
	As	2.0	500ml	MET1-57-Z	Z-AS02032	7/1/2008	4	
	Cd	2.0	500ml	MET1-57-M	Z-CD02004	7/1/2008	4	
	Pb	2.0	500ml	MET1-57-R	A2-PB02135	7/1/2008	4	
	Se	2.0	500ml	MET1-57-K	Z-SE01120	7/1/2008	4	
	Tl	2.0	500ml	MET1-54-M	Z-TL01097	2/1/2008	4	
	Cu	2.0	500ml	MET1-55-O	Z-CU02084	5/1/2008	4	
SS5-MET1-61-P	HNO3	25.0	500ml	JT BAKER	E17044	-	-	Expires:2/1/08
	As	50.0	500ml	MET1-57-Z	Z-AS02032	7/1/2008	100	
	Se	50.0	500ml	MET1-59-C	Z-SE01120	8/1/2008	100	
	Tl	50.0	500ml	MET1-54-M	Z-TL01097	2/1/2008	100	
SS6-MET1-62-A	HNO3	25	500ml	JT BAKER	E27027	-	-	Expires: 2/1/08
	B	50	500ml	MET1-59-A	715006	1/2/2009	100	
	Mo	50	500ml	MET1-54-K	Z-MO02012	2/1/2008	100	

GFLCSW (MET1-60-A)	HNO3	10.0	1000ml	JT BAKER	E01042	-	-	Expires: 3/1/08
	As, Pb, Se, Tl	5.0	1000ml	QCP-CICV-3	Z-CICP19048	3/1/2008	2.5	
	Cd	-	-	QCP-CICV-3	Z-CICP19048	3/1/2008	1.25	
	Cu	2.5	1000ml	MET-55-O	Z-CU02084	5/1/2008	2.5	
QCP-CICV-1 (MET1-63-A)	Ca, Mg, Na, K	no dilution	-	IV	A2-MEB236021	1/1/2009	2500	Expires:1/1/09
	Al, Ba	no dilution	-	IV	A2-MEB236021	1/1/2009	1000	
	Fe	no dilution	-	IV	A2-MEB236021	1/1/2009	500	
	Co, Mn, Ni, V, Zn	no dilution	-	IV	A2-MEB236021	1/1/2009	250	
	Cu, Ag	no dilution	-	IV	A2-MEB236021	1/1/2009	125	
	Cr	no dilution	-	IV	A2-MEB236021	1/1/2009	100	
	Be	no dilution	-	IV	A2-MEB236021	1/1/2009	25	
QCP-CICV-2 (MET1-63-B)	Sb	no dilution	-	IV	Z-CICP19033	1/1/2009	500	Expires: 1/1/09
QCP-CICV-3 (MET1-59-J)	As, Pb, Se, Tl	no dilution	-	IV	Z-CICP19048	9/1/2008	500	Expires: 9/1/08
	Cd	no dilution	-	IV	Z-CICP19048	9/1/2008	250	

* Denotes volume of 1000 ppm stock standard.

Element	mls of	ppm	Source	Lot# / Lab Code	Exp. Date

Element Analyzed Se Hydride Instrument K-FLAA-2
Service Request # K11649

Batch QC SR's # _____

Calibration Std. AA1-8-A

Starlims # 104556

Run # 011808-Se

Hydride Data Review Form

	Yes	No	NA
1. ICV within 10% of true Value	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
2. Calibration data included	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
3. CCV's in control	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
4. CCB's and/or ICB's below MRL	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
5. All reported Results within Cal. Range	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
6. All Calculations are Correct	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Comments

Sample quantified via single point MSA.

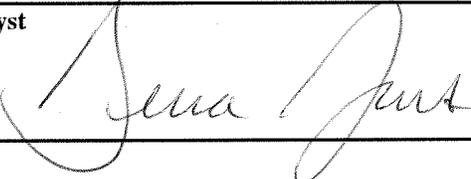
Primary Reviewed by *A* Date *1/18/08*
Secondary Reviewed by *JPB* Date *1/18/08*

COLUMBIA ANALYTICAL SERVICES, INC.
GFAA Run Log

Method: (Circle Method Used) 7742 7062 Other: _____ Element: As Se	Service Request # :
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SAMPLE NUMBER	Dilution Factor	Measured (µg/L)	Recoveries (ICV, CCV, CRA, LCS, Matrix Spk.)	Comments
ICV	-	9.517	95%	
ICB	-	0.281		
CRA	-	0.671	134%	
CCV	-	9.829	98%	
CCB	-	0.207		
K0711649-001	1/2	7.153		1/18/08 Rerun
K0711649-001A	1/2	8.646		1/18/08 Rerun
CCV	-	9.581	96%	
CCB	-	0.301		
K0711649-MB	1/2	0.310		
LCSS DOLT 1/20	1/2+1/10	3.000	85%	
K0711649-001 1/4	1/2+1/4	2.085		X= 4.681
K0711649-001D 1/4	1/2+1/4	2.267		X= 5.159
K0711649-001S 1/20	1/2+1/20	3.720		X= 4.688
K0711649-002 1/4	1/2+1/4	2.355		X= 5.462
K0711649-003 1/4	1/2+1/4	2.085		X= 4.652
K0711649-004 1/4	1/2+1/4	2.485		X= 6.091
K0711649-005 1/4	1/2+1/4	2.143		X= 4.771
K0711649-006 1/4	1/2+1/4	2.100		X= 4.129
CCV	-	9.158	92%	
CCB	-	0.129		
K0711649-007 1/4	1/2+1/4	1.783		X= 3.439
K0711649-008 1/4	1/2+1/4	1.765		1/18/08 X= 3.209
K0711649-001A 1/4	1/2+1/4	2.590		1/18/08 Rerun
K0711649-001A 1/4	1/2+1/4	4.312	45%	
K0711649-001DA 1/4	1/2+1/4	4.464	44%	
K0711649-001SA 1/20	1/2+1/20	7.688	79%	
K0711649-002A 1/4	1/2+1/4	4.511	43%	
K0711649-003A 1/4	1/2+1/4	4.326	45%	
K0711649-004A 1/4	1/2+1/4	4.525	41%	

True Values/QC Limits:	LCSW	Water Spike	LCSS (ERA D045540)	Soil Spike
Arsenic:	2500ppb (80-120%)	1000ppb (80-120%)	146.0mg/kg (80-120%)	1000ppb (80-120%)
Selenium	2500ppb (71-122%)	1000ppb (65-122%)	73.0mg/kg (61-142%)	1000ppb (65-124%)

Analyst 	Date: 1/18/08	Page Number: 1
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Sample ID: Std 5.0
Analyst:

Date Collected: 1/18/2008 9:49:47 AM
Data Type: Original

Replicate Data: Std 5.0

Repl #	SampleConc ug/L	StdConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[5.0]	0.070	0.380	0.075			09:50:10	Yes
2		[5.0]	0.067	0.347	0.072			09:50:44	Yes
3		[5.0]	0.066	0.354	0.071			09:51:19	Yes
Mean:		[5.0]	0.068						
SD:		0.0	0.0023						
%RSD:		0.0	3.36						

Standard number 3 applied. [5.0]
Correlation Coef.: 0.999873 Slope: 0.01350 Intercept: 0.00000

Sequence No.: 5
Sample ID: Std 10.0
Analyst:

Autosampler Location: 5
Date Collected: 1/18/2008 9:52:10 AM
Data Type: Original

Replicate Data: Std 10.0

Repl #	SampleConc ug/L	StdConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[10.0]	0.121	0.669	0.125			09:52:33	Yes
2		[10.0]	0.114	0.614	0.119			09:53:07	Yes
3		[10.0]	0.121	0.655	0.126			09:53:42	Yes
Mean:		[10.0]	0.119						
SD:		0.0	0.0039						
%RSD:		0.0	3.28						

Standard number 4 applied. [10.0]
Correlation Coef.: 0.996782 Slope: 0.01224 Intercept: 0.00000

Sequence No.: 6
Sample ID: Std 15.0
Analyst:

Autosampler Location: 6
Date Collected: 1/18/2008 9:54:33 AM
Data Type: Original

Replicate Data: Std 15.0

Repl #	SampleConc ug/L	StdConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[15.0]	0.185	1.004	0.190			09:54:56	Yes
2		[15.0]	0.178	0.950	0.183			09:55:31	Yes
3		[15.0]	0.178	0.957	0.183			09:56:06	Yes
Mean:		[15.0]	0.181						
SD:		0.0	0.0043						
%RSD:		0.0	2.39						

Standard number 5 applied. [15.0]
Correlation Coef.: 0.998698 Slope: 0.01211 Intercept: 0.00000
The calibration curve may not be linear.

Calibration data for Se 196.03

Equation: Linear Through Zero

ID	Mean Signal (Abs)	Entered Conc. ug/L	Calculated Conc. ug/L	Standard Deviation	%RSD
Cal Blk	0.0000	0	0.000	0.00	20.0
Std 0.5	0.0072	0.5	0.592	0.00	13.3
Std 1.0	0.0129	1.0	1.064	0.00	2.8
Std 5.0	0.0676	5.0	5.580	0.00	3.4
Std 10.0	0.1187	10.0	9.799	0.00	3.3
Std 15.0	0.1805	15.0	14.907	0.00	2.4

Correlation Coef.: 0.998698 Slope: 0.01211 Intercept: 0.00000

Sequence No.: 7
 Sample ID: ICV
 Analyst:

Autosampler Location: 7
 Date Collected: 1/18/2008 9:56:58 AM
 Data Type: Original

Replicate Data: ICV

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.849	9.849	0.119	0.669	0.124			09:57:22	Yes
2	9.405	9.405	0.114	0.618	0.119			09:57:56	Yes
3	9.296	9.296	0.113	0.610	0.117			09:58:30	Yes
Mean:	9.517	9.517	0.115						
SD:	0.293	0.293	0.0036						
%RSD:	3.080	3.080	3.08						

QC value within limits for Se 196.03 Recovery = 95.17%
 All analyte(s) passed QC.

Sequence No.: 8
 Sample ID: ICB
 Analyst:

Autosampler Location: 1
 Date Collected: 1/18/2008 9:59:22 AM
 Data Type: Original

Replicate Data: ICB

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.259	0.259	0.003	0.037	0.008			09:59:44	Yes
2	0.403	0.403	0.005	0.044	0.010			10:00:18	Yes
3	0.182	0.182	0.002	0.029	0.007			10:00:52	Yes
Mean:	0.281	0.281	0.003						
SD:	0.112	0.112	0.0014						
%RSD:	39.86	39.86	39.86						

QC value within limits for Se 196.03 Recovery = Not calculated
 All analyte(s) passed QC.

Sequence No.: 9
 Sample ID: CRA
 Analyst:

Autosampler Location: 2
 Date Collected: 1/18/2008 10:01:41 AM
 Data Type: Original

Replicate Data: CRA

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.617	0.617	0.007	0.047	0.012			10:02:03	Yes
2	0.693	0.693	0.008	0.059	0.013			10:02:38	Yes
3	0.704	0.704	0.009	0.060	0.013			10:03:12	Yes
Mean:	0.671	0.671	0.008						
SD:	0.047	0.047	0.0006						
%RSD:	7.032	7.032	7.03						

QC value within limits for Se 196.03 Recovery = 134.26%
 All analyte(s) passed QC.

Sequence No.: 10
 Sample ID: CCV
 Analyst:

Autosampler Location: 5
 Date Collected: 1/18/2008 10:04:01 AM
 Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.58	10.58	0.128	0.701	0.133			10:04:24	Yes
2	9.438	9.438	0.114	0.624	0.119			10:04:58	Yes
3	9.472	9.472	0.115	0.618	0.120			10:05:32	Yes
Mean:	9.829	9.829	0.119						

SD: 0.648 0.648 0.0078
%RSD: 6.590 6.590 6.59

QC value within limits for Se 196.03 Recovery = 98.29%
All analyte(s) passed QC.

Sequence No.: 11
Sample ID: CCB
Analyst:

Autosampler Location: 1
Date Collected: 1/18/2008 10:06:23 AM
Data Type: Original

Replicate Data: CCB

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Sequence No.: 12
Sample ID: K0711649-001
Analyst:

Autosampler Location: 9
Date Collected: 1/18/2008 10:08:42 AM
Data Type: Original

Replicate Data: K0711649-001

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics. Includes handwritten signature '1/18/08'.

Sequence No.: 13
Sample ID: K0711649-001A
Analyst:

Autosampler Location: 10
Date Collected: 1/18/2008 10:11:00 AM
Data Type: Original

Replicate Data: K0711649-001A

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 14
Sample ID: CCV
Analyst:

Autosampler Location: 5
Date Collected: 1/18/2008 10:13:20 AM
Data Type: Original

Replicate Data: CCV

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Mean: 9.581 9.581 0.116
 SD: 0.258 0.258 0.0031
 %RSD: 2.693 2.693 2.69

QC value within limits for Se 196.03 Recovery = 95.81%
 All analyte(s) passed QC.

Sequence No.: 15
 Sample ID: CCB
 Analyst:

Autosampler Location: 1
 Date Collected: 1/18/2008 10:15:42 AM
 Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.319	0.319	0.004	0.036	0.009			10:16:03	Yes
2	0.290	0.290	0.004	0.041	0.008			10:16:37	Yes
3	0.295	0.295	0.004	0.033	0.008			10:17:11	Yes
Mean:	0.301	0.301	0.004						
SD:	0.015	0.015	0.0002						
%RSD:	5.108	5.108	5.11						

QC value within limits for Se 196.03 Recovery = Not calculated
 All analyte(s) passed QC.

Analysis Begun

Logged In Analyst: ACQMET10 Technique: AA FIAS-Flame
 Spectrometer Model: AAnalyst 200, S/N 200S5061701 Autosampler Model: AS-90

Sample Information File: C:\data-AA\ACQMET10\Sample Information\011808-Se.sif

Batch ID: 011808-Se

Results Data Set: 011808-Se

Results Library: R:\ICP\WIP\DATA\K-FLAA-02\Results.mdb

Sequence No.: 16

Sample ID: K0711649-MB

Analyst:

Autosampler Location:

Date Collected: 1/18/2008 10:23:32 AM

Data Type: Original

Replicate Data: K0711649-MB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.282	0.282	0.003	0.031	0.008			10:23:48	Yes
2	0.340	0.340	0.004	0.031	0.009			10:24:22	Yes
3	0.307	0.307	0.004	0.032	0.009			10:24:56	Yes
Mean:	0.310	0.310	0.004						
SD:	0.029	0.029	0.0004						
%RSD:	9.456	9.456	9.46						

Sequence No.: 17

Sample ID: LCSS DOLT 1/20

Analyst:

Autosampler Location:

Date Collected: 1/18/2008 10:25:53 AM

Data Type: Original

Replicate Data: LCSS DOLT 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.069	3.069	0.037	0.235	0.042			10:26:09	Yes
2	2.994	2.994	0.036	0.197	0.041			10:26:43	Yes
3	2.938	2.938	0.036	0.213	0.040			10:27:17	Yes
Mean:	3.000	3.000	0.036						
SD:	0.065	0.065	0.0008						
%RSD:	2.178	2.178	2.18						

Sequence No.: 18
 Sample ID: K0711649-001 1/4
 Analyst:

Autosampler Location:
 Date Collected: 1/18/2008 10:28:34 AM
 Data Type: Original

Replicate Data: K0711649-001 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.094	2.094	0.025	0.146	0.030			10:28:51	Yes
2	2.006	2.006	0.024	0.132	0.029			10:29:25	Yes
3	2.156	2.156	0.026	0.148	0.031			10:29:59	Yes
Mean:	2.085	2.085	0.025						
SD:	0.076	0.076	0.0009						
%RSD:	3.626	3.626	3.63						

Sequence No.: 19
 Sample ID: K0711649-001D 1/4
 Analyst:

Autosampler Location:
 Date Collected: 1/18/2008 10:31:07 AM
 Data Type: Original

Replicate Data: K0711649-001D 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.213	2.213	0.027	0.147	0.032			10:31:23	Yes
2	2.241	2.241	0.027	0.153	0.032			10:31:57	Yes
3	2.346	2.346	0.028	0.153	0.033			10:32:31	Yes
Mean:	2.267	2.267	0.027						
SD:	0.070	0.070	0.0008						
%RSD:	3.094	3.094	3.09						

Sequence No.: 20
 Sample ID: K0711649-001S 1/20
 Analyst:

Autosampler Location:
 Date Collected: 1/18/2008 10:33:47 AM
 Data Type: Original

Replicate Data: K0711649-001S 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.782	3.782	0.046	0.269	0.051			10:34:03	Yes
2	3.741	3.741	0.045	0.256	0.050			10:34:38	Yes
3	3.636	3.636	0.044	0.257	0.049			10:35:12	Yes
Mean:	3.720	3.720	0.045						
SD:	0.076	0.076	0.0009						
%RSD:	2.030	2.030	2.03						

Sequence No.: 21
 Sample ID: K0711649-002 1/4
 Analyst:

Autosampler Location:
 Date Collected: 1/18/2008 10:36:43 AM
 Data Type: Original

Replicate Data: K0711649-002 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.321	2.321	0.028	0.144	0.033			10:36:59	Yes
2	2.387	2.387	0.029	0.157	0.034			10:37:34	Yes
3	2.358	2.358	0.029	0.157	0.033			10:38:08	Yes
Mean:	2.355	2.355	0.029						
SD:	0.033	0.033	0.0004						
%RSD:	1.401	1.401	1.40						

Sequence No.: 22
 Sample ID: K0711649-003 1/4

Autosampler Location:
 Date Collected: 1/18/2008 10:39:19 AM

Analyst:

Data Type: Original

Replicate Data: K0711649-003 1/4

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.146	2.146	0.026	0.142	0.031			10:39:35	Yes
2	2.163	2.163	0.026	0.143	0.031			10:40:09	Yes
3	1.946	1.946	0.024	0.104	0.028			10:40:44	Yes
Mean:	2.085	2.085	0.025						
SD:	0.121	0.121	0.0015						
%RSD:	5.787	5.787	5.79						

===== Sequence No.: 23

Autosampler Location:

Sample ID: K0711649-004 1/4

Date Collected: 1/18/2008 10:41:52 AM

Analyst:

Data Type: Original

Replicate Data: K0711649-004 1/4

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.520	2.520	0.031	0.176	0.035			10:42:08	Yes
2	2.363	2.363	0.029	0.158	0.033			10:42:42	Yes
3	2.574	2.574	0.031	0.165	0.036			10:43:16	Yes
Mean:	2.485	2.485	0.030						
SD:	0.109	0.109	0.0013						
%RSD:	4.405	4.405	4.41						

===== Sequence No.: 24

Autosampler Location:

Sample ID: K0711649-005 1/4

Date Collected: 1/18/2008 10:44:24 AM

Analyst:

Data Type: Original

Replicate Data: K0711649-005 1/4

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.134	2.134	0.026	0.135	0.031			10:44:40	Yes
2	2.103	2.103	0.025	0.125	0.030			10:45:15	Yes
3	2.192	2.192	0.027	0.138	0.031			10:45:51	Yes
Mean:	2.143	2.143	0.026						
SD:	0.045	0.045	0.0005						
%RSD:	2.102	2.102	2.10						

===== Sequence No.: 25

Autosampler Location:

Sample ID: K0711649-006 1/4

Date Collected: 1/18/2008 10:46:55 AM

Analyst:

Data Type: Original

Replicate Data: K0711649-006 1/4

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.166	2.166	0.026	0.136	0.031			10:47:12	Yes
2	2.146	2.146	0.026	0.155	0.031			10:47:47	Yes
3	1.988	1.988	0.024	0.108	0.029			10:48:22	Yes
Mean:	2.100	2.100	0.025						
SD:	0.098	0.098	0.0012						
%RSD:	4.653	4.653	4.65						

===== Sequence No.: 26

Autosampler Location:

Sample ID: CCV

Date Collected: 1/18/2008 10:49:31 AM

Analyst:

Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.456	9.456	0.115	0.620	0.119			10:49:48	Yes
2	9.030	9.030	0.109	0.601	0.114			10:50:22	Yes
3	8.989	8.989	0.109	0.591	0.114			10:50:57	Yes
Mean:	9.158	9.158	0.111						
SD:	0.259	0.259	0.0031						
%RSD:	2.824	2.824	2.82						

Sequence No.: 27

Sample ID: CCB

Analyst:

Autosampler Location:

Date Collected: 1/18/2008 10:51:58 AM

Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.141	0.141	0.002	0.016	0.007			10:52:14	Yes
2	0.216	0.216	0.003	0.036	0.007			10:52:48	Yes
3	0.029	0.029	0.000	0.008	0.005			10:53:23	Yes
Mean:	0.129	0.129	0.002						
SD:	0.094	0.094	0.0011						
%RSD:	73.15	73.15	73.15						

Sequence No.: 28

Sample ID: K0711649-007 1/4

Analyst:

Autosampler Location:

Date Collected: 1/18/2008 10:54:34 AM

Data Type: Original

Replicate Data: K0711649-007 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	1.719	1.719	0.021	0.097	0.026			10:54:50	Yes
2	1.807	1.807	0.022	0.119	0.027			10:55:25	Yes
3	1.823	1.823	0.022	0.121	0.027			10:55:59	Yes
Mean:	1.783	1.783	0.022						
SD:	0.056	0.056	0.0007						
%RSD:	3.158	3.158	3.16						

Sequence No.: 29

Sample ID: K0711649-008 1/4

Analyst:

Autosampler Location:

Date Collected: 1/18/2008 10:57:03 AM

Data Type: Original

Replicate Data: K0711649-008 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	1.842	1.842	0.022	0.128	0.027			10:57:20	Yes
2	1.787	1.787	0.022	0.128	0.027			10:57:54	Yes
3	1.667	1.667	0.020	0.094	0.025			10:58:29	Yes
Mean:	1.765	1.765	0.021						
SD:	0.090	0.090	0.0011						
%RSD:	5.075	5.075	5.08						

Sequence No.: 30

Sample ID: K0711649-001A 1/4

Analyst:

Autosampler Location:

Date Collected: 1/18/2008 10:59:37 AM

Data Type: Original

Replicate Data: K0711649-001A 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
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Handwritten signature and date: 1/18/08

1	2.499	2.499	0.030	0.183	0.035			10:59:53	Yes
2	2.567	2.567	0.031	0.185	0.036			11:00:27	Yes
3	2.704	2.704	0.033	0.208	0.038			11:01:00	Yes
Mean:	2.590	2.590	0.031						
SD:	0.105	0.105	0.0013						
%RSD:	4.040	4.040	4.04						

1/18/08

Sequence No.: 31
Sample ID: K0711649-001A 1/4
Analyst:

Autosampler Location:
Date Collected: 1/18/2008 12:15:14 PM
Data Type: Original

Replicate Data: K0711649-001A 1/4

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.382	4.382	0.053	0.341	0.058			12:15:32	Yes
2	4.216	4.216	0.051	0.318	0.056			12:16:07	Yes
3	4.339	4.339	0.053	0.333	0.057			12:16:42	Yes
Mean:	4.312	4.312	0.052						
SD:	0.086	0.086	0.0010						
%RSD:	2.000	2.000	2.00						

Sequence No.: 32
Sample ID: K0711649-001DA 1/4
Analyst:

Autosampler Location:
Date Collected: 1/18/2008 12:17:46 PM
Data Type: Original

Replicate Data: K0711649-001DA 1/4

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.669	4.669	0.057	0.365	0.061			12:18:03	Yes
2	4.318	4.318	0.052	0.326	0.057			12:18:39	Yes
3	4.407	4.407	0.053	0.327	0.058			12:19:14	Yes
Mean:	4.464	4.464	0.054						
SD:	0.182	0.182	0.0022						
%RSD:	4.086	4.086	4.09						

Sequence No.: 33
Sample ID: K0711649-001SA 1/20
Analyst:

Autosampler Location:
Date Collected: 1/18/2008 12:20:17 PM
Data Type: Original

Replicate Data: K0711649-001SA 1/20

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	8.027	8.027	0.097	0.579	0.102			12:20:34	Yes
2	7.435	7.435	0.090	0.543	0.095			12:21:09	Yes
3	7.603	7.603	0.092	0.530	0.097			12:21:44	Yes
Mean:	7.688	7.688	0.093						
SD:	0.305	0.305	0.0037						
%RSD:	3.971	3.971	3.97						

Sequence No.: 34
Sample ID: K0711649-002A 1/4
Analyst:

Autosampler Location:
Date Collected: 1/18/2008 12:22:48 PM
Data Type: Original

Replicate Data: K0711649-002A 1/4

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.602	4.602	0.056	0.357	0.061			12:23:04	Yes
2	4.506	4.506	0.055	0.338	0.059			12:23:39	Yes
3	4.423	4.423	0.054	0.332	0.058			12:24:14	Yes

Mean: 4.511 4.511 0.055
 SD: 0.090 0.090 0.0011
 %RSD: 1.989 1.989 1.99

Sequence No.: 35

Sample ID: K0711649-003A 1/4

Analyst:

Autosampler Location:

Date Collected: 1/18/2008 12:25:17 PM

Data Type: Original

Replicate Data: K0711649-003A 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.400	4.400	0.053	0.339	0.058			12:25:33	Yes
2	4.278	4.278	0.052	0.318	0.057			12:26:08	Yes
3	4.299	4.299	0.052	0.313	0.057			12:26:42	Yes
Mean:	4.326	4.326	0.052						
SD:	0.065	0.065	0.0008						
%RSD:	1.509	1.509	1.51						

Sequence No.: 36

Sample ID: K0711649-004A 1/4

Analyst:

Autosampler Location:

Date Collected: 1/18/2008 12:28:02 PM

Data Type: Original

Replicate Data: K0711649-004A 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.596	4.596	0.056	0.350	0.061			12:28:18	Yes
2	4.530	4.530	0.055	0.344	0.060			12:28:53	Yes
3	4.447	4.447	0.054	0.330	0.059			12:29:27	Yes
Mean:	4.525	4.525	0.055						
SD:	0.075	0.075	0.0009						
%RSD:	1.656	1.656	1.66						

Sequence No.: 37

Sample ID: K0711649-05A 1/4

Analyst:

Autosampler Location:

Date Collected: 1/18/2008 12:30:31 PM

Data Type: Original

Replicate Data: K0711649-05A 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.387	4.387	0.053	0.333	0.058			12:30:48	Yes
2	4.289	4.289	0.052	0.314	0.057			12:31:22	Yes
3	4.490	4.490	0.054	0.322	0.059			12:31:57	Yes
Mean:	4.389	4.389	0.053						
SD:	0.100	0.100	0.0012						
%RSD:	2.289	2.289	2.29						

Sequence No.: 38

Sample ID: CCV

Analyst:

Autosampler Location:

Date Collected: 1/18/2008 12:33:06 PM

Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.843	9.843	0.119	0.667	0.124			12:33:22	Yes
2	9.178	9.178	0.111	0.634	0.116			12:33:57	Yes
3	9.090	9.090	0.110	0.633	0.115			12:34:31	Yes
Mean:	9.370	9.370	0.113						
SD:	0.412	0.412	0.0050						
%RSD:	4.395	4.395	4.39						

Sequence No.: 39
Sample ID: CCB
Analyst:

Autosampler Location:
Date Collected: 1/18/2008 12:35:31 PM
Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.251	0.251	0.003	0.041	0.008			12:35:47	Yes
2	0.277	0.277	0.003	0.039	0.008			12:36:22	Yes
3	0.280	0.280	0.003	0.040	0.008			12:36:56	Yes
Mean:	0.269	0.269	0.003						
SD:	0.016	0.016	0.0002						
%RSD:	5.828	5.828	5.83						

Sequence No.: 40
Sample ID: K0711649-006A 1/4
Analyst:

Autosampler Location:
Date Collected: 1/18/2008 12:37:54 PM
Data Type: Original

Replicate Data: K0711649-006A 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.796	4.796	0.058	0.357	0.063			12:38:10	Yes
2	4.517	4.517	0.055	0.335	0.060			12:38:44	Yes
3	4.616	4.616	0.056	0.337	0.061			12:39:19	Yes
Mean:	4.643	4.643	0.056						
SD:	0.142	0.142	0.0017						
%RSD:	3.054	3.054	3.05						

Sequence No.: 41
Sample ID: K0711649-007A 1/4
Analyst:

Autosampler Location:
Date Collected: 1/18/2008 12:40:15 PM
Data Type: Original

Replicate Data: K0711649-007A 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.411	4.411	0.053	0.330	0.058			12:40:32	Yes
2	4.382	4.382	0.053	0.338	0.058			12:41:06	Yes
3	4.332	4.332	0.052	0.322	0.057			12:41:40	Yes
Mean:	4.375	4.375	0.053						
SD:	0.040	0.040	0.0005						
%RSD:	0.923	0.923	0.92						

Sequence No.: 42
Sample ID: K0711649-008A 1/4
Analyst:

Autosampler Location:
Date Collected: 1/18/2008 12:42:39 PM
Data Type: Original

Replicate Data: K0711649-008A 1/4

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.745	4.745	0.057	0.371	0.062			12:42:55	Yes
2	4.438	4.438	0.054	0.323	0.059			12:43:29	Yes
3	4.361	4.361	0.053	0.334	0.058			12:44:02	Yes
Mean:	4.515	4.515	0.055						
SD:	0.203	0.203	0.0025						
%RSD:	4.501	4.501	4.50						

Sequence No.: 43

Autosampler Location:

Sample ID: CCV
Analyst:

Date Collected: 1/18/2008 12:45:17 PM
Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.739	9.739	0.118	0.669	0.123			12:45:32	Yes
2	9.227	9.227	0.112	0.631	0.117			12:46:04	Yes
3	9.401	9.401	0.114	0.636	0.119			12:46:37	Yes
Mean:	9.456	9.456	0.115						
SD:	0.261	0.261	0.0032						
%RSD:	2.755	2.755	2.75						

=====
Sequence No.: 44

Sample ID: CCB
Analyst:

Autosampler Location:

Date Collected: 1/18/2008 12:47:30 PM
Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.317	0.317	0.004	0.049	0.009			12:47:46	Yes
2	0.247	0.247	0.003	0.033	0.008			12:48:18	Yes
3	0.154	0.154	0.002	0.013	0.007			12:48:51	Yes
Mean:	0.239	0.239	0.003						
SD:	0.081	0.081	0.0010						
%RSD:	34.03	34.03	34.03						

February 7, 2008

Analytical Report for Service Request No: K0712111

Kathy Tegtmeyer
New Fields Environmental
4720 Walnut St., Suite 200
Boulder, CO 80301

RE: Se in Tissue

Dear Kathy:

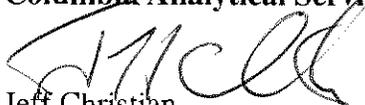
Enclosed are the results of the samples submitted to our laboratory on December 21, 2007. For your reference, these analyses have been assigned our service request number K0712111.

All analyses were performed according to our laboratory's quality assurance program. Where applicable, the methods cited conform to the Methods Update Rule (effective 4/11/2007), which relates to the use of analytical methods for the drinking water and waste water programs. The test results meet requirements of the NELAC standards. Exceptions are noted in the case narrative report where applicable. All results are intended to be considered in their entirety, and Columbia Analytical Services, Inc. (CAS) is not responsible for use of less than the complete report. Results apply only to the items submitted to the laboratory for analysis and individual items (samples) analyzed, as listed in the report.

Please call if you have any questions. My extension is 3316. You may also contact me via Email at JChristian@caslab.com.

Respectfully submitted,

Columbia Analytical Services, Inc.



Jeff Christian
Laboratory Director

JC/lb

Page 1 of 131

Acronyms

ASTM	American Society for Testing and Materials
A2LA	American Association for Laboratory Accreditation
CARB	California Air Resources Board
CAS Number	Chemical Abstract Service registry Number
CFC	Chlorofluorocarbon
CFU	Colony-Forming Unit
DEC	Department of Environmental Conservation
DEQ	Department of Environmental Quality
DHS	Department of Health Services
DOE	Department of Ecology
DOH	Department of Health
EPA	U. S. Environmental Protection Agency
ELAP	Environmental Laboratory Accreditation Program
GC	Gas Chromatography
GC/MS	Gas Chromatography/Mass Spectrometry
LUFT	Leaking Underground Fuel Tank
M	Modified
MCL	Maximum Contaminant Level is the highest permissible concentration of a substance allowed in drinking water as established by the USEPA.
MDL	Method Detection Limit
MPN	Most Probable Number
MRL	Method Reporting Limit
NA	Not Applicable
NC	Not Calculated
NCASI	National Council of the Paper Industry for Air and Stream Improvement
ND	Not Detected
NIOSH	National Institute for Occupational Safety and Health
PQL	Practical Quantitation Limit
RCRA	Resource Conservation and Recovery Act
SIM	Selected Ion Monitoring
TPH	Total Petroleum Hydrocarbons
tr	Trace level is the concentration of an analyte that is less than the PQL but greater than or equal to the MDL.

Inorganic Data Qualifiers

- * The result is an outlier. See case narrative.
- # The control limit criteria is not applicable. See case narrative.
- B The analyte was found in the associated method blank at a level that is significant relative to the sample result.
- E The result is an estimate amount because the value exceeded the instrument calibration range.
- J The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
 - i The MRL/MDL has been elevated due to a matrix interference.
- X See case narrative.

Metals Data Qualifiers

- # The control limit criteria is not applicable. See case narrative.
- B The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- E The percent difference for the serial dilution was greater than 10%, indicating a possible matrix interference in the sample.
- M The duplicate injection precision was not met.
- N The Matrix Spike sample recovery is not within control limits. See case narrative.
- S The reported value was determined by the Method of Standard Additions (MSA).
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
- W The post-digestion spike for furnace AA analysis is out of control limits, while sample absorbance is less than 50% of spike absorbance.
 - i The MRL/MDL has been elevated due to a matrix interference.
- X See case narrative.
- * The duplicate analysis not within control limits. See case narrative.
- + The correlation coefficient for the MSA is less than 0.995.

Organic Data Qualifiers

- * The result is an outlier. See case narrative.
- # The control limit criteria is not applicable. See case narrative.
- A A tentatively identified compound, a suspected aldol-condensation product.
- B The analyte was found in the associated method blank at a level that is significant relative to the sample result.
- C The analyte was qualitatively confirmed using GC/MS techniques, pattern recognition, or by comparing to historical data.
- D The reported result is from a dilution.
- E The result is an estimate amount because the value exceeded the instrument calibration range.
- J The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- N The result is presumptive. The analyte was tentatively identified, but a confirmation analysis was not performed.
- P The GC or HPLC confirmation criteria was exceeded. The relative percent difference is greater than 40% between the two analytical results (25% for CLP Pesticides).
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
 - i The MRL/MDL has been elevated due to a chromatographic interference.
- X See case narrative.

Additional Petroleum Hydrocarbon Specific Qualifiers

- F The chromatographic fingerprint of the sample matches the elution pattern of the calibration standard.
- L The chromatographic fingerprint of the sample resembles a petroleum product, but the elution pattern indicates the presence of a greater amount of lighter molecular weight constituents than the calibration standard.
- H The chromatographic fingerprint of the sample resembles a petroleum product, but the elution pattern indicates the presence of a greater amount of heavier molecular weight constituents than the calibration standard.
- O The chromatographic fingerprint of the sample resembles an oil, but does not match the calibration standard.
- Y The chromatographic fingerprint of the sample resembles a petroleum product eluting in approximately the correct carbon range, but the elution pattern does not match the calibration standard.
- Z The chromatographic fingerprint does not resemble a petroleum product.

Columbia Analytical Services, Inc.
Kelso, WA
State Certifications, Accreditations, and Licenses

Program	Number
Alaska DEC UST	UST-040
Arizona DHS	AZ0339
Arkansas - DEQ	88-0637
California DHS	2286
Colorado DPHE	-
Florida DOH	E87412
Hawaii DOH	-
Idaho DHW	-
Indiana DOH	C-WA-01
Louisiana DEQ	3016
Louisiana DHH	LA050010
Maine DHS	WA0035
Michigan DEQ	9949
Minnesota DOH	053-999-368
Montana DPHHS	CERT0047
Nevada DEP	WA35
New Jersey DEP	WA005
New Mexico ED	-
North Carolina DWQ	605
Oklahoma DEQ	9801
Oregon - DHS	WA200001
South Carolina DHEC	61002
Utah DOH	COLU
Washington DOE	C1203
Wisconsin DNR	998386840
Wyoming (EPA Region 8)	-



Case Narrative

COLUMBIA ANALYTICAL SERVICES, INC.

Client: New Fields Environmental
Project: Tissue - Se
Sample Matrix: Tissue

Service Request No.: K0712111
Date Received: 1/21/07

CASE NARRATIVE

All analyses were performed consistent with the quality assurance program of Columbia Analytical Services, Inc. (CAS). This report contains analytical results for samples designated for Tier III validation deliverables including summary forms and all of the associated raw data for each of the analyses. When appropriate to the method, method blank results have been reported with each analytical test.

Sample Receipt

Tissue samples were received for analysis at Columbia Analytical Services on 1/21/07. The samples were received in good condition and consistent with the accompanying chain of custody form. The samples were stored frozen at -20°C upon receipt at the laboratory.

Total Metals

General Comments:

The samples were freeze-dried to determine moisture and to allow complete homogenization of the dry material. The dried material was milled to a fine meal, and then sub-sampled for digestion. A thorough digestion was performed prior to instrumental analysis to convert all Selenium species to Selenate. Prior to hydride formation, the valence was adjusted by reduction to Selenite.

No anomalies associated with the analysis of these samples were observed.

Approved by _____



Date _____

2/8/08

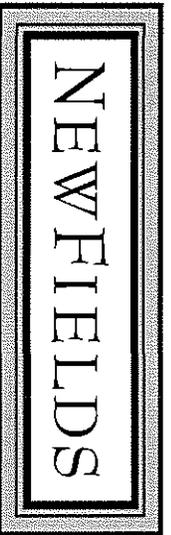
**Chain of Custody
Documentation**

Chain of Custody

Page 1 of 3

Project Contact: Sean Covington/Kathy Tegtmeyer PO 0442-004-900.70
 Courier/Airbill: Columbia Analytical Services, Inc.

Shipped to: 1317 South 13th Ave
 Kelso, WA 98626
 Telephone: (360) 430-7733



4720 Walnut St, Suite 200
 Boulder, CO 80301
 Phone: 303-442-0267
 Fax: 303-442-3679

COC #:

10712111

Sample ID	Sample Date	Sample Time	Matrix	Tot/Dis	Analysis	Preservative	Lab QC	Comments
SM1007-SNPFH-FT0058	10/25/2007	1200	Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNPFH-FT0059	10/25/2007	1200	Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNPFH-FT0060	10/25/2007	1345	Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNPFH-FT0061	10/25/2007	1415	Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNPFH-FT0062	10/25/2007	1410	Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNPFH-FT0063	10/25/2007	1430	Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNPFH-FT0064	10/25/2007	1335	Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1007-SNPFH-FT0065	10/25/2007	1540	Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0066	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0067	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0068	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0069	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0070	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0071	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0072	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0073	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0074	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0075	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		

Sampler Signature: *[Signature]*

Relinquished by: *[Signature]*

Date/Time: 12/29/07 01:50

Received by: *[Signature]*

Date/Time: 12/29/07 1:50

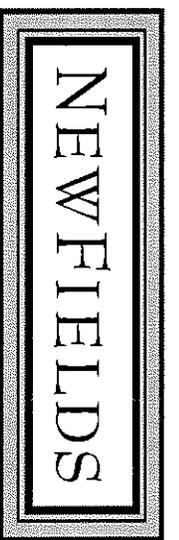
LAB USE ONLY - Sample condition on Receipt

Chain of Custody

Project Contact: Sean Covington/Kathy Tegtmeyer PO 0442-004-900.70
 Courier/Airbill: Columbia Analytical Services, Inc.

Shipped to: 1317 South 13th Ave
 Kelso, WA 98626

Telephone: (360) 430-7733



4720 Walnut St, Suite 200
 Boulder, CO 80301
 Phone: 303-442-0267
 Fax: 303-442-3679

col tape # 40312

COC #:

10712111

Sample ID	Sample Date	Sample Time	Matrix	Tot/Diss	Analysis	Preservative	Lab QC	Comments
SM1107-LSV2c-FT0076	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0077	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0078	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-LSV2c-FT0079	11/14/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC150-FT0080	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC150-FT0081	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC150-FT0082	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC150-FT0083	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC150-FT0084	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC150-FT0085	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC150-FT0086	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC150-FT0087	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC150-FT0088	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC350-FT0089	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC350-FT0090	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1107-CC350-FT0091	11/15/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		
SM1207-SPCFH-FT0092	12/4/2007		Egg Tissue	Tot	Selenium, % Solids	Dry Ice		

Sampler Signature: *[Signature]* LAB USE ONLY - Sample condition on Receipt

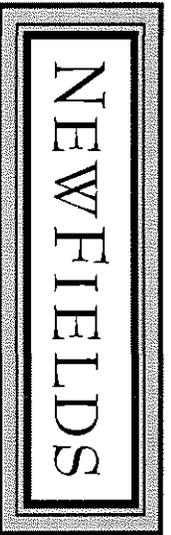
Relinquished by: *[Signature]* Date/Time: *12/29/07 01:50*
 Received by: *[Signature]* Date/Time: *12/29/07 13:00*

Chain of Custody Page 3 of 3

Project Contact: Sean Covington/Kathy Tegtmeyer PO 0442-004-900.70
 Courier/Airbill:

Shipped to: Columbia Analytical Services, Inc.

1317 South 13th Ave
 Kelso, WA 98626
 Telephone: (360) 430-7733



4720 Walnut St., Suite 200
 Boulder, CO 80301
 Phone: 303-442-0267
 Fax: 303-442-3679

COC #: 10712111

Sample ID	Sample Date	Sample Time	Matrix	Tox/ Diss	Analysis	Preservative	Lab QC	Comments
Total Number of Containers: <u>35</u> Individual Lines Reflect Single Containers, Except for Aqueous Analyses Assigned as Laboratory QC								

Sampler Signature: [Signature]

Relinquished by: [Signature] Date/Time: 12/29/07 @ 1500

Received by: [Signature] Date/Time: 12/29/07 1200

LAB USE ONLY - Sample condition on Receipt

**Columbia Analytical Services, Inc.
Cooler Receipt and Preservation Form**

PC JCW

Client / Project: New fields Service Request **K07** 12111

Received: 12-21-2007 Opened: 12-21-2007 By: DJ

1. Samples were received via? US Mail ~~Fed Ex~~ UPS DHL GH GS PDX Courier Hand Delivered
2. Samples were received in: (circle) Cooler Box Envelope Other _____ NA
3. Were custody seals on coolers? NA Y N If yes, how many and where? Front
- If present, were custody seals intact? Y N If present, were they signed and dated? Y N
4. Is shipper's air-bill filed? If not, record air-bill number: 7904 1025 9599 NA Y N

5. Temperature of cooler(s) upon receipt (°C): -8.4
- Temperature Blank (°C): _____
6. If applicable, list Chain of Custody Numbers: _____
7. Were custody papers properly filled out (ink, signed, etc.)? NA Y N
8. Packing material used. Inserts Bubble Wrap Gel Packs Wet Ice Sleeves Other Dry Ice
9. Did all bottles arrive in good condition (unbroken)? Indicate in the table below. NA Y N
10. Were all sample labels complete (i.e analysis, preservation, etc.)? Y N
11. Did all sample labels and tags agree with custody papers? Indicate in the table below. Y N
12. Were the correct types of bottles used for the tests indicated? NA Y N
13. Were all of the preserved bottles received at the lab with the appropriate pH? Indicate in the table below. NA Y N
14. Were VOA vials and 1631 Mercury bottles checked for absence of air bubbles? Indicate in the table below. NA Y N
- Are CWA Microbiology samples received with >1/2 the 24hr. hold time remaining from collection? NA Y N
16. Was C12/Res negative? NA Y N

Sample ID on Bottle	Sample ID on COC	Sample ID on Bottle	Sample ID on COC

Sample ID	Bottle Count	Bottle Type	Out of Temp	Head-space	Broken	pH	Reagent	Volume added	Reagent Lot Number	Initials

Additional Notes, Discrepancies, & Resolutions: _____

Total Solids

COLUMBIA ANALYTICAL SERVICES, INC.

Analytical Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0712111
Date Collected: 10/25-11/14/07
Date Received: 12/21/07

Solids, Total

Prep Method: NONE
 Analysis Method: Freeze Dry
 Test Notes:

Units: PERCENT
 Basis: Wet

Sample Name	Lab Code	Date Analyzed	Result	Result Notes
SM1007-SNFH-FT0058	K0712111-001	1/10/08	31.2	
SM1007-SNFH-FT0059	K0712111-002	1/10/08	26.0	
SM1007-SNFH-FT0060	K0712111-003	1/10/08	31.1	
SM1007-SNFH-FT0061	K0712111-004	1/10/08	23.8	
SM1007-SNFH-FT0062	K0712111-005	1/10/08	31.0	
SM1007-SNFH-FT0063	K0712111-006	1/10/08	29.5	
SM1007-SNFH-FT0064	K0712111-007	1/10/08	27.9	
SM1007-SNFH-FT0065	K0712111-008	1/10/08	29.8	
SM1107-LSV2c-FT0066	K0712111-009	1/10/08	32.4	
SM1107-LSV2c-FT0067	K0712111-010	1/10/08	33.9	
SM1107-LSV2c-FT0068	K0712111-011	1/10/08	32.1	
SM1107-LSV2c-FT0069	K0712111-012	1/10/08	32.9	
SM1107-LSV2c-FT0070	K0712111-013	1/10/08	25.8	
SM1107-LSV2c-FT0071	K0712111-014	1/10/08	32.2	
SM1107-LSV2c-FT0072	K0712111-015	1/10/08	32.1	
SM1107-LSV2c-FT0073	K0712111-016	1/10/08	33.4	
SM1107-LSV2c-FT0074	K0712111-017	1/10/08	30.6	
SM1107-LSV2c-FT0075	K0712111-018	1/10/08	31.4	
SM1107-LSV2c-FT0076	K0712111-019	1/10/08	31.7	
SM1107-LSV2c-FT0077	K0712111-020	1/10/08	31.7	

COLUMBIA ANALYTICAL SERVICES, INC.

Analytical Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0712111
Date Collected: 11/14-12/4/07
Date Received: 12/21/07

Solids, Total

Prep Method: NONE
Analysis Method: Freeze Dry
Test Notes:

Units: PERCENT
Basis: Wet

Sample Name	Lab Code	Date Analyzed	Result	Result Notes
SM1107-LSV2c-FT0078	K0712111-021	1/10/08	32.6	
SM1107-LSV2c-FT0079	K0712111-022	1/10/08	33.9	
SM1107-CC150-FT0080	K0712111-023	1/10/08	31.9	
SM1107-CC150-FT0081	K0712111-024	1/10/08	30.6	
SM1107-CC150-FT0082	K0712111-025	1/10/08	30.5	
SM1107-CC150-FT0083	K0712111-026	1/10/08	30.4	
SM1107-CC150-FT0084	K0712111-027	1/10/08	31.0	
SM1107-CC150-FT0085	K0712111-028	1/10/08	32.2	
SM1107-CC150-FT0086	K0712111-029	1/10/08	32.2	
SM1107-CC150-FT0087	K0712111-030	1/10/08	33.1	
SM1107-CC150-FT0088	K0712111-031	1/10/08	31.9	
SM1107-CC350-FT0089	K0712111-032	1/10/08	29.9	
SM1107-CC350-FT0090	K0712111-033	1/10/08	31.9	
SM1107-CC350-FT0091	K0712111-034	1/10/08	33.5	
SM1207-SPCFH-FT0092	K0712111-035	1/10/08	33.2	

COLUMBIA ANALYTICAL SERVICES, INC.

QA/QC Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0712111
Date Collected: 10/25/07
Date Received: 12/21/07
Date Extracted: NA
Date Analyzed: 1/10/08

Duplicate Summary
Total Metals

Sample Name: SM1007-SNFH-FT0058
Lab Code: K0712111-001D
Test Notes:

Units: PERCENT
Basis: Wet

Analyte	Prep Method	Analysis Method	Sample Result	Duplicate Sample Result	Average	Relative Percent Difference	Result Notes
Solids, Total	NA	Freeze Dry	31.2	28.3	29.8	10	

COLUMBIA ANALYTICAL SERVICES, INC.

QA/QC Report

Client: New Fields Environmental
Project: Se in Tissue
Sample Matrix: Tissue

Service Request: K0712111
Date Collected: 11/14/07
Date Received: 12/21/07
Date Extracted: NA
Date Analyzed: 1/10/08

Duplicate Summary
Total Metals

Sample Name: SM1107-LSV2c-FT0070
Lab Code: K0712111-013D
Test Notes:

Units: PERCENT
Basis: Wet

Analyte	Prep Method	Analysis Method	Sample Result	Duplicate Sample Result	Average	Relative Percent Difference	Result Notes
Solids, Total	NA	Freeze Dry	25.8	29.1	27.4	12	

Service Request #:

K0712111

Analytical Batch

Analysis For:

Freeze Dried Solids

KA0628234

Lab Code	Wet Weight (g)	Tare (g)	Tare + Dry Wt.(g)	Dry Weight (g)	% Total Solids
K0712111-001	24.59	81.87	89.53	7.66	31.2
ELV-10-06-02	24.62	81.38	88.89	6.96	
-001D	24.62	81.93	88.34	6.96	28.3
-002	24.68	81.93	88.34	6.41	26.0
-003	24.51	81.83	89.45	7.62	31.1
-004	24.37	81.86	87.67	5.81	23.8
-005	24.00	81.96	89.40	7.44	31.0
-006	23.56	81.91	88.86	6.95	29.5
-007	23.69	81.97	88.59	6.62	27.9
-008	24.95	82.06	89.49	7.43	29.8
-009	23.13	81.73	89.23	7.50	32.4
-010	4.33	81.18	82.65	1.47	33.9
-011	15.12	81.84	86.70	4.80	32.1
-012	10.18	81.31	84.66	3.35	32.9
-013	24.93	81.87	88.30	6.43	25.8
-013D	24.87	81.30	88.54	7.24	29.1
-014	14.89	82.00	86.80	4.80	32.2
-015	4.58	81.04	82.51	1.47	32.1
-016	3.32	81.46	82.57	1.11	33.4
-017	24.15	81.75	89.14	7.39	30.6
-018	14.73	81.93	86.56	4.63	31.4
-019	9.73	81.04	84.12	3.08	31.7
-020	17.79	81.38	87.02	5.64	31.7
-021	8.09	81.78	84.43	2.64	32.6
-022	23.47	81.39	89.35	7.96	33.9
-023	24.44	81.18	88.97	7.79	31.9
-024	10.48	82.02	85.23	3.21	30.6

AB11/14/08

Time In: 4:00 p.m. 1/11/08

Time Out: 9:00 a.m. 1/14/08

Comments:

Balance 23

\bar{x} = RPD =

Analyst: <u>[Signature]</u>	Date: <u>1/10/08</u>
Reviewed By: <u>[Signature]</u>	Date: <u>1/30/08</u>

Service Request #:

K0712111

Analysis For:

Freeze Dried Solids

Lab Code	Wet Weight (g)	Tare (g)	Tare + Dry Wt.(g)	Dry Weight (g)	% Total Solids
K0712111-025	11.12	81.54	84.93	3.39	30.5
-026	24.64	81.47	88.95	7.48	30.4
-027	22.47	82.05	89.01	6.96	31.0
-028	24.77	81.97	89.95	7.98	32.2
-029	8.30	82.24	EW 87.52 81.91	2.107	32.2
-030	18.36	81.45	87.55	6.07	33.1
-031	23.00	82.01	89.35	7.34	31.9
-032	24.74	82.08	89.47	7.39	29.9
-033	23.64	81.54	89.07	7.53	31.9
-034	21.06	82.06	89.11	7.05	33.5
-035	24.60	81.31	89.47	8.16	33.2
EW 1/10/08					

Time In: _____

Time Out: _____

Comments: _____

\bar{x} = _____ RPD = _____

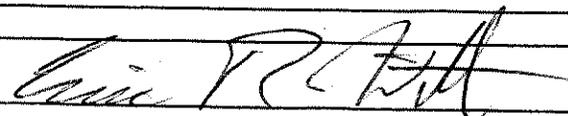
Analyst: <u>[Signature]</u>	Date: <u>1/10/08</u>
Reviewed By: <u>[Signature]</u>	Date: <u>1/30/08</u>

Sample Number(s): As Listed	Service Request Number(s): K0712111
--------------------------------	--

TISSUE COMPOSITION DATA

Laboratory ID	Weight (g)	Tare (g)			Matrix	Length
K0712111-01	249.37 256.10	275.57			Fish eggs	
-02	237.34	275.95				
-03	638.05 ^{362.50}	275.55				
-04	41.79	275.68				
-05	359.85	275.49				
-06	186.99	275.83				
-07	196.20	275.91				
-08	238.70	275.74				
-09	25.28	275.09				
-10	5.15	275.02				
-11	15.62	275.91				
-12	10.69	274.79				
-13	125.82	276.96				
-14	15.40	277.09				
-15	4.88	276.85				
-16	3.58	276.31				
-17	38.11	277.22				
-18	15.09	277.03				
-19	10.09	276.73				
-20	18.17	276.45				
-21	8.58	276.35				
-22	28.87	276.76				
-23	33.31	277.00				
-24	11.15	276.74				
-25	11.59	278.02				

Comments:

Analyst: 	Date: 1/10/08
Reviewed: 	Date: 1/30/08

Sample Number(s): As Listed	Service Request Number(s): K0712111
--------------------------------	--

TISSUE COMPOSITION DATA

Laboratory ID	Weight (g)	Tare (g)			Matrix	Length
K0712111-26	42.58	277.17			fish eggs	
-27	23.04	278.07				
-28	29.49	278.23				
-29	8.72	277.73				
-30	18.85	278.29				
-31	37.67	278.18				
-32	38.69	278.33				
-33	30.08	277.28				
-34	21.74	278.25				
-35	68.60	277.40				
<div style="font-size: 2em; font-family: cursive;">I W</div> <div style="font-size: 1.5em; font-family: cursive;">1/10/08</div>						

Comments:

Analyst: <i>[Signature]</i>	Date: I-10-08
Reviewed: <i>[Signature]</i>	Date: 1/30/08

Metals

Columbia Analytical Services

- Cover Page -
INORGANIC ANALYSIS DATA PACKAGE

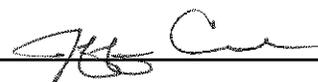
Client: New Fields Environmental
Project Name: Se in Tissue
Project No.:

Service Request: K0712111

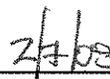
<u>Sample Name:</u>	<u>Lab Code:</u>
SM1007-SNFH-FT0058	K0712111-001
SM1007-SNFH-FT0058D	K0712111-001D
SM1007-SNFH-FT0058S	K0712111-001S
SM1007-SNFH-FT0059	K0712111-002
SM1007-SNFH-FT0060	K0712111-003
SM1007-SNFH-FT0061	K0712111-004
SM1007-SNFH-FT0062	K0712111-005
SM1007-SNFH-FT0063	K0712111-006
SM1007-SNFH-FT0064	K0712111-007
SM1007-SNFH-FT0065	K0712111-008
SM1107-LSV2c-FT0066	K0712111-009
SM1107-LSV2c-FT0067	K0712111-010
SM1107-LSV2c-FT0068	K0712111-011
SM1107-LSV2c-FT0069	K0712111-012
SM1107-LSV2c-FT0070	K0712111-013
SM1107-LSV2c-FT0071	K0712111-014
SM1107-LSV2c-FT0072	K0712111-015
SM1107-LSV2c-FT0073	K0712111-016
SM1107-LSV2c-FT0074	K0712111-017
SM1107-LSV2c-FT0075	K0712111-018
SM1107-LSV2c-FT0076	K0712111-019
SM1107-LSV2c-FT0077	K0712111-020
SM1107-LSV2c-FT0078	K0712111-021
SM1107-LSV2c-FT0079	K0712111-022
SM1107-LSV2c-FT0079D	K0712111-022D
SM1107-LSV2c-FT0079S	K0712111-022S
SM1107-CC150-FT0080	K0712111-023
SM1107-CC150-FT0081	K0712111-024
SM1107-CC150-FT0082	K0712111-025
SM1107-CC150-FT0083	K0712111-026
SM1107-CC150-FT0084	K0712111-027
SM1107-CC150-FT0085	K0712111-028
SM1107-CC150-FT0086	K0712111-029
SM1107-CC150-FT0087	K0712111-030
SM1107-CC150-FT0088	K0712111-031
SM1107-CC350-FT0089	K0712111-032
SM1107-CC350-FT0090	K0712111-033
SM1107-CC350-FT0091	K0712111-034
SM1207-SPCFH-FT0092	K0712111-035

Comments:

Approved By:



Date:



Columbia Analytical Services

**- Cover Page -
INORGANIC ANALYSIS DATA PACKAGE**

Client: New Fields Environmental
Project Name: Se in Tissue
Project No.:

Service Request: K0712111

Sample Name:

Method Blank

Method Blank

Lab Code:

K0712111-MB

K0712111-MB2

Comments:

Approved By: _____

Date: _____

Metals

- 1 -

INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental Service Request: K0712111
Project No.: NA Date Collected: 11/14/07
Project Name: Se in Tissue Date Received: 12/21/07
Matrix: TISSUE Units: mg/Kg
Basis: DRY

Sample Name: SM1107-LSV2c-FT0077 Lab Code: K0712111-020

Analyte	Analysis Method	MRL	MDL	Dilution Factor	Date Extracted	Date Analyzed	Result	C	Q
Selenium	7742	0.96	0.19	20.0	01/28/08	02/05/08	12.5		

Comments:

Metals

- 1 -

INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental Service Request: K0712111
Project No.: NA Date Collected: 11/15/07
Project Name: Se in Tissue Date Received: 12/21/07
Matrix: TISSUE Units: mg/Kg
Basis: DRY

Sample Name: SM1107-CC150-FT0080 Lab Code: K0712111-023

Analyte	Analysis Method	MRL	MDL	Dilution Factor	Date Extracted	Date Analyzed	Result	C	Q
Selenium	7742	0.47	0.09	10.0	01/28/08	02/05/08	12.8		

Comments:

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium	10.0	9.74	97	10.0	9.85	98	10.04	100	7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium				10.0	10.11	101	10.04	100	7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium				10.0	9.68	97	10.10	101	7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium				10.0	10.02	100			7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium	10.0	10.03	100	10.0	10.23	102	10.42	104	7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium				10.0	10.28	103			7742

Metals

- 2b -

CRDL STANDARD FOR AA AND ICP

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Concentration Units: ug/L

Analyte	CRDL Standard for AA			CRDL Standard for ICP				
	True	Found	%R	Initial		Final		
	True	Found	%R	True	Found	%R	Found	%R
Selenium	0.5	0.54	108.0					

Metals

- 2b -

CRDL STANDARD FOR AA AND ICP

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Concentration Units: ug/L

Analyte	CRDL Standard for AA			CRDL Standard for ICP				
	True	Found	%R	Initial		Final		
	True	Found	%R	True	Found	%R	Found	%R
Selenium	0.5	0.54	108.0					

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)		Continuing Calibration Blank (ug/L)						Method
	1	C	1	C	2	C	3	C	
Selenium	0.1	U	0.1	U	0.1	U	0.1	U	7742

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)	C	Continuing Calibration Blank (ug/L)						Method
			1	C	2	C	3	C	
Selenium			0.2	B	0.1	U	0.1	U	7742

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)	Continuing Calibration Blank (ug/L)						Method	
		C	1	C	2	C	3		C
Selenium			0.1	U	0.1	U	0.1	U	7742

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)		Continuing Calibration Blank (ug/L)						Method
	1	C	1	C	2	C	3	C	
Selenium	0.1	U	0.1	U	0.1	U	0.1	U	7742

Metals

- 5A -

SPIKE SAMPLE RECOVERY

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Units: MG/KG

Project Name: Se in Tissue

Basis: DRY

Matrix: TISSUE

Sample Name: SM1007-SNFH-FT0058S

Lab Code: K0712111-001S

Analyte	Control Limit %R	Spike Result	C	Sample Result	C	Spike Added	%R	Q	Method
Selenium	65 - 124	13.4		0.76		16.32	77.5		7742

An empty field in the Control Limit column indicates the control limit is not applicable

Metals

- 5A -

SPIKE SAMPLE RECOVERY

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Units: MG/KG

Project Name: Se in Tissue

Basis: DRY

Matrix: TISSUE

Sample Name: SM1107-LSV2c-FT0079S

Lab Code: K0712111-022S

Analyte	Control Limit %R	Spike Result C	Sample Result C	Spike Added	%R	Q	Method
Selenium	65 - 124	44.3	28.1	16.32	99.3		7742

An empty field in the Control Limit column indicates the control limit is not applicable

Metals

- 5B -

POST SPIKE SAMPLE RECOVERY

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Units: UG/L

Project Name: Se in Tissue

Basis: DRY

Matrix: WATER

Sample Name: SM1107-LSV2c-FT0068A

Lab Code: K0712111-011A

Analyte	Control Limit %R	Spike Result C	Sample Result C	Spike Added	%R	Q	Method
Selenium	75-125	13.66	10.45	3.0	107		7742

Metals

- 5B -

POST SPIKE SAMPLE RECOVERY

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Units: UG/L

Project Name: Se in Tissue

Basis: DRY

Matrix: WATER

Sample Name: SM1107-LSV2c-FT0079A

Lab Code: K0712111-022A

Analyte	Control Limit %R	Spike Result	C	Sample Result	C	Spike Added	%R	Q	Method
Selenium	75-125	11.36		8.32		3.0	101.3		7742

Metals
- 6 -
DUPLICATES

Client: New Fields Environmental Service Request: K0712111
Project No.: NA Units: MG/KG
Project Name: Se in Tissue Basis: DRY
Matrix: TISSUE

Sample Name: SM1007-SNFH-FT0058D Lab Code: K0712111-001D

Analyte	Control Limit	Sample (S)	C	Duplicate (D)	C	RPD	Q	Method
Selenium		0.76		0.80		5.1		7742

An empty field in the Control Limit column indicates the control limit is not applicable.

Metals

- 6 -

DUPLICATES

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Units: MG/KG

Project Name: Se in Tissue

Basis: DRY

Matrix: TISSUE

Sample Name: SM1107-LSV2c-FT0079D

Lab Code: K0712111-022D

Analyte	Control Limit	Sample (S)	C	Duplicate (D)	C	RPD	Q	Method
Selenium	30	28.1		30.2		7.2		7742

An empty field in the Control Limit column indicates the control limit is not applicable.

Metals

- 7 -

LABORATORY CONTROL SAMPLE

Client: New Fields Environmental Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Aqueous LCS Source: Solid LCS Source: NRCC DOLT 3

Analyte	Aqueous (ug/L)			Solid (mg/kg)					
	True	Found	%R	True	Found	C	Limits	%R	
Selenium				7.06	7.9		5.26	9.05	111.9

Metals

- 7 -

LABORATORY CONTROL SAMPLE

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Aqueous LCS Source:

Solid LCS Source: NRCC DOLT 3

Analyte	Aqueous (ug/L)			Solid (mg/kg)				
	True	Found	%R	True	Found	C	Limits	%R
Selenium				7.06	8.1		5.26 9.05	114.7

Metals

- 10 -

DETECTION LIMITS

Client: New Fields Environmental Service Request: K0712111
Project No.: NA
Project Name: Se in Tissue

ICP/ICP-MS ID #: _____
GFAA ID #: K-FLAA-02 AA ID #: _____

Analyte	Wave-length (nm)	Back-ground	MRL ug/L	MDL ug/L	M
Selenium			0.5	0.1	F

Comments: _____

Metals
-13-
PREPARATION LOG

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Method: F

Sample ID	Preparation Date	Initial Volume	Final Volume (mL)
K0712111-001	1/28/08	0.3470	30.0
K0712111-001D	1/28/08	0.3050	30.0
K0712111-001S	1/28/08	0.3070	30.0
K0712111-002	1/28/08	0.3230	30.0
K0712111-003	1/28/08	0.3180	30.0
K0712111-004	1/28/08	0.3150	30.0
K0712111-005	1/28/08	0.3020	30.0
K0712111-006	1/28/08	0.3200	30.0
K0712111-007	1/28/08	0.3090	30.0
K0712111-008	1/28/08	0.3100	30.0
K0712111-009	1/28/08	0.3280	30.0
K0712111-010	1/28/08	0.3400	30.0
K0712111-011	1/28/08	0.3480	30.0
K0712111-012	1/28/08	0.3050	30.0
K0712111-013	1/28/08	0.3330	30.0
K0712111-014	1/28/08	0.3410	30.0
K0712111-015	1/28/08	0.3150	30.0
K0712111-016	1/28/08	0.3100	30.0
K0712111-017	1/28/08	0.3070	30.0
K0712111-018	1/28/08	0.3140	30.0
K0712111-019	1/28/08	0.3390	30.0
K0712111-020	1/28/08	0.3120	30.0
K0712111-MB	1/28/08	0.3000	30.0
LCSS DOLT	1/28/08	0.3420	30.0

Metals
-13-
PREPARATION LOG

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Method: F

Sample ID	Preparation Date	Initial Volume	Final Volume (mL)
K0712111-021	1/28/08	0.3500	30.0
K0712111-022	1/28/08	0.3550	30.0
K0712111-022D	1/28/08	0.3390	30.0
K0712111-022S	1/28/08	0.3070	30.0
K0712111-023	1/28/08	0.3200	30.0
K0712111-024	1/28/08	0.3220	30.0
K0712111-025	1/28/08	0.3100	30.0
K0712111-026	1/28/08	0.3400	30.0
K0712111-027	1/28/08	0.3210	30.0
K0712111-028	1/28/08	0.3090	30.0
K0712111-029	1/28/08	0.3620	30.0
K0712111-030	1/28/08	0.3120	30.0
K0712111-031	1/28/08	0.3620	30.0
K0712111-032	1/28/08	0.3120	30.0
K0712111-033	1/28/08	0.3430	30.0
K0712111-034	1/28/08	0.3190	30.0
K0712111-035	1/28/08	0.3020	30.0
K0712111-MB2	1/28/08	0.3000	30.0
LCSS DOLT2	1/28/08	0.3390	30.0

Metals
- 14 -
ANALYSIS RUN LOG

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Instrument ID Number: K-FLAA-02

Method: F

Start Date: 2/5/08

End Date: 2/5/08

Sample No.	D/F	Time	% R	Analytes																									
				A L	S B	A S	B A	B E	C D	C A	C R	C O	C U	F E	P B	M G	M N	H G	N I	K	S E	A G	N A	T L	V L	Z N	C N		
ZZZZZZ	20	10:52																											
CCV3	1	10:55																			X								
CCB3	1	10:57																			X								
K0712111-017	20	10:59																			X								
K0712111-018	20	11:02																			X								
K0712111-019	20	11:04																			X								
K0712111-020	20	11:07																			X								
K0712111-MB2	2	11:09																			X								
ZZZZZZ	20	11:11																											
K0712111-021	20	11:14																			X								
K0712111-022	40	11:16																			X								
K0712111-022D	40	11:18																			X								
K0712111-022S	40	11:20																			X								
CCV4	1	11:23																			X								
CCB4	1	11:25																			X								
K0712111-023	10	11:27																			X								
K0712111-024	20	11:30																			X								
K0712111-025	20	11:32																			X								
K0712111-026	20	11:35																			X								
K0712111-027	20	11:37																			X								
K0712111-028	20	11:39																			X								
K0712111-029	20	11:42																			X								
K0712111-030	20	11:44																			X								
K0712111-031	20	11:47																			X								
K0712111-032	20	11:49																			X								
CCV5	1	11:51																			X								
CCB5	1	11:54																			X								
ZZZZZZ	20	11:56																											
K0712111-034	20	11:58																			X								
ZZZZZZ	20	12:00																											
ZZZZZZ	4	12:03																											
K0712111-022A	40	12:05	101.3																		X								

* - Denotes additional elements (other than the standard CLP elements) are represented on another Form 14

Metals
- 14 -
ANALYSIS RUN LOG

Client: New Fields Environmental

Service Request: K0712111

Project No.: NA

Project Name: Se in Tissue

Instrument ID Number: K-FLAA-02

Method: F

Start Date: 2/5/08

End Date: 2/5/08

Sample No.	D/F	Time	% R	Analytes																									
				A L	S B	A S	B A	B E	C D	C A	C R	C O	C U	F E	P B	M G	M N	H G	N I	K	S E	A G	N A	T L	V	Z N	C N		
CCV6	1	12:07																			X								
CCB6	1	12:10																			X								
ZZZZZZ	40	12:14																											
ZZZZZZ	40	12:17																											
K0712111-001	4	12:20																			X								
K0712111-001D	4	12:22																			X								
K0712111-004	4	12:24																			X								
ZZZZZZ	20	12:27																											
ZZZZZZ	20	12:30																											
K0712111-016	40	12:32																			X								
ZZZZZZ	20	12:35																											
K0712111-035	2	12:37																			X								
CCV7	1	12:40																			X								
CCB7	1	12:42																			X								
ZZZZZZ	40	13:03																											
K0712111-033	20	13:06																			X								
ZZZZZZ	20	13:08																											
K0712111-011	40	13:11																			X								
K0712111-010	40	13:13																			X								
ZZZZZZ	20	13:16																											
ZZZZZZ	20	13:19																											
ZZZZZZ	40	13:24																											
CCV8	1	13:27																			X								
CCB8	1	13:29																			X								
ZZZZZZ	10	13:32																											
ZZZZZZ	40	13:35																											
ZZZZZZ	40	13:44																											
K0712111-011A	40	14:14	42.7																		X								
CCV9	1	14:16																			X								
CCB9	1	14:19																			X								

* - Denotes additional elements (other than the standard CLP elements) are represented on another Form 14

Columbia Analytical Services
Metals Tissue Digestion Sheet

Service Request Number(s): K0712111 1-20

Star Lims Run No.: L1844 Analysis for: ICP ICP-MS GFAA
 Method: Tissue other: Se hydride

Sample	Initial Weight (g)	freeze Dry	Wet	Final Volume (ml)	Matrix
IPB			X	30mls	15% HNO ₃
Dolt	0.342		↓		
Dorm					
K0712111-01	0.347	X			
- 01x	0.305				
- 02	0.323				
- 03	0.318				
- 04	0.315				
- 05	0.302				
- 06	0.320				
- 07	0.309				
- 08	0.310				
- 09	0.328				
- 10	0.340				
- 11	0.348				
- 12	0.305				
- 13	0.333				
- 14	0.341				
- 15	0.315				
- 16	0.310				
- 17	0.307				
- 18	0.314				
- 19	0.339				
- 20	0.312				
- 015	0.307				

AB 1/25/08

LAB 1/28/08

Time Digestion Started: 4:00pm 1/28/08 Oven Temp: 105°C

Lot # Acids Used: HNO₃ M511-104 C

LCS: Dorm-2, Dolt-3

Time Digestion Ended: 1:00am 1/30/08

Oven Temp: 107°C

Balance I.D.: Lochner

- QCP CICV-1, MET1-63-A, _____ mls. added
- QCP CICV-2, MET1-63-B, _____ mls. added
- QCP CICV-3, MET1-59-J, _____ mls. Added
- SS6, MET1-62-A, _____ mls. Added

SPIKE INFO

- SS1-MET1-61-R, 0.3 mls added
- SS5-MET1-61-P, 0.05 mls added
- SS6-MET1-62-A, 0.05 mls added

Additional spikes: _____

Comments: _____

Analyst <u>Angela Brack</u>	Date <u>1/28/08</u>
Reviewer <u>A</u>	Date <u>1/30/08</u>

TissueDig.xls
1/7/2008

METALS SPIKE FORM

Service Request # K071211-20
 Q.C. Sample # K071211

Circle type of digest: GFAA ICP FAA ICP-MS Other: Initials / Date: UTB, 1/28/08
 Circle type of sample: Soil Water Misc. Sludge Oil Other: TISSUE

Solution Name	Element	mls of 1000ppm Solution	Final Volume	Source	Lot#	Exp. Date	Solution Conc. mg/L	Enter mls Added
SS1-MET1-61-R	HNO3	50.0	1000ml	JT Baker	E17044	-	-	0.3
	Al	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	200	
	Ag	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Ba	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	200	
	Bc	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Cd	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Co	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Cr	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	20	
	Cu	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	25	
	Fe	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	100	
	Pb	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Mn	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Ni	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Sb	50*	1000ml	MET1-53-A	619312	3/7/2008	50	
V	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50		
Zn	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50		
SS4-MET1-63-D	HNO3	25.0	500ml	JT BAKER	E27027	-	-	Expires: 7/1/08
	As	2.0	500ml	MET1-57-Z	Z-AS02032	7/1/2008	4	
	Cd	2.0	500ml	MET1-57-M	Z-CD02004	7/1/2008	4	
	Pb	2.0	500ml	MET1-57-R	A2-PB02135	7/1/2008	4	
	Se	2.0	500ml	MET1-57-K	Z-SE01120	7/1/2008	4	
	Tl	2.0	500ml	MET1-59-B	704313	1/2/2009	4	
SS5-MET1-63-E	HNO3	25.0	500ml	JT BAKER	E27027	-	-	0.05 Expires: 7/1/08
	As	50.0	500ml	MET1-57-Z	Z-AS02032	7/1/2008	100	
	Se	50.0	500ml	MET1-59-C	Z-SE01120	8/1/2008	100	
	Tl	50.0	500ml	MET1-59-B	704313	1/2/2009	100	
SS6-MET1-63-F	HNO3	25	500ml	JT BAKER	E27027	-	-	0.05 Expires: 9/26/08
	B	50	500ml	MET1-59-A	715006	1/2/2009	100	
	Mo	50	500ml	MET1-55-C	705726	9/26/2008	100	

GFLCSW (MET1-60-A)	HNO3	10.0	1000ml	JT BAKER	E01042	-	-	Expires: 3/1/08
	As, Pb, Se, Tl	5.0	1000ml	QCP-CICV-3	Z-CICP19048	3/1/2008	2.5	
	Cd	-	-	QCP-CICV-3	Z-CICP19048	3/1/2008	1.25	
	Cu	2.5	1000ml	MET-55-O	Z-CU0284	5/1/2008	2.5	
QCP-CICV-1 (MET1-63-A)	Ca, Mg, Na, K	no dilution	-	IV	A2-MEB236021	1/1/2009	2500	Expires: 1/1/09
	Al, Ba	no dilution	-	IV	A2-MEB236021	1/1/2009	1000	
	Fe	no dilution	-	IV	A2-MEB236021	1/1/2009	500	
	Co, Mn, Ni, V, Zn	no dilution	-	IV	A2-MEB236021	1/1/2009	250	
	Cu, Ag	no dilution	-	IV	A2-MEB236021	1/1/2009	125	
	Cr	no dilution	-	IV	A2-MEB236021	1/1/2009	100	
	Be	no dilution	-	IV	A2-MEB236021	1/1/2009	25	
QCP-CICV-2 (MET1-63-B)	Sb	no dilution	-	IV	Z-CICP19033	1/1/2009	500	Expires: 1/1/09
QCP-CICV-3 (MET1-63-C)	As, Pb, Se, Tl	no dilution	-	IV	Z-CICP19048	1/1/2009	500	Expires: 1/1/09
	Cd	no dilution	-	IV	Z-CICP19048	1/1/2009	250	

* Denotes volume of 1000 ppm stock standard.

Element	mls of	ppm	Source	Lot# / Lab Code	Exp. Date

Columbia Analytical Services
Metals Tissue Digestion Sheet

Service Request Number(s): K0712111 21-35

Star Lims Run No.: W1845 Analysis for: ICP · ICP-MS GFAA
 Method: Tissue other: Se hydride

Sample	Initial Weight (g)	freeze Dry	Wet	Final Volume (ml)	Matrix
VB			X	30m08.	15/HND2
Dolt	0.339		↓		
K0712111-21	0.350	X			
- 22	0.355				
- 22D	0.339				
- 22S	0.307				
- 23	0.320				
- 24	0.322				
- 25	0.310				
- 26	0.340				
- 27	0.321				
- 28	0.309				
- 29	0.302				
- 30	0.312				
- 31	0.312				
- 32	0.312				
- 33	0.343				
- 34	0.319				
- 35	0.302				
<u>AB 1/28/08</u>					

Time Digestion Started: 4:00 p.m. 1/28/08 Oven Temp: 105°C
 Lot # Acids Used: HNO3 MS1164 C

Time Digestion Ended: _____
 Oven Temp: _____

LCS: Dorm-2, Dolt-3

Balance I.D.: loaner

- QCP CICV-1, MET1-63-A, _____ mls. added
- QCP CICV-2, MET1-63-B, _____ mls. added
- QCP CICV-3, MET1-59-J, _____ mls. Added
- SS6, MET1-62-A, _____ mls. Added

SPIKE INFO

- SS1-MET1-61-R, 0.3 mls added
- SS5-MET1-61-P, 0.05 mls added
- SS6-MET1-62-A, 0.05 mls added

Additional spikes: _____

Comments: _____

Analyst <u>Angela Brack</u>	Date <u>1/28/08</u>
Reviewer _____	Date <u>1/30/08</u>

TissueDig.xls
1/7/2008

METALS SPIKE FORM

Service Request # K071211 (21-35)
 Q.C. Sample # K071211-22

Circle type of digest: GFAA ICP FAA ICP-MS Other: Initials / Date: ARB, 1/28/08
 Circle type of sample: Soil Water Misc. Sludge Oil Other: TISSUE

Solution Name	Element	mls of 1000ppm Solution	Final Volume	Source	Lot#	Exp. Date	Solution Conc. mg/L	Enter mls Added
SS1-MET1-61-R	HNO3	50.0	1000ml	JT Baker	E17044	-	-	0.3
	Al	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	200	
	Ag	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Ba	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	200	
	Be	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Cd	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	5	
	Co	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Cr	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	20	
	Cu	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	25	
	Fe	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	100	
	Pb	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Mn	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Ni	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50	
	Sb	50*	1000ml	MET1-53-A	619312	3/7/2008	50	
V	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50		
Zn	100	1000ml	MET1-60-J	A2-MEB246032	10/1/2008	50		
SS4-MET1-63-D	HNO3	25.0	500ml	JT BAKER	E27027	-	-	Expires: 7/1/08
	As	2.0	500ml	MET1-57-Z	Z-AS02032	7/1/2008	4	
	Cd	2.0	500ml	MET1-57-M	Z-CD02004	7/1/2008	4	
	Pb	2.0	500ml	MET1-57-R	A2-PB02135	7/1/2008	4	
	Se	2.0	500ml	MET1-57-K	Z-SE01120	7/1/2008	4	
	Tl	2.0	500ml	MET1-59-B	704313	1/2/2009	4	
	Cu	2.0	500ml	MET1-62-G	726801	6/3/2009	4	
SS5-MET1-63-E	HNO3	25.0	500ml	JT BAKER	E27027	-	-	Expires: 7/1/08
	As	50.0	500ml	MET1-57-Z	Z-AS02032	7/1/2008	100	
	Se	50.0	500ml	MET1-59-C	Z-SE01120	8/1/2008	100	
	Tl	50.0	500ml	MET1-59-B	704313	1/2/2009	100	
SS6-MET1-63-F	HNO3	25	500ml	JT BAKER	E27027	-	-	Expires: 9/26/08
	B	50	500ml	MET1-59-A	715006	1/2/2009	100	
	Mo	50	500ml	MET1-55-C	705726	9/26/2008	100	

GFLCSW (MET1-60-A)	HNO3	10.0	1000ml	JT BAKER	E01042	-	-	Expires: 3/1/08
	As, Pb, Se, Tl	5.0	1000ml	QCP-CICV-3	Z-CICP19048	3/1/2008	2.5	
	Cd	-	-	QCP-CICV-3	Z-CICP19048	3/1/2008	1.25	
	Cu	2.5	1000ml	MET-55-O	Z-CU0284	5/1/2008	2.5	
QCP-CICV-1 (MET1-63-A)	Ca, Mg, Na, K	no dilution	-	IV	A2-MEB236021	1/1/2009	2500	Expires: 1/1/09
	Al, Ba	no dilution	-	IV	A2-MEB236021	1/1/2009	1000	
	Fe	no dilution	-	IV	A2-MEB236021	1/1/2009	500	
	Co, Mn, Ni, V, Zn	no dilution	-	IV	A2-MEB236021	1/1/2009	250	
	Cu, Ag	no dilution	-	IV	A2-MEB236021	1/1/2009	125	
	Cr	no dilution	-	IV	A2-MEB236021	1/1/2009	100	
	Be	no dilution	-	IV	A2-MEB236021	1/1/2009	25	
QCP-CICV-2 (MET1-63-B)	Sb	no dilution	-	IV	Z-CICP19033	1/1/2009	500	Expires: 1/1/09
QCP-CICV-3 (MET1-63-C)	As, Pb, Se, Tl	no dilution	-	IV	Z-CICP19048	1/1/2009	500	Expires: 1/1/09
	Cd	no dilution	-	IV	Z-CICP19048	1/1/2009	250	

* Denotes volume of 1000 ppm stock standard.

Element	mls of	ppm	Source	Lot# / Lab Code	Exp. Date

Element Analyzed Se Hydride Instrument K-FLAA-2
Service Request # K0712111

Batch QC SR's # _____

Calibration Std. AA1-8-A

Starlims # 105967

Run # 020508-Se

Hydride Data Review Form

	Yes	No	NA
1. ICV within 10% of true Value	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
2. Calibration data included	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
3. CCV's in control	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
4. CCB's and/or ICB's below MRL	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
5. All reported Results within Cal. Range	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>
6. All Calculations are Correct	<input checked="" type="checkbox"/>	<input type="checkbox"/>	<input type="checkbox"/>

Comments

Primary Reviewed by [Signature] Date 2/6/08

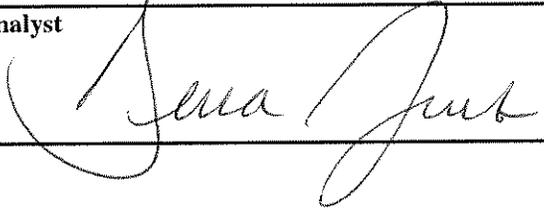
Secondary Reviewed by JDB Date 2/6/08

COLUMBIA ANALYTICAL SERVICES, INC.
GFAA Run Log

Method: (Circle Method Used) 7742 7062 Other: _____ Element: Ar Se	Service Request # :
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SAMPLE NUMBER	Dilution Factor	Measured (µg/L)	Recoveries (ICV, CCV, CRA, LCS, Matrix Spk.)	Comments
ICV	-	9.736	97%	
ICB	-	0.032		
CRA	-	0.539	108%	
CCV	-	9.855	99%	
CCB	-	0.011		
K0712111-MB	1/2	-0.101		
DOLT K0712111	1/2+1/10	1.984		Rerun on 2nd run
K0712111-001	1/2+1/5	1.385		Rerun
K0712111-001D	1/2+1/5	1.231		Rerun
K0712111-001S	1/2+1/20	3.416	77%	
K0712111-002	1/2+1/2	2.532		
K0712111-003	1/2+1/2	2.190		
K0712111-004	1/2+1/2	3.592		Rerun
K0712111-005	1/2+1/2	3.095		
K0712111-006	1/2+1/2	3.318		
CCV	-	10.039	100%	
CCB	-	0.030		
K0712111-007	1/2+1/2	2.575		
K0712111-008	1/2+1/2	2.470		
K0712111-009	1/2+1/5	13.997		
K0712111-010	1/2+1/5	26.684		Rerun
K0712111-011	1/2+1/5	27.366		Rerun
K0712111-012	1/2+1/10	13.623		
K0712111-013	1/2+1/10	14.930		
K0712111-014	1/2+1/10	10.543		
K0712111-015	1/2+1/10	9.311		
K0712111-016	1/2+1/10	16.828		Rerun
CCV	-	10.114	101%	
CCB	-	-0.006		
K0712111-017	1/2+1/10	6.771		

True Values/QC Limits:	LCSW	Water Spike	LCSS (ERA D045540)	Soil Spike
Arsenic:	2500ppb (80-120%)	1000ppb (80-120%)	146.0mg/kg (80-120%)	1000ppb (80-120%)
Selenium	2500ppb (71-122%)	1000ppb (65-122%)	73.0mg/kg (61-142%)	1000ppb (65-124%)

Analyst 	Date: 2/6/08	Page Number: 1
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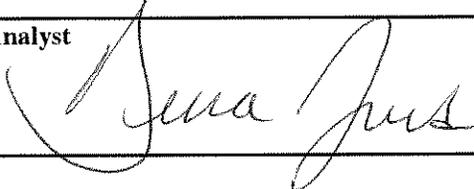
COLUMBIA ANALYTICAL SERVICES, INC.

GFAA Run Log

Method: (Circle Method Used) 7742 7062 Other: Element: As <u>Se</u>	Service Request # :
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SAMPLE NUMBER	Dilution Factor	Measured (µg/L)	Recoveries (ICV, CCV, CRA, LCS, Matrix Spk.)	Comments
K0712111-018	1/2+1/10	7.023		
K0712111-019	1/2+1/10	11.593		
K0712111-020	1/2+1/10	6.491		
K0712111-MB	1/2	-0.033		
DOLT K0712111	1/2+1/10	1.784		2/6/08 Rerun on 2nd run
K0712111-021	1/2+1/10	6.558		
K0712111-022	1/2+1/20	8.324		
K0712111-022D	1/2+1/20	8.542		
K0712111-022S	1/2+1/20	11.338	72%	
CCV	-	10.037	100%	
CCB	-	0.241		
K0712111-023	1/2+1/5	13.617		
K0712111-024	1/2+1/10	4.528		
K0712111-025	1/2+1/10	4.382		
K0712111-026	1/2+1/10	4.751		
K0712111-027	1/2+1/10	4.869		
K0712111-028	1/2+1/10	3.842		
K0712111-029	1/2+1/10	3.979		
K0712111-030	1/2+1/10	3.566		
K0712111-031	1/2+1/10	3.720		
K0712111-032	1/2+1/10	7.260		
CCV	-	9.678	97%	
CCB	-	-0.008		
K0712111-033	1/2+1/10	5.032		2/6/08 Rerun
K0712111-034	1/2+1/10	5.026		
K0712111-035	1/2+1/10	0.636		2/6/08 Rerun
K0712111-001A	1/2+1/2	4.788		2/6/08 Rerun
K0712111-022A	1/2+1/20	11.362	101%	Post Spike = 3 ppb
CCV	-	10.096	101%	
CCB	-	0.039		

True Values/QC Limits:	LCSW	Water Spike	LCSS (ERA D045540)	Soil Spike
Arsenic:	2500ppb (80-120%)	1000ppb (80-120%)	146.0mg/kg (80-120%)	1000ppb (80-120%)
Selenium	2500ppb (71-122%)	1000ppb (65-122%)	73.0mg/kg (61-142%)	1000ppb (65-124%)

Analyst 	Date: 2/6/08	Page Number: 2
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COLUMBIA ANALYTICAL SERVICES, INC.
GFAA Run Log

Method: (Circle Method Used) 7742 7062 Other: _____ Element: <u>As Se</u>	Service Request # :
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SAMPLE NUMBER	Dilution Factor	Measured (µg/L)	Recoveries (ICV, CCV, CRA, LCS, Matrix Spk.)	Comments
DOLT K0712111 1/40	1/2+1/20	1.215		See 2nd run
DOLT K0712111 1/40	1/2+1/20	1.149		See 2nd run
K0712111-001 1/2	1/2+1/2	2.189		
K0712111-001D 1/2	1/2+1/2	2.022		
K0712111-004 1/2	1/2+1/2	2.427		
K0712111-010 1/10	1/2+1/10	19.362		2/16/08 Rerun
K0712111-011 1/10	1/2+1/10	17.222		2/16/08 Rerun
K0712111-016 1/20	1/2+1/20	10.034		
K0712111-033 1/10	1/2+1/10	4.908		Rerun
K0712111-035	1/2	3.696		
CCV	-	9.911	99%	
CCB	-	0.011		
K0712111-0016A 1/20	1/2+1/20	12.404		Rerun
K0712111-033 1/10	1/2+1/10	3.923		
K0712111-010 1/20	1/2+1/10	16.898		2/16/08
K0712111-011 1/20	1/2+1/20	10.452		
K0712111-010 1/20	1/2+1/20	11.413		
DOLT K0712111 1/20	1/2+1/10	2.316		See 2nd run
DOLT K0712111	1/2+1/10	2.085		See 2nd run
K0712111-016A 1/20	1/2+1/20	12.154		Rerun
CCV	-	10.445	104%	
CCB	-	0.073		
DOLT K0712111 1/10	1/2+1/5	2.889		2/16/08 See 2nd run
K0712111-011 1/20	1/2+1/20	12.375		Rerun
K0712111-011A 1/20	1/2+1/20	12.836		Rerun
K0712111-011A 1/20	1/2+1/20	13.665	107%	Post Spike = 3 ppb
CCV	-	10.022	100%	
CCB	-	-0.085		

True Values/QC Limits:	LCSW	Water Spike	LCSS (ERA D045540)	Soil Spike
Arsenic:	2500ppb (80-120%)	1000ppb (80-120%)	146.0mg/kg (80-120%)	1000ppb (80-120%)
Selenium	2500ppb (71-122%)	1000ppb (65-122%)	73.0mg/kg (61-142%)	1000ppb (65-124%)

Analyst: <i>Jana Jant</i>	Date: <i>2/16/08</i>	Page Number: <i>3</i>
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Sample ID: Std 5.0
Analyst:

Date Collected: 2/5/2008 9:42:51 AM
Data Type: Original

Replicate Data: Std 5.0

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[5.0]	0.072	0.355	0.077			09:43:14	Yes
2		[5.0]	0.071	0.348	0.076			09:43:48	Yes
3		[5.0]	0.071	0.350	0.076			09:44:23	Yes
Mean:		[5.0]	0.071						
SD:		0.0	0.0006						
%RSD:		0.0	0.91						
Standard number 3 applied. [5.0]									
Correlation Coef.: 0.999891 Slope: 0.01425 Intercept: 0.00000									

Sequence No.: 5
Sample ID: Std 10.0
Analyst:

Autosampler Location: 5
Date Collected: 2/5/2008 9:45:13 AM
Data Type: Original

Replicate Data: Std 10.0

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[10.0]	0.139	0.690	0.144			09:45:35	Yes
2		[10.0]	0.139	0.681	0.144			09:46:10	Yes
3		[10.0]	0.142	0.689	0.147			09:46:44	Yes
Mean:		[10.0]	0.140						
SD:		0.0	0.0016						
%RSD:		0.0	1.15						
Standard number 4 applied. [10.0]									
Correlation Coef.: 0.999915 Slope: 0.01405 Intercept: 0.00000									

Sequence No.: 6
Sample ID: Std 15.0
Analyst:

Autosampler Location: 6
Date Collected: 2/5/2008 9:47:35 AM
Data Type: Original

Replicate Data: Std 15.0

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[15.0]	0.215	0.939	0.220			09:47:58	Yes
2	Changing BOC	[15.0]	0.225	1.064	0.230			09:48:33	Yes
3		[15.0]	0.222	1.031	0.227			09:49:07	Yes
Mean:		[15.0]	0.221						
SD:		0.0	0.0051						
%RSD:		0.0	2.32						
Standard number 5 applied. [15.0]									
Correlation Coef.: 0.999417 Slope: 0.01448 Intercept: 0.00000									
Changing BOC									

Calibration data for Se 196.03

Equation: Linear Through Zero

ID	Mean Signal (Abs)	Entered Conc. ug/L	Calculated Conc. ug/L	Standard Deviation	%RSD
Cal Blk	0.0000	0	0.000	0.00	6.9
Std 0.5	0.0075	0.5	0.515	0.00	2.3
Std 1.0	0.0136	1.0	0.940	0.00	3.0
Std 5.0	0.0714	5.0	4.929	0.00	0.9
Std 10.0	0.1399	10.0	9.665	0.00	1.2
Std 15.0	0.2206	15.0	15.239	0.01	2.3
Correlation Coef.: 0.999417 Slope: 0.01448 Intercept: 0.00000					

Sequence No.: 7
 Sample ID: ICV
 Analyst:

Autosampler Location: 7
 Date Collected: 2/5/2008 9:49:59 AM
 Data Type: Original

Replicate Data: ICV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.770	9.770	0.141	0.707	0.146			09:50:23	Yes
2	9.927	9.927	0.144	0.700	0.149			09:50:57	Yes
3	9.510	9.510	0.138	0.674	0.143			09:51:32	Yes
Mean:	9.736	9.736	0.141						
SD:	0.211	0.211	0.0030						
%RSD:	2.163	2.163	2.16						

QC value within limits for Se 196.03 Recovery = 97.36%
 All analyte(s) passed QC.

Sequence No.: 8
 Sample ID: ICB
 Analyst:

Autosampler Location: 1
 Date Collected: 2/5/2008 9:52:24 AM
 Data Type: Original

Replicate Data: ICB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.078	0.078	0.001	0.025	0.006			09:52:45	Yes
2	0.044	0.044	0.001	0.021	0.006			09:53:19	Yes
3	-0.025	-0.025	-0.000	0.015	0.005			09:53:53	Yes
Mean:	0.032	0.032	0.000						
SD:	0.053	0.053	0.0008						
%RSD:	163.8	163.8	163.85						

QC value within limits for Se 196.03 Recovery = Not calculated
 All analyte(s) passed QC.

Sequence No.: 9
 Sample ID: CRA
 Analyst:

Autosampler Location: 2
 Date Collected: 2/5/2008 9:54:42 AM
 Data Type: Original

Replicate Data: CRA

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.549	0.549	0.008	0.051	0.013			09:55:04	Yes
2	0.500	0.500	0.007	0.043	0.012			09:55:38	Yes
3	0.566	0.566	0.008	0.056	0.013			09:56:12	Yes
Mean:	0.539	0.539	0.008						
SD:	0.034	0.034	0.0005						
%RSD:	6.354	6.354	6.35						

QC value within limits for Se 196.03 Recovery = 107.74%
 All analyte(s) passed QC.

Sequence No.: 10
 Sample ID: CCV
 Analyst:

Autosampler Location: 5
 Date Collected: 2/5/2008 9:57:01 AM
 Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.891	9.891	0.143	0.708	0.148			09:57:24	Yes
2	9.861	9.861	0.143	0.695	0.148			09:57:58	Yes
3	9.812	9.812	0.142	0.692	0.147			09:58:32	Yes

Mean: 9.855 9.855 0.143
 SD: 0.040 0.040 0.0006
 %RSD: 0.408 0.408 0.41

QC value within limits for Se 196.03 Recovery = 98.55%
 All analyte(s) passed QC.

Sequence No.: 11
 Sample ID: CCB
 Analyst:

Autosampler Location: 1
 Date Collected: 2/5/2008 9:59:23 AM
 Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.046	0.046	0.001	0.021	0.006			09:59:44	Yes
2	0.001	0.001	0.000	0.017	0.005			10:00:18	Yes
3	-0.014	-0.014	-0.000	0.014	0.005			10:00:52	Yes
Mean:	0.011	0.011	0.000						
SD:	0.031	0.031	0.0005						
%RSD:	286.9	286.9	286.93						

QC value within limits for Se 196.03 Recovery = Not calculated
 All analyte(s) passed QC.

Sequence No.: 12
 Sample ID: K0712111-MB
 Analyst:

Autosampler Location: 9
 Date Collected: 2/5/2008 10:01:41 AM
 Data Type: Original

Replicate Data: K0712111-MB

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	-0.073	-0.073	-0.001	0.012	0.004			10:02:02	Yes
2	-0.125	-0.125	-0.002	-0.003	0.003			10:02:36	Yes
3	-0.106	-0.106	-0.002	0.009	0.003			10:03:10	Yes
Mean:	-0.101	-0.101	-0.001						
SD:	0.026	0.026	0.0004						
%RSD:	26.06	26.06	26.06						

Sequence No.: 13
 Sample ID: DOLT K0712111
 Analyst:

Autosampler Location: 10
 Date Collected: 2/5/2008 10:04:00 AM
 Data Type: Original

Replicate Data: DOLT K0712111

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	1.889	1.889	0.027	0.110	0.032			10:04:21	Yes
2	2.054	2.054	0.030	0.168	0.035			10:04:55	Yes
3	2.009	2.009	0.029	0.168	0.034			10:05:29	Yes
Mean:	1.984	1.984	0.029						
SD:	0.085	0.085	0.0012						
%RSD:	4.300	4.300	4.30						

Sequence No.: 14
 Sample ID: K0712111-001
 Analyst:

Autosampler Location: 11
 Date Collected: 2/5/2008 10:06:19 AM
 Data Type: Original

Replicate Data: K0712111-001

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	1.442	1.442	0.021	0.121	0.026			10:06:41	Yes
2	1.346	1.346	0.019	0.113	0.024			10:07:15	Yes

3	1.367	1.367	0.020	0.112	0.025	10:07:50	Yes
Mean:	1.385	1.385	0.020				
SD:	0.050	0.050	0.0007				
%RSD:	3.634	3.634	3.63				

Sequence No.: 15
 Sample ID: K0712111-001D
 Analyst:

Autosampler Location: 12
 Date Collected: 2/5/2008 10:08:40 AM
 Data Type: Original

Replicate Data: K0712111-001D

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	1.222	1.222	0.018	0.104	0.023			10:09:03	Yes
2	1.185	1.185	0.017	0.093	0.022			10:09:37	Yes
3	1.287	1.287	0.019	0.103	0.024			10:10:11	Yes
Mean:	1.231	1.231	0.018						
SD:	0.052	0.052	0.0007						
%RSD:	4.204	4.204	4.20						

Sequence No.: 16
 Sample ID: K0712111-001S
 Analyst:

Autosampler Location: 13
 Date Collected: 2/5/2008 10:11:01 AM
 Data Type: Original

Replicate Data: K0712111-001S

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.414	3.414	0.049	0.258	0.054			10:11:24	Yes
2	3.413	3.413	0.049	0.260	0.054			10:11:59	Yes
3	3.421	3.421	0.050	0.250	0.054			10:12:34	Yes
Mean:	3.416	3.416	0.049						
SD:	0.004	0.004	0.0001						
%RSD:	0.130	0.130	0.13						

Sequence No.: 17
 Sample ID: K0712111-002
 Analyst:

Autosampler Location: 14
 Date Collected: 2/5/2008 10:13:26 AM
 Data Type: Original

Replicate Data: K0712111-002

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.566	2.566	0.037	0.218	0.042			10:13:49	Yes
2	2.503	2.503	0.036	0.210	0.041			10:14:24	Yes
3	2.527	2.527	0.037	0.209	0.042			10:14:58	Yes
Mean:	2.532	2.532	0.037						
SD:	0.032	0.032	0.0005						
%RSD:	1.256	1.256	1.26						

Sequence No.: 18
 Sample ID: K0712111-003
 Analyst:

Autosampler Location: 15
 Date Collected: 2/5/2008 10:15:50 AM
 Data Type: Original

Replicate Data: K0712111-003

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.226	2.226	0.032	0.193	0.037			10:16:14	Yes
2	2.212	2.212	0.032	0.195	0.037			10:16:48	Yes
3	2.131	2.131	0.031	0.180	0.036			10:17:22	Yes
Mean:	2.190	2.190	0.032						
SD:	0.051	0.051	0.0007						

%RSD: 2.337 2.337 2.34

Sequence No.: 19
 Sample ID: K0712111-004
 Analyst:

Autosampler Location: 16
 Date Collected: 2/5/2008 10:18:14 AM
 Data Type: Original

Replicate Data: K0712111-004

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.693	2.693	0.039	0.287	0.044			10:18:38	Yes
2	4.094	4.094	0.059	0.312	0.064			10:19:13	Yes
3	3.989	3.989	0.058	0.312	0.063			10:19:47	Yes
Mean:	3.592	3.592	0.052						
SD:	0.781	0.781	0.0113						
%RSD:	21.74	21.74	21.74						

Sequence No.: 20
 Sample ID: K0712111-005
 Analyst:

Autosampler Location: 17
 Date Collected: 2/5/2008 10:20:40 AM
 Data Type: Original

Replicate Data: K0712111-005

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.179	3.179	0.046	0.273	0.051			10:21:00	Yes
2	3.059	3.059	0.044	0.254	0.049			10:21:35	Yes
3	3.048	3.048	0.044	0.251	0.049			10:22:09	Yes
Mean:	3.095	3.095	0.045						
SD:	0.072	0.072	0.0010						
%RSD:	2.341	2.341	2.34						

Sequence No.: 21
 Sample ID: K0712111-006
 Analyst:

Autosampler Location: 18
 Date Collected: 2/5/2008 10:22:57 AM
 Data Type: Original

Replicate Data: K0712111-006

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.369	3.369	0.049	0.275	0.054			10:23:18	Yes
2	3.311	3.311	0.048	0.263	0.053			10:23:52	Yes
3	3.275	3.275	0.047	0.262	0.052			10:24:27	Yes
Mean:	3.318	3.318	0.048						
SD:	0.048	0.048	0.0007						
%RSD:	1.433	1.433	1.43						

Sequence No.: 22
 Sample ID: CCV
 Analyst:

Autosampler Location: 5
 Date Collected: 2/5/2008 10:25:16 AM
 Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.04	10.04	0.145	0.704	0.150			10:25:39	Yes
2	9.969	9.969	0.144	0.693	0.149			10:26:13	Yes
3	10.10	10.10	0.146	0.713	0.151			10:26:49	Yes
Mean:	10.04	10.04	0.145						
SD:	0.068	0.068	0.0010						
%RSD:	0.676	0.676	0.68						

QC value within limits for Se 196.03 Recovery = 100.39%
 All analyte(s) passed QC.

Sequence No.: 23
 Sample ID: CCB
 Analyst:

Autosampler Location: 1
 Date Collected: 2/5/2008 10:27:40 AM
 Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.073	0.073	0.001	0.027	0.006			10:28:01	Yes
2	0.044	0.044	0.001	0.022	0.006			10:28:35	Yes
3	-0.026	-0.026	-0.000	0.023	0.005			10:29:10	Yes
Mean:	0.030	0.030	0.000						
SD:	0.051	0.051	0.0007						
%RSD:	169.1	169.1	169.13						

QC value within limits for Se 196.03 Recovery = Not calculated
 All analyte(s) passed QC.

Sequence No.: 24
 Sample ID: K0712111-007
 Analyst:

Autosampler Location: 19
 Date Collected: 2/5/2008 10:29:58 AM
 Data Type: Original

Replicate Data: K0712111-007

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.645	2.645	0.038	0.229	0.043			10:30:19	Yes
2	2.541	2.541	0.037	0.217	0.042			10:30:53	Yes
3	2.540	2.540	0.037	0.209	0.042			10:31:28	Yes
Mean:	2.575	2.575	0.037						
SD:	0.061	0.061	0.0009						
%RSD:	2.357	2.357	2.36						

Sequence No.: 25
 Sample ID: K0712111-008
 Analyst:

Autosampler Location: 20
 Date Collected: 2/5/2008 10:32:17 AM
 Data Type: Original

Replicate Data: K0712111-008

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.482	2.482	0.036	0.211	0.041			10:32:39	Yes
2	2.442	2.442	0.035	0.201	0.040			10:33:12	Yes
3	2.487	2.487	0.036	0.193	0.041			10:33:47	Yes
Mean:	2.470	2.470	0.036						
SD:	0.025	0.025	0.0004						
%RSD:	1.012	1.012	1.01						

Sequence No.: 26
 Sample ID: K0712111-009
 Analyst:

Autosampler Location: 21
 Date Collected: 2/5/2008 10:34:36 AM
 Data Type: Original

Replicate Data: K0712111-009

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	13.95	13.95	0.202	1.036	0.207			10:34:58	Yes
2	13.87	13.87	0.201	1.036	0.206			10:35:32	Yes
3	14.17	14.17	0.205	1.036	0.210			10:36:06	Yes
Mean:	14.00	14.00	0.203						
SD:	0.159	0.159	0.0023						
%RSD:	1.134	1.134	1.13						

Sequence No.: 27
Sample ID: K0712111-010
Analyst:

Autosampler Location: 22
Date Collected: 2/5/2008 10:36:57 AM
Data Type: Original

Replicate Data: K0712111-010

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	26.60	26.60	0.385	2.023	0.390			10:37:19	Yes
Sample concentration is greater than that of the highest standard.									
2	26.80	26.80	0.388	2.011	0.393			10:37:53	Yes
Sample concentration is greater than that of the highest standard.									
3	26.65	26.65	0.386	1.983	0.391			10:38:28	Yes
Sample concentration is greater than that of the highest standard.									
Mean:	26.68	26.68	0.386						
SD:	0.105	0.105	0.0015						
%RSD:	0.392	0.392	0.39						
Sample concentration is greater than that of the highest standard.									

Sequence No.: 28
Sample ID: K0712111-011
Analyst:

Handwritten: K 2/5/08

Autosampler Location: 23
Date Collected: 2/5/2008 10:39:52 AM
Data Type: Original

Replicate Data: K0712111-011

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	25.51	25.51	0.369	1.881	0.374			10:40:14	Yes
Sample concentration is greater than that of the highest standard.									
2	29.48	29.48	0.427	2.783	0.432			10:40:48	Yes
Sample concentration is greater than that of the highest standard.									
3	27.11	27.11	0.392	1.964	0.397			10:41:22	Yes
Sample concentration is greater than that of the highest standard.									
Mean:	27.37	27.37	0.396						
SD:	1.994	1.994	0.0289						
%RSD:	7.287	7.287	7.29						
Sample concentration is greater than that of the highest standard.									

Sequence No.: 29
Sample ID: K0712111-012
Analyst:

Autosampler Location: 24
Date Collected: 2/5/2008 10:42:45 AM
Data Type: Original

Replicate Data: K0712111-012

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	14.20	14.20	0.205	0.980	0.210			10:43:08	Yes
2	13.28	13.28	0.192	0.931	0.197			10:43:42	Yes
3	13.39	13.39	0.194	0.935	0.199			10:44:16	Yes
Mean:	13.62	13.62	0.197						
SD:	0.499	0.499	0.0072						
%RSD:	3.664	3.664	3.66						

Sequence No.: 30
Sample ID: K0712111-013
Analyst:

Autosampler Location: 25
Date Collected: 2/5/2008 10:45:07 AM
Data Type: Original

Replicate Data: K0712111-013

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	15.08	15.08	0.218	1.055	0.223			10:45:30	Yes
2	14.90	14.90	0.216	1.043	0.221			10:46:04	Yes

1	11.49	11.49	0.166	0.825	0.171	11:05:04	Yes
2	11.39	11.39	0.165	0.800	0.170	11:05:39	Yes
3	11.90	11.90	0.172	0.818	0.177	11:06:13	Yes
Mean:	11.59	11.59	0.168				
SD:	0.268	0.268	0.0039				
%RSD:	2.312	2.312	2.31				

Sequence No.: 39
 Sample ID: K0712111-020
 Analyst:

Autosampler Location: 32
 Date Collected: 2/5/2008 11:07:06 AM
 Data Type: Original

Replicate Data: K0712111-020

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	6.453	6.453	0.093	0.468	0.098			11:07:28	Yes
2	6.530	6.530	0.095	0.463	0.099			11:08:03	Yes
3	6.491	6.491	0.094	0.451	0.099			11:08:37	Yes
Mean:	6.491	6.491	0.094						
SD:	0.039	0.039	0.0006						
%RSD:	0.594	0.594	0.59						

Sequence No.: 40
 Sample ID: K0712111-MB
 Analyst:

Autosampler Location: 33
 Date Collected: 2/5/2008 11:09:26 AM
 Data Type: Original

Replicate Data: K0712111-MB

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	-0.040	-0.040	-0.001	0.008	0.004			11:09:46	Yes
2	-0.051	-0.051	-0.001	0.017	0.004			11:10:20	Yes
3	-0.008	-0.008	-0.000	0.015	0.005			11:10:54	Yes
Mean:	-0.033	-0.033	-0.000						
SD:	0.023	0.023	0.0003						
%RSD:	68.64	68.64	68.64						

Sequence No.: 41
 Sample ID: DOLT K0712111
 Analyst:

Autosampler Location: 34
 Date Collected: 2/5/2008 11:11:43 AM
 Data Type: Original

Replicate Data: DOLT K0712111

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	1.797	1.797	0.026	0.149	0.031			11:12:03	Yes
2	1.819	1.819	0.026	0.146	0.031			11:12:37	Yes
3	1.735	1.735	0.025	0.141	0.030			11:13:12	Yes
Mean:	1.784	1.784	0.026						
SD:	0.043	0.043	0.0006						
%RSD:	2.433	2.433	2.43						

Sequence No.: 42
 Sample ID: K0712111-021
 Analyst:

Autosampler Location: 35
 Date Collected: 2/5/2008 11:14:00 AM
 Data Type: Original

Replicate Data: K0712111-021

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	6.634	6.634	0.096	0.468	0.101			11:14:21	Yes
2	6.493	6.493	0.094	0.456	0.099			11:14:55	Yes
3	6.546	6.546	0.095	0.454	0.100			11:15:30	Yes

Mean: 6.558 6.558 0.095
SD: 0.071 0.071 0.0010
%RSD: 1.088 1.088 1.09

Sequence No.: 43
Sample ID: K0712111-022
Analyst:

Autosampler Location: 36
Date Collected: 2/5/2008 11:16:18 AM
Data Type: Original

Replicate Data: K0712111-022

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 44
Sample ID: K0712111-022D
Analyst:

Autosampler Location: 37
Date Collected: 2/5/2008 11:18:36 AM
Data Type: Original

Replicate Data: K0712111-022D

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 45
Sample ID: K0712111-022S
Analyst:

Autosampler Location: 38
Date Collected: 2/5/2008 11:20:56 AM
Data Type: Original

Replicate Data: K0712111-022S

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 46
Sample ID: CCV
Analyst:

Autosampler Location: 5
Date Collected: 2/5/2008 11:23:16 AM
Data Type: Original

Replicate Data: CCV

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

QC value within limits for Se 196.03 Recovery = 100.37%
All analyte(s) passed QC.

Sequence No.: 47
Sample ID: CCB
Analyst:

Autosampler Location: 1
Date Collected: 2/5/2008 11:25:39 AM
Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.068	0.068	0.001	0.021	0.006			11:26:00	Yes
2	0.025	0.025	0.000	0.010	0.005			11:26:34	Yes
3	0.631	0.631	0.009	0.105	0.014			11:27:08	Yes
Mean:	0.241	0.241	0.003						
SD:	0.338	0.338	0.0049						
%RSD:	140.0	140.0	139.96						

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Sequence No.: 48
Sample ID: K0712111-023
Analyst:

Autosampler Location: 39
Date Collected: 2/5/2008 11:27:57 AM
Data Type: Original

Replicate Data: K0712111-023

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	13.51	13.51	0.196	1.026	0.200			11:28:19	Yes
2	13.78	13.78	0.199	1.022	0.204			11:28:53	Yes
3	13.57	13.57	0.196	0.953	0.201			11:29:27	Yes
Mean:	13.62	13.62	0.197						
SD:	0.141	0.141	0.0020						
%RSD:	1.035	1.035	1.03						

Sequence No.: 49
Sample ID: K0712111-024
Analyst:

Autosampler Location: 40
Date Collected: 2/5/2008 11:30:17 AM
Data Type: Original

Replicate Data: K0712111-024

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.631	4.631	0.067	0.358	0.072			11:30:40	Yes
2	4.502	4.502	0.065	0.336	0.070			11:31:14	Yes
3	4.453	4.453	0.064	0.326	0.069			11:31:49	Yes
Mean:	4.528	4.528	0.066						
SD:	0.092	0.092	0.0013						
%RSD:	2.027	2.027	2.03						

Sequence No.: 50
Sample ID: K0712111-025
Analyst:

Autosampler Location: 41
Date Collected: 2/5/2008 11:32:39 AM
Data Type: Original

Replicate Data: K0712111-025

Repl #	SampleConc ug/L	StdConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.514	4.514	0.065	0.333	0.070			11:33:02	Yes
2	4.423	4.423	0.064	0.320	0.069			11:33:36	Yes
3	4.210	4.210	0.061	0.289	0.066			11:34:11	Yes
Mean:	4.382	4.382	0.063						
SD:	0.156	0.156	0.0023						

%RSD: 3.566 3.566 3.57

Sequence No.: 51
Sample ID: K0712111-026
Analyst:

Autosampler Location: 42
Date Collected: 2/5/2008 11:35:00 AM
Data Type: Original

Replicate Data: K0712111-026

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.719	4.719	0.068	0.353	0.073			11:35:23	Yes
2	4.781	4.781	0.069	0.350	0.074			11:35:57	Yes
3	4.753	4.753	0.069	0.350	0.074			11:36:32	Yes
Mean:	4.751	4.751	0.069						
SD:	0.031	0.031	0.0005						
%RSD:	0.657	0.657	0.66						

Sequence No.: 52
Sample ID: K0712111-027
Analyst:

Autosampler Location: 43
Date Collected: 2/5/2008 11:37:25 AM
Data Type: Original

Replicate Data: K0712111-027

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.929	4.929	0.071	0.352	0.076			11:37:47	Yes
2	4.865	4.865	0.070	0.350	0.075			11:38:22	Yes
3	4.813	4.813	0.070	0.335	0.075			11:38:56	Yes
Mean:	4.869	4.869	0.070						
SD:	0.058	0.058	0.0008						
%RSD:	1.194	1.194	1.19						

Sequence No.: 53
Sample ID: K0712111-028
Analyst:

Autosampler Location: 44
Date Collected: 2/5/2008 11:39:47 AM
Data Type: Original

Replicate Data: K0712111-028

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.851	3.851	0.056	0.280	0.061			11:40:10	Yes
2	3.863	3.863	0.056	0.273	0.061			11:40:44	Yes
3	3.810	3.810	0.055	0.281	0.060			11:41:19	Yes
Mean:	3.842	3.842	0.056						
SD:	0.028	0.028	0.0004						
%RSD:	0.725	0.725	0.73						

Sequence No.: 54
Sample ID: K0712111-029
Analyst:

Autosampler Location: 45
Date Collected: 2/5/2008 11:42:10 AM
Data Type: Original

Replicate Data: K0712111-029

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.012	4.012	0.058	0.295	0.063			11:42:35	Yes
2	3.987	3.987	0.058	0.291	0.063			11:43:10	Yes
3	3.937	3.937	0.057	0.288	0.062			11:43:44	Yes
Mean:	3.979	3.979	0.058						
SD:	0.038	0.038	0.0006						
%RSD:	0.959	0.959	0.96						

Sequence No.: 55
 Sample ID: K0712111-030
 Analyst:

Autosampler Location: 46
 Date Collected: 2/5/2008 11:44:35 AM
 Data Type: Original

 Replicate Data: K0712111-030

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.570	3.570	0.052	0.263	0.057			11:44:59	Yes
2	3.679	3.679	0.053	0.272	0.058			11:45:34	Yes
3	3.449	3.449	0.050	0.256	0.055			11:46:08	Yes
Mean:	3.566	3.566	0.052						
SD:	0.115	0.115	0.0017						
%RSD:	3.228	3.228	3.23						

Sequence No.: 56
 Sample ID: K0712111-031
 Analyst:

Autosampler Location: 47
 Date Collected: 2/5/2008 11:47:00 AM
 Data Type: Original

 Replicate Data: K0712111-031

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.714	3.714	0.054	0.290	0.059			11:47:24	Yes
2	3.710	3.710	0.054	0.275	0.059			11:47:58	Yes
3	3.737	3.737	0.054	0.272	0.059			11:48:33	Yes
Mean:	3.720	3.720	0.054						
SD:	0.015	0.015	0.0002						
%RSD:	0.399	0.399	0.40						

Sequence No.: 57
 Sample ID: K0712111-032
 Analyst:

Autosampler Location: 48
 Date Collected: 2/5/2008 11:49:24 AM
 Data Type: Original

 Replicate Data: K0712111-032

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	7.293	7.293	0.106	0.525	0.111			11:49:45	Yes
2	7.261	7.261	0.105	0.512	0.110			11:50:19	Yes
3	7.225	7.225	0.105	0.511	0.110			11:50:54	Yes
Mean:	7.260	7.260	0.105						
SD:	0.034	0.034	0.0005						
%RSD:	0.468	0.468	0.47						

Sequence No.: 58
 Sample ID: CCV
 Analyst:

Autosampler Location: 5
 Date Collected: 2/5/2008 11:51:42 AM
 Data Type: Original

 Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.649	9.649	0.140	0.696	0.145			11:52:05	Yes
2	9.777	9.777	0.142	0.682	0.146			11:52:39	Yes
3	9.608	9.608	0.139	0.689	0.144			11:53:14	Yes
Mean:	9.678	9.678	0.140						
SD:	0.088	0.088	0.0013						
%RSD:	0.911	0.911	0.91						

QC value within limits for Se 196.03 Recovery = 96.78%
 All analyte(s) passed QC.

Sequence No.: 59

Autosampler Location: 1

Sample ID: CCB
Analyst:

Date Collected: 2/5/2008 11:54:05 AM
Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StdConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.023	0.023	0.000	0.023	0.005			11:54:26	Yes
2	0.008	0.008	0.000	0.026	0.005			11:55:00	Yes
3	-0.055	-0.055	-0.001	0.013	0.004			11:55:34	Yes
Mean:	-0.008	-0.008	-0.000						
SD:	0.042	0.042	0.0006						
%RSD:	509.2	509.2	509.23						

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Sequence No.: 60

Sample ID: K0712111-033

Analyst:

Autosampler Location: 49

Date Collected: 2/5/2008 11:56:23 AM

Data Type: Original

Replicate Data: K0712111-033

Repl #	SampleConc ug/L	StdConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.468	9.468	0.137	0.486	0.142			11:56:44	Yes
2	1.032	1.032	0.015	-0.484	0.020			11:57:18	Yes
Changing BOC									
3	4.595	4.595	0.067	0.308	0.071			11:57:53	Yes
Mean:	5.032	5.032	0.073						
SD:	4.235	4.235	0.0613						
%RSD:	84.17	84.17	84.17						
Changing BOC									

Sequence No.: 61

Sample ID: K0712111-034

Analyst:

Autosampler Location: 50

Date Collected: 2/5/2008 11:58:41 AM

Data Type: Original

Replicate Data: K0712111-034

Repl #	SampleConc ug/L	StdConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.975	4.975	0.072	0.331	0.077			11:59:02	Yes
2	4.986	4.986	0.072	0.312	0.077			11:59:36	Yes
3	5.117	5.117	0.074	0.352	0.079			12:00:10	Yes
Mean:	5.026	5.026	0.073						
SD:	0.079	0.079	0.0011						
%RSD:	1.577	1.577	1.58						

Sequence No.: 62

Sample ID: K0712111-035

Analyst:

Autosampler Location: 51

Date Collected: 2/5/2008 12:00:59 PM

Data Type: Original

Replicate Data: K0712111-035

Repl #	SampleConc ug/L	StdConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.742	0.742	0.011	0.072	0.016			12:01:20	Yes
2	0.582	0.582	0.008	0.053	0.013			12:01:54	Yes
3	0.584	0.584	0.008	0.064	0.013			12:02:28	Yes
Mean:	0.636	0.636	0.009						
SD:	0.092	0.092	0.0013						
%RSD:	14.41	14.41	14.41						

Sequence No.: 63
 Sample ID: K0712111-001A
 Analyst:

Autosampler Location: 52
 Date Collected: 2/5/2008 12:03:17 PM
 Data Type: Original

A2/6/08

Replicate Data: K0712111-001A

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.743	4.743	0.069	0.387	0.074			12:03:38	Yes
2	4.689	4.689	0.068	0.365	0.073			12:04:12	Yes
3	4.932	4.932	0.071	0.387	0.076			12:04:46	Yes
Mean:	4.788	4.788	0.069						
SD:	0.127	0.127	0.0018						
%RSD:	2.660	2.660	2.66						

Sequence No.: 64
 Sample ID: K0712111-022A
 Analyst:

Autosampler Location: 53
 Date Collected: 2/5/2008 12:05:35 PM
 Data Type: Original

Replicate Data: K0712111-022A

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.94	10.94	0.158	0.767	0.163			12:05:56	Yes
2	11.00	11.00	0.159	0.766	0.164			12:06:30	Yes
3	12.15	12.15	0.176	0.780	0.181			12:07:05	Yes
Mean:	11.36	11.36	0.164						
SD:	0.686	0.686	0.0099						
%RSD:	6.037	6.037	6.04						

Sequence No.: 65
 Sample ID: CCV
 Analyst:

Autosampler Location: 5
 Date Collected: 2/5/2008 12:07:55 PM
 Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.05	10.05	0.145	0.733	0.150			12:08:18	Yes
2	10.39	10.39	0.150	0.731	0.155			12:08:51	Yes
3	9.847	9.847	0.143	0.724	0.147			12:09:26	Yes
Mean:	10.10	10.10	0.146						
SD:	0.276	0.276	0.0040						
%RSD:	2.734	2.734	2.73						

QC value within limits for Se 196.03 Recovery = 100.96%
 All analyte(s) passed QC.

Sequence No.: 66
 Sample ID: CCB
 Analyst:

Autosampler Location: 1
 Date Collected: 2/5/2008 12:10:17 PM
 Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.094	0.094	0.001	0.026	0.006			12:10:38	Yes
2	0.011	0.011	0.000	0.017	0.005			12:11:12	Yes
3	0.012	0.012	0.000	0.020	0.005			12:11:46	Yes
Mean:	0.039	0.039	0.001						
SD:	0.048	0.048	0.0007						
%RSD:	121.8	121.8	121.83						

QC value within limits for Se 196.03 Recovery = Not calculated
 All analyte(s) passed QC.

#	ug/L	ug/L	Signal	Area	Height	Area	Height	Time	Stored
1	1.999	1.999	0.029	0.168	0.034			12:22:42	Yes
2	2.039	2.039	0.030	0.170	0.034			12:23:16	Yes
3	2.029	2.029	0.029	0.174	0.034			12:23:50	Yes
Mean:	2.022	2.022	0.029						
SD:	0.020	0.020	0.0003						
%RSD:	1.010	1.010	1.01						

Sequence No.: 71
 Sample ID: K0712111-004 1/2
 Analyst:

Autosampler Location:
 Date Collected: 2/5/2008 12:24:59 PM
 Data Type: Original

Replicate Data: K0712111-004 1/2

Repl #	SampleConc ug/L	StdConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.505	2.505	0.036	0.206	0.041			12:25:15	Yes
2	2.383	2.383	0.035	0.201	0.039			12:25:50	Yes
3	2.392	2.392	0.035	0.198	0.040			12:26:25	Yes
Mean:	2.427	2.427	0.035						
SD:	0.068	0.068	0.0010						
%RSD:	2.794	2.794	2.79						

Sequence No.: 72
 Sample ID: K0712111-010 1/10
 Analyst:

Autosampler Location:
 Date Collected: 2/5/2008 12:27:23 PM
 Data Type: Original

Replicate Data: K0712111-010 1/10

Repl #	SampleConc ug/L	StdConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	17.40	17.40	0.252	1.241	0.257			12:27:39	Yes
	Sample concentration is greater than that of the highest standard.								
2	17.51	17.51	0.253	1.256	0.258			12:28:14	Yes
	Sample concentration is greater than that of the highest standard.								
3	23.17	23.17	0.335	1.995	0.340			12:28:48	Yes
	Sample concentration is greater than that of the highest standard.								
Mean:	19.36	19.36	0.280						
SD:	3.301	3.301	0.0478						
%RSD:	17.05	17.05	17.05						
	Sample concentration is greater than that of the highest standard.								

Sequence No.: 73
 Sample ID: K0712111-011 1/10
 Analyst:

A 2/6/08

Autosampler Location:
 Date Collected: 2/5/2008 12:30:16 PM
 Data Type: Original

Replicate Data: K0712111-011 1/10

Repl #	SampleConc ug/L	StdConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	17.22	17.22	0.249	1.168	0.254			12:30:32	Yes
	Sample concentration is greater than that of the highest standard.								
2	17.15	17.15	0.248	1.219	0.253			12:31:07	Yes
	Sample concentration is greater than that of the highest standard.								
3	17.30	17.30	0.250	1.211	0.255			12:31:42	Yes
	Sample concentration is greater than that of the highest standard.								
Mean:	17.22	17.22	0.249						
SD:	0.076	0.076	0.0011						
%RSD:	0.443	0.443	0.44						
	Sample concentration is greater than that of the highest standard.								

Sequence No.: 74
 Sample ID: K0712111-016 1/20

Autosampler Location:
 Date Collected: 2/5/2008 12:32:53 PM

Analyst:

Data Type: Original

Replicate Data: K0712111-016 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.03	10.03	0.145	0.699	0.150			12:33:10	Yes
2	10.05	10.05	0.145	0.692	0.150			12:33:44	Yes
3	10.02	10.02	0.145	0.680	0.150			12:34:18	Yes
Mean:	10.03	10.03	0.145						
SD:	0.015	0.015	0.0002						
%RSD:	0.153	0.153	0.15						

Sequence No.: 75

Sample ID: K0712111-033 1/10

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 12:35:17 PM

Data Type: Original

A 2/6/08

Replicate Data: K0712111-033 1/10

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.066	4.066	0.059	0.297	0.064			12:35:33	Yes
2	5.678	5.678	0.082	0.545	0.087			12:36:07	Yes
3	4.981	4.981	0.072	0.325	0.077			12:36:42	Yes
Mean:	4.908	4.908	0.071						
SD:	0.809	0.809	0.0117						
%RSD:	16.48	16.48	16.48						

Sequence No.: 76

Sample ID: K0712111-035

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 12:37:40 PM

Data Type: Original

Replicate Data: K0712111-035

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.664	3.664	0.053	0.316	0.058			12:37:57	Yes
2	3.704	3.704	0.054	0.271	0.059			12:38:31	Yes
3	3.721	3.721	0.054	0.306	0.059			12:39:05	Yes
Mean:	3.696	3.696	0.054						
SD:	0.029	0.029	0.0004						
%RSD:	0.786	0.786	0.79						

Sequence No.: 77

Sample ID: CCV

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 12:40:05 PM

Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.569	9.569	0.139	0.683	0.143			12:40:21	Yes
2	10.35	10.35	0.150	0.699	0.155			12:40:56	Yes
3	9.816	9.816	0.142	0.707	0.147			12:41:30	Yes
Mean:	9.911	9.911	0.143						
SD:	0.399	0.399	0.0058						
%RSD:	4.024	4.024	4.02						

Sequence No.: 78

Sample ID: CCB

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 12:42:27 PM

Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.062	0.062	0.001	0.020	0.006			12:42:43	Yes
2	0.007	0.007	0.000	0.018	0.005			12:43:17	Yes
3	-0.036	-0.036	-0.001	0.017	0.004			12:43:52	Yes
Mean:	0.011	0.011	0.000						
SD:	0.049	0.049	0.0007						
%RSD:	439.1	439.1	439.14						

Sequence No.: 79

Sample ID: K0712111-0016A 1/20

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 1:03:20 PM

Data Type: Original

2/6/08

Replicate Data: K0712111-0016A 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	12.40	12.40	0.179	0.851	0.184			13:03:37	Yes
2	12.68	12.68	0.184	0.852	0.188			13:04:11	Yes
3	12.13	12.13	0.176	0.848	0.181			13:04:46	Yes
Mean:	12.40	12.40	0.180						
SD:	0.273	0.273	0.0039						
%RSD:	2.199	2.199	2.20						

Sequence No.: 80

Sample ID: K0712111-033 1/10

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 1:06:05 PM

Data Type: Original

Replicate Data: K0712111-033 1/10

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.018	4.018	0.058	0.300	0.063			13:06:21	Yes
2	3.843	3.843	0.056	0.283	0.061			13:06:56	Yes
3	3.910	3.910	0.057	0.281	0.062			13:07:32	Yes
Mean:	3.923	3.923	0.057						
SD:	0.089	0.089	0.0013						
%RSD:	2.256	2.256	2.26						

Sequence No.: 81

Sample ID: K0712111-010 1/20

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 1:08:54 PM

Data Type: Original

2/6/08

Replicate Data: K0712111-010 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	17.04	17.04	0.247	1.258	0.252			13:09:10	Yes
2	16.74	16.74	0.242	1.235	0.247			13:09:45	Yes
3	16.92	16.92	0.245	1.371	0.250			13:10:19	Yes
Mean:	16.90	16.90	0.245						
SD:	0.149	0.149	0.0022						
%RSD:	0.880	0.880	0.88						

Sample concentration is greater than that of the highest standard.

Sample concentration is greater than that of the highest standard.

Sample concentration is greater than that of the highest standard.

Sequence No.: 82

Sample ID: K0712111-011 1/20

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 1:11:29 PM

Data Type: Original

Replicate Data: K0712111-011 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.63	10.63	0.154	0.738	0.159			13:11:45	Yes
2	10.40	10.40	0.151	0.729	0.155			13:12:20	Yes
3	10.32	10.32	0.149	0.717	0.154			13:12:54	Yes
Mean:	10.45	10.45	0.151						
SD:	0.162	0.162	0.0023						
%RSD:	1.549	1.549	1.55						

Sequence No.: 83

Sample ID: K0712111-010 1/20

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 1:13:54 PM

Data Type: Original

Replicate Data: K0712111-010 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	11.37	11.37	0.165	0.810	0.170			13:14:10	Yes
2	11.40	11.40	0.165	0.940	0.170			13:14:45	Yes
3	11.46	11.46	0.166	0.799	0.171			13:15:19	Yes
Mean:	11.41	11.41	0.165						
SD:	0.045	0.045	0.0007						
%RSD:	0.395	0.395	0.39						

Sequence No.: 84

Sample ID: DOLT K0712111 1/20

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 1:16:31 PM

Data Type: Original

Replicate Data: DOLT K0712111 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.313	2.313	0.033	0.176	0.038			13:16:48	Yes
2	2.358	2.358	0.034	0.205	0.039			13:17:22	Yes
3	2.278	2.278	0.033	0.188	0.038			13:17:57	Yes
Mean:	2.316	2.316	0.034						
SD:	0.040	0.040	0.0006						
%RSD:	1.747	1.747	1.75						

2/16/08

Sequence No.: 85

Sample ID: DOLT K0712111

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 1:19:01 PM

Data Type: Original

Replicate Data: DOLT K0712111

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.153	2.153	0.031	0.190	0.036			13:19:17	Yes
2	2.074	2.074	0.030	0.181	0.035			13:19:51	Yes
3	2.029	2.029	0.029	0.173	0.034			13:20:25	Yes
Mean:	2.085	2.085	0.030						
SD:	0.062	0.062	0.0009						
%RSD:	2.990	2.990	2.99						

Sequence No.: 86

Sample ID: K0712111-016A 1/20

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 1:24:36 PM

Data Type: Original

Replicate Data: K0712111-016A 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
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#	ug/L	ug/L	Signal	Area	Height	Area	Height	Stored
1	12.37	12.37	0.179	0.855	0.184		13:24:52	Yes
2	11.71	11.71	0.169	0.824	0.174		13:25:27	Yes
3	12.39	12.39	0.179	0.870	0.184		13:26:01	Yes
Mean:	12.15	12.15	0.176					
SD:	0.389	0.389	0.0056					
%RSD:	3.203	3.203	3.20					

2/6/08

Sequence No.: 87
Sample ID: CCV
Analyst:

Autosampler Location:
Date Collected: 2/5/2008 1:27:03 PM
Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StdConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.07	10.07	0.146	0.744	0.151			13:27:20	Yes
2	10.39	10.39	0.150	0.738	0.155			13:27:55	Yes
3	10.88	10.88	0.157	0.770	0.162			13:28:30	Yes
Mean:	10.45	10.45	0.151						
SD:	0.408	0.408	0.0059						
%RSD:	3.902	3.902	3.90						

Sequence No.: 88
Sample ID: CCB
Analyst:

Autosampler Location:
Date Collected: 2/5/2008 1:29:26 PM
Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StdConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.280	0.280	0.004	0.070	0.009			13:29:42	Yes
2	0.012	0.012	0.000	0.011	0.005			13:30:16	Yes
3	-0.074	-0.074	-0.001	0.003	0.004			13:30:51	Yes
Mean:	0.073	0.073	0.001						
SD:	0.184	0.184	0.0027						
%RSD:	253.7	253.7	253.68						

Sequence No.: 89
Sample ID: DOLT K0712111 1/10
Analyst:

Autosampler Location:
Date Collected: 2/5/2008 1:32:52 PM
Data Type: Original

2/6/08

Replicate Data: DOLT K0712111 1/10

Repl #	SampleConc ug/L	StdConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.848	2.848	0.041	0.255	0.046			13:33:08	Yes
2	2.872	2.872	0.042	0.239	0.047			13:33:42	Yes
3	2.947	2.947	0.043	0.249	0.048			13:34:16	Yes
Mean:	2.889	2.889	0.042						
SD:	0.052	0.052	0.0007						
%RSD:	1.791	1.791	1.79						

Sequence No.: 90
Sample ID: K0712111-011 1/20
Analyst:

Autosampler Location:
Date Collected: 2/5/2008 1:35:30 PM
Data Type: Original

2/6/08

Replicate Data: K0712111-011 1/20

Repl #	SampleConc ug/L	StdConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	12.08	12.08	0.175	0.854	0.180			13:35:46	Yes
2	12.32	12.32	0.178	0.879	0.183			13:36:21	Yes

3	12.73	12.73	0.184	0.870	0.189			13:36:55	Yes
Mean:	12.38	12.38	0.179						
SD:	0.326	0.326	0.0047						
%RSD:	2.638	2.638	2.64						

Sequence No.: 91

Sample ID: K0712111-011A 1/20

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 1:44:59 PM

Data Type: Original

Replicate Data: K0712111-011A 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	13.16	13.16	0.190	0.883	0.195			13:45:15	Yes
2	12.79	12.79	0.185	0.872	0.190			13:45:49	Yes
3	12.56	12.56	0.182	0.879	0.187			13:46:23	Yes
Mean:	12.84	12.84	0.186						
SD:	0.299	0.299	0.0043						
%RSD:	2.326	2.326	2.33						

Sequence No.: 92

Sample ID: K0712111-011A 1/20

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 2:14:00 PM

Data Type: Original

Replicate Data: K0712111-011A 1/20

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	13.44	13.44	0.195	0.922	0.199			14:14:17	Yes
2	13.77	13.77	0.199	0.940	0.204			14:14:51	Yes
3	13.78	13.78	0.200	0.945	0.204			14:15:25	Yes
Mean:	13.66	13.66	0.198						
SD:	0.196	0.196	0.0028						
%RSD:	1.436	1.436	1.44						

Sequence No.: 93

Sample ID: CCV

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 2:16:30 PM

Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.20	10.20	0.148	0.751	0.153			14:16:46	Yes
2	10.10	10.10	0.146	0.701	0.151			14:17:21	Yes
3	9.761	9.761	0.141	0.709	0.146			14:17:55	Yes
Mean:	10.02	10.02	0.145						
SD:	0.232	0.232	0.0034						
%RSD:	2.312	2.312	2.31						

Sequence No.: 94

Sample ID: CCB

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 2:19:09 PM

Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	-0.054	-0.054	-0.001	0.020	0.004			14:19:25	Yes
2	-0.051	-0.051	-0.001	0.025	0.004			14:19:59	Yes
3	-0.149	-0.149	-0.002	0.008	0.003			14:20:34	Yes
Mean:	-0.085	-0.085	-0.001						
SD:	0.056	0.056	0.0008						

%RSD: 65.92 65.92 65.92

Service Request Summary

Folder #: K0712111
Client Name: New Fields Environmental
Project Name: Se in Tissue
Project Number:

Report To: Kathy Tegtmeyer
 New Fields Environmental
 4720 Walnut St., Suite 200
 Boulder, CO 80301

Phone Number: 303-442-0267 Ext. 1006
Cell Number:
Fax Number:

E-mail: ktegmeyer@newfields.com

Project Chemist: Jeff Christian
Originating Lab: KEISO
Logged By: FADAIR
Date Received: 12/21/2007
Internal Due Date: 01/13/2008

QAPP: LAB QAP
Qualifier Set: CAS Standard
Formset: CAS Standard
Merged?: Y
Report to MDL?: Y
P.O. Number:
EDD: BASIC_WQC

35 - 1 each-Plastic Bag Ziplock Unpreserved
 35 - 16 oz-Glass Jar WM CLEAR Teflon Liner Unpreserved
Location: K-Yukon-04

CAS Samp No.	Client Samp No.	Matrix	Collected	Frz Dry/ Frz Dry	Metals	Homogen/ Homogen	Se T/ 7742
K0712111-001	SM1007-SNPH-FT0058	Tissue	10/25/07 1200	V	V	V	V
K0712111-002	SM1007-SNPH-FT0059	Tissue	10/25/07 1200	V	V	V	V
K0712111-003	SM1007-SNPH-FT0060	Tissue	10/25/07 1345	V	V	V	V
K0712111-004	SM1007-SNPH-FT0061	Tissue	10/25/07 1415	V	V	V	V
K0712111-005	SM1007-SNPH-FT0062	Tissue	10/25/07 1410	V	V	V	V
K0712111-006	SM1007-SNPH-FT0063	Tissue	10/25/07 1430	V	V	V	V
K0712111-007	SM1007-SNPH-FT0064	Tissue	10/25/07 1535	V	V	V	V
K0712111-008	SM1007-SNPH-FT0065	Tissue	10/25/07 1540	V	V	V	V
K0712111-009	SM1107-LSV2c-FT0066	Tissue	11/14/07	V	V	V	V
K0712111-010	SM1107-LSV2c-FT0067	Tissue	11/14/07	V	V	V	V
K0712111-011	SM1107-LSV2c-FT0068	Tissue	11/14/07	V	V	V	V
K0712111-012	SM1107-LSV2c-FT0069	Tissue	11/14/07	V	V	V	V
K0712111-013	SM1107-LSV2c-FT0070	Tissue	11/14/07	V	V	V	V
K0712111-014	SM1107-LSV2c-FT0071	Tissue	11/14/07	V	V	V	V
K0712111-015	SM1107-LSV2c-FT0072	Tissue	11/14/07	V	V	V	V
K0712111-016	SM1107-LSV2c-FT0073	Tissue	11/14/07	V	V	V	V
K0712111-017	SM1107-LSV2c-FT0074	Tissue	11/14/07	V	V	V	V
K0712111-018	SM1107-LSV2c-FT0075	Tissue	11/14/07	V	V	V	V
K0712111-019	SM1107-LSV2c-FT0076	Tissue	11/14/07	V	V	V	V
K0712111-020	SM1107-LSV2c-FT0077	Tissue	11/14/07	V	V	V	V
K0712111-021	SM1107-LSV2c-FT0078	Tissue	11/14/07	V	V	V	V
K0712111-022	SM1107-LSV2c-FT0079	Tissue	11/14/07	V	V	V	V
K0712111-023	SM1107-CC150-FT0080	Tissue	11/15/07	V	V	V	V
K0712111-024	SM1107-CC150-FT0081	Tissue	11/15/07	V	V	V	V

Service Request Summary

Folder #: K0712111
Client Name: New Fields Environmental
Project Name: Se in Tissue
Project Number:

Report To: Kathy Tegtmeyer

New Fields Environmental
 4720 Walnut St., Suite 200
 Boulder, CO 80301
Phone Number: 303-442-0267 **Ext.** 1006
Cell Number:
Fax Number:

E-mail: kegtmeyer@newfields.com

Project Chemist: Jeff Christian
Originating Lab: KELSO
Logged By: FADAIR
Date Received: 12/21/2007
Internal Due Date: 01/13/2008

QAPP: LAB QAP
Qualifier Set: CAS Standard
Formset: CAS Standard
Merged?: Y
Report to MDL?: Y

P.O. Number:
EDD: BASIC_WQC

35 - 1 each-Plastic Bag Ziplock Unpreserved
 35 - 16 oz-Glass Jar WM CLEAR Teflon Liner Unpreserved
Location: K-Yukon-04

			Metals	
			Frz Dry/ Frz Dry	Se T/ 7742
K0712111-025	SM1107-CC150-FT0082	Tissue	11/15/07	V
K0712111-026	SM1107-CC150-FT0083	Tissue	11/15/07	V
K0712111-027	SM1107-CC150-FT0084	Tissue	11/15/07	V
K0712111-028	SM1107-CC150-FT0085	Tissue	11/15/07	V
K0712111-029	SM1107-CC150-FT0086	Tissue	11/15/07	V
K0712111-030	SM1107-CC150-FT0087	Tissue	11/15/07	V
K0712111-031	SM1107-CC150-FT0088	Tissue	11/15/07	V
K0712111-032	SM1107-CC350-FT0089	Tissue	11/15/07	V
K0712111-033	SM1107-CC350-FT0090	Tissue	11/15/07	V
K0712111-034	SM1107-CC350-FT0091	Tissue	11/15/07	V
K0712111-035	SM1207-SPCFH-FT0092	Tissue	12/4/07	V

Element Analyzed Se Hydride Instrument K-FLAA-2
Service Request # K0712111

Batch QC SR's # _____

Calibration Std. AA1-8-A Analytical Batch _____
Starlims # _____ KA0630252
Run # 06014-Se

Hydride Data Review Form

	Yes	No	NA
1. ICV within 10% of true Value	<u>X</u>	_____	_____
2. Calibration data included	<u>X</u>	_____	_____
3. CCV's in control	<u>X</u>	_____	_____
4. CCB's and/or ICB's below MRL	<u>X</u>	_____	_____
5. All reported Results within Cal. Range	<u>X</u>	_____	_____
6. All Calculations are Correct	<u>X</u>	_____	_____

Comments

Primary Reviewed by [Signature] Date 2/6/08
Secondary Reviewed by JOB Date 2/6/08

Sample ID: Std 5.0
Analyst:

Date Collected: 2/5/2008 4:39:08 PM
Data Type: Original

Replicate Data: Std 5.0

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[5.0]	0.072	0.374	0.077			16:39:31	Yes
2		[5.0]	0.070	0.365	0.075			16:40:05	Yes
3		[5.0]	0.071	0.372	0.077			16:40:40	Yes
Mean:		[5.0]	0.071						
SD:		0.0	0.0010						
%RSD:		0.0	1.42						

Standard number 3 applied. [5.0]
Correlation Coef.: 0.999990 Slope: 0.01421 Intercept: 0.00000

=====

Sequence No.: 5
Sample ID: Std 10.0
Analyst:

Autosampler Location: 5
Date Collected: 2/5/2008 4:41:30 PM
Data Type: Original

Replicate Data: Std 10.0

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[10.0]	0.137	0.722	0.143			16:41:53	Yes
2		[10.0]	0.136	0.707	0.141			16:42:27	Yes
3		[10.0]	0.138	0.702	0.143			16:43:01	Yes
Mean:		[10.0]	0.137						
SD:		0.0	0.0011						
%RSD:		0.0	0.77						

Standard number 4 applied. [10.0]
Correlation Coef.: 0.999771 Slope: 0.01382 Intercept: 0.00000

=====

Sequence No.: 6
Sample ID: Std 15.0
Analyst:

Autosampler Location: 6
Date Collected: 2/5/2008 4:43:53 PM
Data Type: Original

Replicate Data: Std 15.0

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[15.0]	0.194	1.045	0.199			16:44:16	Yes
2		[15.0]	0.191	1.001	0.196			16:44:50	Yes
3		[15.0]	0.196	1.006	0.201			16:45:24	Yes
Mean:		[15.0]	0.194						
SD:		0.0	0.0024						
%RSD:		0.0	1.23						

Standard number 5 applied. [15.0]
Correlation Coef.: 0.998645 Slope: 0.01325 Intercept: 0.00000
The calibration curve may not be linear.

Calibration data for Se 196.03

Equation: Linear Through Zero

ID	Mean Signal (Abs)	Entered Conc. ug/L	Calculated Conc. ug/L	Standard Deviation	%RSD
Cal Blk	0.0000	0	0.000	0.00	8.6
Std 0.5	0.0073	0.5	0.549	0.00	3.5
Std 1.0	0.0141	1.0	1.061	0.00	4.7
Std 5.0	0.0711	5.0	5.363	0.00	1.4
Std 10.0	0.1372	10.0	10.353	0.00	0.8
Std 15.0	0.1936	15.0	14.611	0.00	1.2

Correlation Coef.: 0.998645 Slope: 0.01325 Intercept: 0.00000

Sequence No.: 7
 Sample ID: ICV
 Analyst:

Autosampler Location: 7
 Date Collected: 2/5/2008 4:46:16 PM
 Data Type: Original

Replicate Data: ICV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.18	10.18	0.135	0.712	0.140			16:46:40	Yes
2	9.926	9.926	0.132	0.694	0.137			16:47:14	Yes
3	9.999	9.999	0.132	0.687	0.138			16:47:50	Yes
Mean:	10.03	10.03	0.133						
SD:	0.129	0.129	0.0017						
%RSD:	1.287	1.287	1.29						

QC value within limits for Se 196.03 Recovery = 100.34%
 All analyte(s) passed QC.

Sequence No.: 8
 Sample ID: ICB
 Analyst:

Autosampler Location: 1
 Date Collected: 2/5/2008 4:48:42 PM
 Data Type: Original

Replicate Data: ICB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.098	0.098	0.001	0.020	0.007			16:49:03	Yes
2	0.102	0.102	0.001	0.025	0.007			16:49:37	Yes
3	0.073	0.073	0.001	0.032	0.006			16:50:11	Yes
Mean:	0.091	0.091	0.001						
SD:	0.016	0.016	0.0002						
%RSD:	17.25	17.25	17.25						

QC value within limits for Se 196.03 Recovery = Not calculated
 All analyte(s) passed QC.

Sequence No.: 9
 Sample ID: CRA
 Analyst:

Autosampler Location: 2
 Date Collected: 2/5/2008 4:51:01 PM
 Data Type: Original

Replicate Data: CRA

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.522	0.522	0.007	0.052	0.012			16:51:22	Yes
2	0.587	0.587	0.008	0.061	0.013			16:51:56	Yes
3	0.512	0.512	0.007	0.058	0.012			16:52:30	Yes
Mean:	0.540	0.540	0.007						
SD:	0.041	0.041	0.0005						
%RSD:	7.514	7.514	7.51						

QC value within limits for Se 196.03 Recovery = 108.02%
 All analyte(s) passed QC.

Sequence No.: 10
 Sample ID: CCV
 Analyst:

Autosampler Location: 5
 Date Collected: 2/5/2008 4:53:20 PM
 Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.35	10.35	0.137	0.720	0.142			16:53:43	Yes
2	10.06	10.06	0.133	0.702	0.139			16:54:17	Yes
3	10.29	10.29	0.136	0.708	0.142			16:54:52	Yes
Mean:	10.23	10.23	0.136						

SD: 0.150 0.150 0.0020
%RSD: 1.467 1.467 1.47

QC value within limits for Se 196.03 Recovery = 102.34%
All analyte(s) passed QC.

Sequence No.: 11
Sample ID: CCB
Analyst:

Autosampler Location: 1
Date Collected: 2/5/2008 4:55:43 PM
Data Type: Original

Replicate Data: CCB

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Sequence No.: 12
Sample ID: LCS-DOLT K0712111 (1)
Analyst:

Autosampler Location: 9
Date Collected: 2/5/2008 4:58:01 PM
Data Type: Original

Replicate Data: LCS-DOLT K0712111 (1)

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 13
Sample ID: LCS-DOLT K0712111 (2)
Analyst:

Autosampler Location: 10
Date Collected: 2/5/2008 5:00:20 PM
Data Type: Original

Replicate Data: LCS-DOLT K0712111 (2)

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

See Run
JAS 2/5/08

Sequence No.: 14
Sample ID: CCV
Analyst:

Autosampler Location: 5
Date Collected: 2/5/2008 5:02:39 PM
Data Type: Original

Replicate Data: CCV

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 1 replicate row and summary statistics.

2	10.41	10.41	0.138	0.708	0.143	17:03:37	Yes
3	10.38	10.38	0.138	0.714	0.143	17:04:11	Yes
Mean:	10.42	10.42	0.138				
SD:	0.045	0.045	0.0006				
%RSD:	0.431	0.431	0.43				

QC value within limits for Se 196.03 Recovery = 104.18%
All analyte(s) passed QC.

Sequence No.: 15

Autosampler Location: 1

Sample ID: CCB

Date Collected: 2/5/2008 5:05:02 PM

Analyst:

Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StdConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.115	0.115	0.002	0.025	0.007			17:05:23	Yes
2	0.030	0.030	0.000	0.017	0.006			17:05:57	Yes
3	0.016	0.016	0.000	0.016	0.005			17:06:31	Yes
Mean:	0.054	0.054	0.001						
SD:	0.054	0.054	0.0007						
%RSD:	100.0	100.0	100.04						

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Analysis Begun

Logged In Analyst: ACQMET10

Technique: AA FIAS-Flame

Spectrometer Model: AAnalyst 200, S/N 200S5061701

Autosampler Model: AS-90

Sample Information File: C:\data-AA\ACQMET10\Sample Information\020508-Se2.sif

Batch ID: 020508-Se2

Results Data Set: 060614-Se

Results Library: R:\ICP\WIP\DATA\K-FLAA-02\Results.mdb

Sequence No.: 16

Autosampler Location:

Sample ID: LCS-DOLT K0712111 (2)

Date Collected: 2/5/2008 5:09:09 PM

Analyst:

Data Type: Original

Replicate Data: LCS-DOLT K0712111 (2)

Repl #	SampleConc ug/L	StdConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	4.399	4.399	0.058	0.321	0.063			17:09:25	Yes
2	4.441	4.441	0.059	0.312	0.064			17:09:59	Yes
3	4.341	4.341	0.058	0.305	0.063			17:10:34	Yes
Mean:	4.393	4.393	0.058						
SD:	0.050	0.050	0.0007						
%RSD:	1.142	1.142	1.14						

Sequence No.: 17

Autosampler Location:

Sample ID: CCV

Date Collected: 2/5/2008 5:12:20 PM

Analyst:

Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StdConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.37	10.37	0.137	0.730	0.143			17:12:36	Yes
2	10.29	10.29	0.136	0.719	0.142			17:13:10	Yes
3	10.16	10.16	0.135	0.714	0.140			17:13:44	Yes
Mean:	10.28	10.28	0.136						
SD:	0.107	0.107	0.0014						

RSD: 1.042 1.042 1.04

Sequence No.: 18

Sample ID: CCB

Analyst:

Autosampler Location:

Date Collected: 2/5/2008 5:18:05 PM

Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.136	0.136	0.002	0.041	0.007			17:18:21	Yes
2	-0.026	-0.026	-0.000	0.016	0.005			17:18:55	Yes
3	0.009	0.009	0.000	0.010	0.005			17:19:30	Yes
Mean:	0.040	0.040	0.001						
SD:	0.085	0.085	0.0011						
RSD:	214.8	214.8	214.77						

December 20, 2007

Analytical Report for Service Request No: K0710142

Kathy Tegtmeyer
New Fields Environmental
4720 Walnut St., Suite 200
Boulder, CO 80301

Dear Kathy:

Enclosed are the results of the samples submitted to our laboratory on October 30, 2007. For your reference, these analyses have been assigned our service request number K0710142.

All analyses were performed according to our laboratory's quality assurance program. Where applicable, the methods cited conform to the Methods Update Rule (effective 4/11/2007), which relates to the use of analytical methods for the drinking water and waste water programs. The test results meet requirements of the NELAC standards. Exceptions are noted in the case narrative report where applicable. All results are intended to be considered in their entirety, and Columbia Analytical Services, Inc. (CAS) is not responsible for use of less than the complete report. Results apply only to the items submitted to the laboratory for analysis and individual items (samples) analyzed, as listed in the report.

Please call if you have any questions. My extension is 3316. You may also contact me via Email at JChristian@caslab.com.

Respectfully submitted,

Columbia Analytical Services, Inc.



Jeff Christian
Laboratory Director

JC/lb

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Acronyms

ASTM	American Society for Testing and Materials
A2LA	American Association for Laboratory Accreditation
CARB	California Air Resources Board
CAS Number	Chemical Abstract Service registry Number
CFC	Chlorofluorocarbon
CFU	Colony-Forming Unit
DEC	Department of Environmental Conservation
DEQ	Department of Environmental Quality
DHS	Department of Health Services
DOE	Department of Ecology
DOH	Department of Health
EPA	U. S. Environmental Protection Agency
ELAP	Environmental Laboratory Accreditation Program
GC	Gas Chromatography
GC/MS	Gas Chromatography/Mass Spectrometry
LUFT	Leaking Underground Fuel Tank
M	Modified
MCL	Maximum Contaminant Level is the highest permissible concentration of a substance allowed in drinking water as established by the USEPA.
MDL	Method Detection Limit
MPN	Most Probable Number
MRL	Method Reporting Limit
NA	Not Applicable
NC	Not Calculated
NCASI	National Council of the Paper Industry for Air and Stream Improvement
ND	Not Detected
NIOSH	National Institute for Occupational Safety and Health
PQL	Practical Quantitation Limit
RCRA	Resource Conservation and Recovery Act
SIM	Selected Ion Monitoring
TPH	Total Petroleum Hydrocarbons
tr	Trace level is the concentration of an analyte that is less than the PQL but greater than or equal to the MDL.

Inorganic Data Qualifiers

- * The result is an outlier. See case narrative.
- # The control limit criteria is not applicable. See case narrative.
- B The analyte was found in the associated method blank at a level that is significant relative to the sample result.
- E The result is an estimate amount because the value exceeded the instrument calibration range.
- J The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
 - i The MRL/MDL has been elevated due to a matrix interference.
- X See case narrative.

Metals Data Qualifiers

- # The control limit criteria is not applicable. See case narrative.
- B The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- E The percent difference for the serial dilution was greater than 10%, indicating a possible matrix interference in the sample.
- M The duplicate injection precision was not met.
- N The Matrix Spike sample recovery is not within control limits. See case narrative.
- S The reported value was determined by the Method of Standard Additions (MSA).
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
- W The post-digestion spike for furnace AA analysis is out of control limits, while sample absorbance is less than 50% of spike absorbance.
 - i The MRL/MDL has been elevated due to a matrix interference.
- X See case narrative.
- * The duplicate analysis not within control limits. See case narrative.
- + The correlation coefficient for the MSA is less than 0.995.

Organic Data Qualifiers

- * The result is an outlier. See case narrative.
- # The control limit criteria is not applicable. See case narrative.
- A A tentatively identified compound, a suspected aldol-condensation product.
- B The analyte was found in the associated method blank at a level that is significant relative to the sample result.
- C The analyte was qualitatively confirmed using GC/MS techniques, pattern recognition, or by comparing to historical data.
- D The reported result is from a dilution.
- E The result is an estimate amount because the value exceeded the instrument calibration range.
- J The result is an estimated concentration that is less than the MRL but greater than or equal to the MDL.
- N The result is presumptive. The analyte was tentatively identified, but a confirmation analysis was not performed.
- P The GC or HPLC confirmation criteria was exceeded. The relative percent difference is greater than 40% between the two analytical results (25% for CLP Pesticides).
- U The compound was analyzed for, but was not detected ("Non-detect") at or above the MRL/MDL.
 - i The MRL/MDL has been elevated due to a chromatographic interference.
- X See case narrative.

Additional Petroleum Hydrocarbon Specific Qualifiers

- F The chromatographic fingerprint of the sample matches the elution pattern of the calibration standard.
- L The chromatographic fingerprint of the sample resembles a petroleum product, but the elution pattern indicates the presence of a greater amount of lighter molecular weight constituents than the calibration standard.
- H The chromatographic fingerprint of the sample resembles a petroleum product, but the elution pattern indicates the presence of a greater amount of heavier molecular weight constituents than the calibration standard.
- O The chromatographic fingerprint of the sample resembles an oil, but does not match the calibration standard.
- Y The chromatographic fingerprint of the sample resembles a petroleum product eluting in approximately the correct carbon range, but the elution pattern does not match the calibration standard.
- Z The chromatographic fingerprint does not resemble a petroleum product.

Columbia Analytical Services, Inc.
Kelso, WA
State Certifications, Accreditations, and Licenses

Program	Number
Alaska DEC UST	UST-040
Arizona DHS	AZ0339
Arkansas - DEQ	88-0637
California DHS	2286
Colorado DPHE	-
Florida DOH	E87412
Hawaii DOH	-
Idaho DHW	-
Indiana DOH	C-WA-01
Louisiana DEQ	3016
Louisiana DHH	LA050010
Maine DHS	WA0035
Michigan DEQ	9949
Minnesota DOH	053-999-368
Montana DPHHS	CERT0047
Nevada DEP	WA35
New Jersey DEP	WA005
New Mexico ED	-
North Carolina DWQ	605
Oklahoma DEQ	9801
Oregon - DHS	WA200001
South Carolina DHEC	61002
Utah DOH	COLU
Washington DOE	C1203
Wisconsin DNR	998386840
Wyoming (EPA Region 8)	-



Case Narrative

COLUMBIA ANALYTICAL SERVICES, INC.

Client: New Fields Environmental
Project: Tissue - Se
Sample Matrix: Tissue

Service Request No.: K0710142
Date Received: 10/30/07

CASE NARRATIVE

All analyses were performed consistent with the quality assurance program of Columbia Analytical Services, Inc. (CAS). This report contains analytical results for samples designated for Tier III validation deliverables including summary forms and all of the associated raw data for each of the analyses. When appropriate to the method, method blank results have been reported with each analytical test.

Sample Receipt

Tissue samples were received for analysis at Columbia Analytical Services on 10/30/07. The samples were received in good condition and consistent with the accompanying chain of custody form. The samples were stored frozen at -20°C upon receipt at the laboratory.

Note that the samples were held until 11/29/07 pending approval from New Fields Environmental to proceed with the analysis.

Total Metals

General Comments:

The samples were freeze-dried to determine moisture and to allow complete homogenization of the dry material. The dried material was milled to a fine meal, and then sub-sampled for digestion. A thorough digestion was performed prior to instrumental analysis to convert all Selenium species to Selenate. Prior to hydride formation, the valence was adjusted by reduction to Selenite.

No anomalies associated with the analysis of these samples were observed.

Approved by _____



Date _____

12/20/07

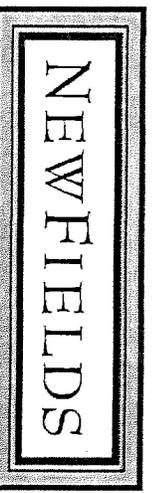
**Chain of Custody
Documentation**

Chain of Custody

Project Contact: Howard Rouse PO 0442-004-900
 Courier/Airbill: 7982 969 8081

Shipped to: COLUMBIA ANALYTICAL SERVICES, INC

Telephone: 1317 SOUTH 13TH AVE
KEESO, WA 98626
(360) 430-3733



4720 Walnut St., Suite 200
 Boulder, CO 80301
 Phone: 303-442-0267
 Fax: 303-442-3679

LAB # 102907-1 COC #: 102907-1

Sample ID	Sample Date	Sample Time	Matrix	Tox/ Diss	Analysis	Preservative	Lab QC	Comments
SM1007-CC3A-FT001	10/22/07	11:30	FISH TISSUE		SELENIUM TOTAL	NONE		ARCHIVE PENDING
SM1007-CC3A-FT001	10/22/07	11:30	FISH TISSUE		PERCENT SOLIDS	NONE		FURTHER INSTRUCTIONS
SM1007-CC3A-FT001	10/22/07	11:30	FISH TISSUE		SELENIUM, TOTAL	NONE		FROM K. FEGTMEYER,
SM1007-CC3A-FT001	10/22/07	11:30	FISH TISSUE		PERCENT SOLIDS	NONE		NEWFIELDS
SM1007-CC3A-FT101	10/24/07	11:43	FISH TISSUE		SELENIUM, TOTAL	NONE		(ALL SAMPLES)
SM1007-CC3A-FT101	10/24/07	11:43	FISH TISSUE		PERCENT SOLIDS	NONE		PLEASE
SM1007-CC3A-FT101	10/24/07	11:43	FISH TISSUE		SELENIUM, TOTAL	NONE		ARCHIVE
SM1007-CC3A-FT001	10/24/07	10:52	FISH TISSUE		PERCENT SOLIDS	NONE		THE
SM1007-CC3A-FT001	10/24/07	11:43	FISH TISSUE		SELENIUM, TOTAL	NONE		SAMPLES
SM1007-CC3A-FT001	10/24/07	11:43	FISH TISSUE		PERCENT SOLIDS	NONE		FROZEN
SM1007-CC3A-FT002	10/24/07	11:00	FISH TISSUE		SELENIUM, TOTAL	NONE		
SM1007-CC3A-FT002	10/24/07	11:00	FISH TISSUE		PERCENT SOLIDS	NONE		

Total Number of Containers: 7 Individual Hines Reflect Single Containers, Except for Aqueous Analyses Assigned as Laboratory QC KL

Sampler Signature: [Signature]

Relinquished by: KIM RABY Date/Time: 10/29/07 1200
 Received by: [Signature] Date/Time: 10/30/07 1430

LAB USE ONLY - Sample condition on Receipt

Columbia Analytical Services, Inc.
Cooler Receipt and Preservation Form

PC YC

Client / Project: Newfields Service Request K07 10/42
 Received: 10/30/07 Opened: 10/30/07 By: A.G.

Samples were received via? US Mail Fed Ex UPS DHL GH GS PDX Courier Hand Delivered
 Samples were received in: (circle) Cooler Box Envelope Other NA
 Were custody seals on coolers? NA Y N If yes, how many and where? _____
 If present, were custody seals intact? Y N If present, were they signed and dated? Y N
 Is shipper's air-bill filed? If not, record air-bill number: _____ NA Y N

Temperature of cooler(s) upon receipt (°C): 2.7
 Temperature Blank (°C): _____
 If applicable, list Chain of Custody Numbers: _____

Were custody papers properly filled out (ink, signed, etc.)? NA Y N
 Packing material used. Inserts Bubble Wrap Gel Packs Wet Ice Sleeves Other
 Did all bottles arrive in good condition (unbroken)? *Indicate in the table below.* NA Y N
 0. Were all sample labels complete (i.e analysis, preservation, etc.)? Y N
 1. Did all sample labels and tags agree with custody papers? *Indicate in the table below.* Y N
 2. Were the correct types of bottles used for the tests indicated? NA Y N
 3. Were all of the preserved bottles received at the lab with the appropriate pH? *Indicate in the table below.* NA Y N
 4. Were VOA vials and 1631 Mercury bottles checked for absence of air bubbles? *Indicate in the table below.* NA Y N
 5. Are CWA Microbiology samples received with >1/2 the 24hr. hold time remaining from collection? NA Y N
 6. Was C12/Res negative? NA Y N

Sample ID on Bottle	Sample ID on COC	Sample ID on Bottle	Sample ID on COC

Sample ID	Bottle Count	Bottle Type	Out of Temp	Head-space	Broken	pH	Reagent	Volume added	Reagent Lot Number	Initials

Additional Notes, Discrepancies, & Resolutions: _____

Total Solids

COLUMBIA ANALYTICAL SERVICES, INC.

Analytical Report

Client: New Fields Environmental
Project: NA
Sample Matrix: Tissue

Service Request: K0710142
Date Collected: 10/22-24/07
Date Received: 10/30/07

Solids, Total

Prep Method: NONE
Analysis Method: Freeze Dry
Test Notes:

Units: PERCENT
Basis: Wet

Sample Name	Lab Code	Date Analyzed	Result	Result Notes
SM1007-CC1A-FT001	K0710142-001	12/5/07	25.8	
SM107-CC1A-FT001-Eggs	K0710142-002	12/5/07	35.3	
SM1007-CC3A-FT101	K0710142-003	12/5/07	25.2	
SM1007-CC3A-FT101-Eggs	K0710142-004	12/5/07	36.1	
SM1007-CC3A-FT001	K0710142-005	12/5/07	26.7	
SM1007-CC3A-FT100	K0710142-006	12/5/07	26.5	
SM1007-CC3A-FT002	K0710142-007	12/5/07	26.8	

COLUMBIA ANALYTICAL SERVICES, INC.

QA/QC Report

Client: New Fields Environmental
Project: NA
Sample Matrix: Tissue

Service Request: K0710142
Date Collected: 10/24/07
Date Received: 10/30/07
Date Extracted: NA
Date Analyzed: 12/5/07

Duplicate Summary
Total Metals

Sample Name: SM1007-CC3A-FT002
Lab Code: K0710142-007D
Test Notes:

Units: PERCENT
Basis: Wet

Analyte	Prep Method	Analysis Method	Sample Result	Duplicate Sample Result	Average	Relative Percent Difference	Result Notes
Solids, Total	NA	Freeze Dry	26.8	26.7	26.8	<1	

Service Request #:

K0710142

Analytical Batch

Analysis For:

Freeze Dried Solids

KA0628246

Lab Code	Wet Weight (g)	Tare (g)	Tare + Dry Wt.(g)	Dry Weight (g)	% Total Solids
K0710142-01	21.875	79.545	85.199	5.054	25.8
- 02	21.178	78.684	86.157	7.473	35.3
- 03	23.435	79.384	85.290	5.900	25.2
- 04	20.020	78.417	85.651	7.234	30.1
- 05	21.707	78.991	84.778	5.787	26.7
- 06	20.832	78.845	84.372	5.527	26.5
- 07	22.294	79.152	85.124	5.974	26.8
- 07D	23.284	78.323	84.550	6.227	26.7
LAB 12/15/07					

Time In: 2:15 p.m. 12/15/07 Time Out: 4:00 p.m. 12/16/07

Comments:

Balance - 21B

\bar{x} =

RPD =

Analyst: Angela Black	Date: 12/15/07
Reviewed By: <i>[Signature]</i>	Date: 12/18/07

Sample Number(s): As Listed Service Request Number(s): K0710142

TISSUE COMPOSITION DATA

Laboratory ID	Weight (g)	Tare (g)		Matrix	Length
01 10142.01	2105.58	471.35	281.03	whole body fish	
02	50.50	113.28		eggs	
03	204.83	472.14	276.98	whole body fish	
04	34.41	113.04		eggs	
05	262.78		276.79	whole body fish	
06	302.12		281.22		
07	379.92		276.75		
LAB 12/5/07					

Comments: Balance 23

Analyst: Angela Black Date: 12/5/07
 Reviewed: Date: 12/13/07

Metals

Columbia Analytical Services

- Cover Page -
INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental
Project Name:
Project No.:

Service Request: K0710142

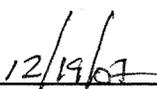
<u>Sample Name:</u>	<u>Lab Code:</u>
<u>SM1007-CC1A-FT001</u>	<u>K0710142-001</u>
<u>SM107-CC1A-FT001-Eggs</u>	<u>K0710142-002</u>
<u>SM1007-CC3A-FT101</u>	<u>K0710142-003</u>
<u>SM1007-CC3A-FT101-Eggs</u>	<u>K0710142-004</u>
<u>SM1007-CC3A-FT001</u>	<u>K0710142-005</u>
<u>SM1007-CC3A-FT100</u>	<u>K0710142-006</u>
<u>SM1007-CC3A-FT002</u>	<u>K0710142-007</u>
<u>SM1007-CC3A-FT002D</u>	<u>K0710142-007D</u>
<u>SM1007-CC3A-FT002S</u>	<u>K0710142-007S</u>
<u>Method Blank</u>	<u>K0710142-MB</u>

Comments:

Approved By:



Date:

 00016

Metals

- 1 -

INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Date Collected: 10/22/07

Project Name: NA

Date Received: 10/30/07

Matrix: TISSUE

Units: mg/Kg

Basis: DRY

Sample Name: SM1007-CC1A-FT001

Lab Code: K0710142-001

Analyte	Analysis Method	MRL	MDL	Dilution Factor	Date Extracted	Date Analyzed	Result	C	Q
Selenium	7742	0.50	0.10	10.0	12/12/07	12/19/07	6.4		

Comments:

00017

Metals

- 1 -

INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Date Collected: 10/24/07

Project Name: NA

Date Received: 10/30/07

Matrix: TISSUE

Units: mg/Kg

Basis: DRY

Sample Name: SM1007-CC3A-FT002

Lab Code: K0710142-007

Analyte	Analysis Method	MRL	MDL	Dilution Factor	Date Extracted	Date Analyzed	Result	C	Q
Selenium	7742	0.49	0.10	10.0	12/12/07	12/19/07	6.4		

Comments:

Metals

- 1 -

INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Date Collected:

Project Name: NA

Date Received:

Matrix: TISSUE

Units: mg/Kg

Basis: DRY

Sample Name: Method Blank

Lab Code: K0710142-MB

Analyte	Analysis Method	MRL	MDL	Dilution Factor	Date Extracted	Date Analyzed	Result	C	Q
Selenium	7742	0.10	0.02	2.0	12/12/07	12/19/07	0.03	B	

Comments:

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Project Name: NA

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium	10.0	10.06	101	10.0	9.87	99	9.28	93	7742

Metals

- 2a -

INITIAL AND CONTINUING CALIBRATION VERIFICATION

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Project Name: NA

ICV Source: Inorganic Ventures

CCV Source: CAS MIXED

Concentration Units: ug/L

Analyte	Initial Calibration			Continuing Calibration					Method
	True	Found	%R(1)	True	Found	%R(1)	Found	%R(1)	
Selenium				10.0	9.22	92	10.01	100	7742

Metals

- 2b -

CRDL STANDARD FOR AA AND ICP

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Project Name: NA

Concentration Units: ug/L

Analyte	CRDL Standard for AA			CRDL Standard for ICP				
	True	Found	%R	Initial		Final		
	True	Found	%R	True	Found	%R	Found	%R
Selenium	0.5	0.59	118.0					

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Project Name: NA

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)		Continuing Calibration Blank (ug/L)						Method
	C	U	1	C	2	C	3	C	
Selenium	0.1	U	0.1	U	0.1	U	0.1	U	7742

Metals

- 3 -

BLANKS

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Project Name: NA

Preparation Blank Matrix (soil/water): WATER

Preparation Blank Concentration Units (ug/L or mg/kg): UG/L

Analyte	Initial Calib. Blank (ug/L)	Continuing Calibration Blank (ug/L)						Method	
		C	1	C	2	C	3		C
Selenium			0.2	B					7742

00029

Metals

- 5A -

SPIKE SAMPLE RECOVERY

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Units: MG/KG

Project Name: NA

Basis: DRY

Matrix: TISSUE

Sample Name: SM1007-CC3A-FT002S

Lab Code: K0710142-007S

Analyte	Control Limit %R	Spike Result	C	Sample Result	C	Spike Added	%R	Q	Method
Selenium	65 - 124	22.2		6.4		15.96	99.0		7742

Metals

- 5B -

POST SPIKE SAMPLE RECOVERY

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Units: UG/L

Project Name: NA

Basis: DRY

Matrix: WATER

Sample Name: K0710642-0002A

Lab Code: K0710642-0002A

Analyte	Control Limit %R	Spike Result	C	Sample Result	C	Spike Added	%R	Q	Method
Selenium	75-125	7.383		2.762		5.0	92.4		7742

00031

Metals

- 6 -

DUPLICATES

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Units: MG/KG

Project Name: NA

Basis: DRY

Matrix: TISSUE

Sample Name: SM1007-CC3A-FT002D

Lab Code: K0710142-007D

Analyte	Control Limit	Sample (S)	C	Duplicate (D)	C	RPD	Q	Method
Selenium	30	6.4		5.6		13.3		7742

COLUMBIA ANALYTICAL SERVICES, INC.

QA/QC Report

Client: New Fields Environmental
Project: NA
LCS Matrix: Tissue

Service Request: K0710142
Date Collected: NA
Date Received: NA
Date Extracted: 12/12/07
Date Analyzed: 12/19/07

Laboratory Control Sample Summary
Total Metals

Sample Name: Laboratory Control Sample
Lab Code: K0710142-LCS
Test Notes:

Units: mg/Kg (ppm)
Basis: Dry

Source: N.R.C.C. Dolt-3

Analyte	Prep Method	Analysis Method	True Value	Result	Percent Recovery	Control Limits	Result Notes
Selenium	PSEP Tissue	7740	7.06	5.37	76	5.26-9.05	

Metals

- 10 -

DETECTION LIMITS

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Project Name: NA

ICP/ICP-MS ID #:

GFAA ID #: K-FLAA-02

AA ID #:

Analyte	Wave-length (nm)	Back-ground	MRL ug/L	MDL ug/L	M
Selenium			0.5	0.1	F

Comments:

Metals
-13-
PREPARATION LOG

Client: New Fields Environmental

Service Request: K0710142

Project No.: NA

Project Name: NA

Method: F

Sample ID	Preparation Date	Initial Volume	Final Volume (mL)
K0710142-001	12/12/07	0.3030	30.0
K0710142-002	12/12/07	0.3030	30.0
K0710142-003	12/12/07	0.3130	30.0
K0710142-004	12/12/07	0.3100	30.0
K0710142-005	12/12/07	0.3040	30.0
K0710142-006	12/12/07	0.3180	30.0
K0710142-007	12/12/07	0.3060	30.0
K0710142-007D	12/12/07	0.3070	30.0
K0710142-007S	12/12/07	0.3140	30.0
K0710142-MB	12/12/07	0.3000	30.0
LCS-DOLT	12/12/07	0.30	30.0
LCS-DORM	12/12/07	0.30	30.0
LCSS DORM	12/12/07	0.30	30.0

Columbia Analytical Services
Metals Tissue Digestion Sheet

Service Request Number(s): K0710042, K0710142

Star Lims Run No.: 59944 Analysis for: ICP ICP-MS GFAA
Method: Tissue other: Se-hydride

Sample	Initial Weight (g)	freeze Dry	Wet	Final Volume (ml)	Matrix
AB			X	30mls	15% HNO ₃
Dolt	0.301				
Dorm	0.302				
K0710042-01	0.309				
- 02	0.306				
- 03	0.354				
- 09	0.307				
- 10	0.214				
K0710142-01	0.303	X			
- 02	0.303				
- 03	0.313				
- 04	0.310				
- 05	0.304				
- 06	0.318				
- 07	0.302				
- 07B	0.307				
- 07S	0.314				
<u>AB 12/12/07</u>					

Time Digestion Started: 3:00p.m. 12/12/07 Oven Temp: 110°C

Lot # Acids Used: HNO₃ MSTO-81°C

LCS: Dorm-2, Dolt-3

Time Digestion Ended: 9:00a.m. 12/15/07

Oven Temp: 107°C

Balance I.D.: 21-B

QCP CICV-1, MET1-59-H, _____ mls. added
QCP CICV-2, MET1-59-I, _____ mls. added
QCP CICV-3, MET1-59-J, _____ mls. Added
SS6, MET1-59-K, _____ mls. Added

SPIKE INFO

SS1-MET1-60-R, 0.3 mls added
SS5-MET1-61-P, 0.05 mls added
SS6-MET1-60-O, 0.05 mls added

Additional spikes: _____

Comments: K0710042-1, 2, 8, 9, 10 are wet sample (limited)

Analyst Angela Black Date 12/12/07
Reviewer _____ Date 12/18/07

TissueDig.xls
12/3/2007

00038

Element Analyzed Se Hydride Instrument K-FLAA-2
Service Request # K10642, K10142

Batch QC SR's # _____

Calibration Std. AA1-8-A

Starlims # 02455

Run # 121907-Se

Hydride Data Review Form

	Yes	No	NA
1. ICV within 10% of true Value	<u>X</u>	_____	_____
2. Calibration data included	<u>X</u>	_____	_____
3. CCV's in control	<u>X</u>	_____	_____
4. CCB's and/or ICB's below MRL	<u>X</u>	_____	_____
5. All reported Results within Cal. Range	<u>X</u>	_____	_____
6. All Calculations are Correct	<u>X</u>	_____	_____

Comments

Primary Reviewed by A Date 12/19/07

Secondary Reviewed by JDB Date 12/17/07

00039

COLUMBIA ANALYTICAL SERVICES, INC.

GFAA Run Log

Method: (Circle Method Used) 7742 7062 Other: _____ Element: As Se	Service Request # :
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SAMPLE NUMBER	Dilution Factor	Measured (µg/L)	Recoveries (ICV, CCV, CRA, LCS, Matrix Spk.)	Comments
ICV	-	10.062	101%	
ICB	-	0.063		
CRA	-	0.594	119%	
CCV	-	9.868	99%	
CCB	-	0.061		
K0710642-PB	1/2	0.139		
LCSS DOLT	1/2+1/10	2.600	75%	
LCSS DORM	1/2+1/2	1.905		Rerun
K0710642-001	1/2+1/5	0.703		Rerun
K0710642-002	1/2+1/5	2.762		
K0710642-008	1/2+1/5	4.059		
K0710642-009	1/2+1/5	7.153		
K0710642-010	1/2+1/5	0.369		Rerun
K0710142-001	1/2+1/5	6.476		
K0710142-002	1/2+1/5	11.584		
CCV	-	9.284	93%	
CCB	-	0.085		
K0710142-003	1/2+1/5	8.350		
K0710142-004	1/2+1/5	10.812		
K0710142-005	1/2+1/5	5.356		
K0710142-006	1/2+1/5	6.628		
K0710142-007	1/2+1/5	6.532		
K0710142-007D	1/2+1/5	5.721		
K0710142-007S	1/2+1/20	5.816	100%	
K0710142-007A	1/2+1/5	9.132		Rerun
CCV	-	9.217	92%	
CCB	-	0.010		
LCSS DORM	1/2	2.709	68%	
K0710642-001	1/2	2.150		
K0710642-010	1/2	1.698		Rerun

12/19/07

12/19/07

True Values/QC Limits:	LCSW	Water Spike	LCSS (ERA D045540)	Soil Spike
Arsenic:	2500ppb (80-120%)	1000ppb (80-120%)	146.0mg/kg (80-120%)	1000ppb (80-120%)
Selenium	2500ppb (71-122%)	1000ppb (80-120%)	73.0mg/kg (61-142%)	1000ppb (80-120%)

Analyst <i>Jenna Just</i>	Date: 12/19/07	Page Number: 1
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Sample ID: Std 5.0

Date Collected: 12/19/2007 8:47:51 AM

Analyst:

Data Type: Original

Replicate Data: Std 5.0

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[5.0]	0.083	0.467	0.087			08:48:14	Yes
2		[5.0]	0.082	0.459	0.087			08:48:48	Yes
3		[5.0]	0.086	0.495	0.091			08:49:22	Yes
Mean:		[5.0]	0.084						
SD:		0.0	0.0021						
%RSD:		0.0	2.53						

Standard number 3 applied. [5.0]
Correlation Coef.: 0.998705 Slope: 0.01686 Intercept: 0.00000

Sequence No.: 5

Autosampler Location: 5

Sample ID: Std 10.0

Date Collected: 12/19/2007 8:50:12 AM

Analyst:

Data Type: Original

Replicate Data: Std 10.0

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[10.0]	0.160	0.916	0.165			08:50:35	Yes
2		[10.0]	0.158	0.893	0.163			08:51:10	Yes
3		[10.0]	0.164	0.887	0.168			08:51:45	Yes
Mean:		[10.0]	0.161						
SD:		0.0	0.0028						
%RSD:		0.0	1.75						

Standard number 4 applied. [10.0]
Correlation Coef.: 0.999303 Slope: 0.01623 Intercept: 0.00000

Sequence No.: 6

Autosampler Location: 6

Sample ID: Std 15.0

Date Collected: 12/19/2007 8:52:36 AM

Analyst:

Data Type: Original

Replicate Data: Std 15.0

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1		[15.0]	0.249	1.402	0.253			08:52:59	Yes
2		[15.0]	0.250	1.374	0.255			08:53:33	Yes
3		[15.0]	0.248	1.354	0.253			08:54:08	Yes
Mean:		[15.0]	0.249						
SD:		0.0	0.0012						
%RSD:		0.0	0.48						

Standard number 5 applied. [15.0]
Correlation Coef.: 0.999604 Slope: 0.01647 Intercept: 0.00000
The calibration curve may not be linear.

Calibration data for Se 196.03

Equation: Linear Through Zero

ID	Mean Signal (Abs)	Entered Conc. ug/L	Calculated Conc. ug/L	Standard Deviation	%RSD
Cal Blk	0.0000	0	0.000	0.00	5.7
Std 0.5	0.0095	0.5	0.576	0.00	14.7
Std 1.0	0.0195	1.0	1.187	0.00	2.7
Std 5.0	0.0835	5.0	5.071	0.00	2.5
Std 10.0	0.1606	10.0	9.753	0.00	1.7
Std 15.0	0.2490	15.0	15.118	0.00	0.5

Correlation Coef.: 0.999604 Slope: 0.01647 Intercept: 0.00000

00043

Sequence No.: 7
Sample ID: ICV
Analyst:

Autosampler Location: 7
Date Collected: 12/19/2007 8:55:00 AM
Data Type: Original

Replicate Data: ICV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.12	10.12	0.167	0.938	0.171			08:55:24	Yes
2	10.08	10.08	0.166	0.926	0.171			08:55:59	Yes
3	9.988	9.988	0.165	0.920	0.169			08:56:33	Yes
Mean:	10.06	10.06	0.166						
SD:	0.067	0.067	0.0011						
%RSD:	0.663	0.663	0.66						

QC value within limits for Se 196.03 Recovery = 100.62%
All analyte(s) passed QC.

Sequence No.: 8
Sample ID: ICB
Analyst:

Autosampler Location: 1
Date Collected: 12/19/2007 8:57:25 AM
Data Type: Original

Replicate Data: ICB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.133	0.133	0.002	0.032	0.007			08:57:47	Yes
2	0.037	0.037	0.001	0.019	0.005			08:58:21	Yes
3	0.018	0.018	0.000	0.014	0.005			08:58:55	Yes
Mean:	0.063	0.063	0.001						
SD:	0.062	0.062	0.0010						
%RSD:	98.62	98.62	98.62						

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Sequence No.: 9
Sample ID: CRA
Analyst:

Autosampler Location: 2
Date Collected: 12/19/2007 8:59:44 AM
Data Type: Original

Replicate Data: CRA

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.580	0.580	0.010	0.070	0.014			09:00:05	Yes
2	0.607	0.607	0.010	0.074	0.015			09:00:40	Yes
3	0.594	0.594	0.010	0.071	0.014			09:01:14	Yes
Mean:	0.594	0.594	0.010						
SD:	0.013	0.013	0.0002						
%RSD:	2.267	2.267	2.27						

QC value within limits for Se 196.03 Recovery = 118.78%
All analyte(s) passed QC.

Sequence No.: 10
Sample ID: CCV
Analyst:

Autosampler Location: 5
Date Collected: 12/19/2007 9:02:04 AM
Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.510	9.510	0.157	0.907	0.161			09:02:27	Yes
2	9.479	9.479	0.156	0.877	0.161			09:03:01	Yes
3	10.61	10.61	0.175	0.903	0.180			09:03:35	Yes
Mean:	9.868	9.868	0.163						

SD: 0.647 0.647 0.0107

%RSD: 6.558 6.558 6.56

QC value within limits for Se 196.03 Recovery = 98.68%

All analyte(s) passed QC.

Sequence No.: 11

Autosampler Location: 1

Sample ID: CCB

Date Collected: 12/19/2007 9:04:27 AM

Analyst:

Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.115	0.115	0.002	0.030	0.007			09:04:48	Yes
2	0.044	0.044	0.001	0.027	0.005			09:05:24	Yes
3	0.025	0.025	0.000	0.013	0.005			09:05:58	Yes
Mean:	0.061	0.061	0.001						
SD:	0.047	0.047	0.0008						
%RSD:	77.23	77.23	77.23						

QC value within limits for Se 196.03 Recovery = Not calculated

All analyte(s) passed QC.

Sequence No.: 12

Autosampler Location: 9

Sample ID: K0710642-PB

Date Collected: 12/19/2007 9:06:47 AM

Analyst:

Data Type: Original

Replicate Data: K0710642-PB

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.157	0.157	0.003	0.030	0.007			09:07:08	Yes
2	0.116	0.116	0.002	0.024	0.007			09:07:42	Yes
3	0.145	0.145	0.002	0.023	0.007			09:08:16	Yes
Mean:	0.139	0.139	0.002						
SD:	0.021	0.021	0.0003						
%RSD:	15.14	15.14	15.14						

Sequence No.: 13

Autosampler Location: 10

Sample ID: LCSS DOLT

Date Collected: 12/19/2007 9:09:05 AM

Analyst:

Data Type: Original

Replicate Data: LCSS DOLT

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.519	2.519	0.041	0.276	0.046			09:09:26	Yes
2	2.680	2.680	0.044	0.282	0.049			09:10:00	Yes
3	2.600	2.600	0.043	0.259	0.047			09:10:34	Yes
Mean:	2.600	2.600	0.043						
SD:	0.080	0.080	0.0013						
%RSD:	3.090	3.090	3.09						

Sequence No.: 14

Autosampler Location: 11

Sample ID: LCSS DORM

Date Collected: 12/19/2007 9:11:24 AM

Analyst:

Data Type: Original

Replicate Data: LCSS DORM

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	1.970	1.970	0.032	0.215	0.037			09:11:46	Yes
2	1.867	1.867	0.031	0.203	0.035			09:12:20	Yes
3	1.878	1.878	0.031	0.200	0.036			09:12:53	Yes

Mean: 1.905 1.905 0.031
SD: 0.057 0.057 0.0009
%RSD: 2.970 2.970 2.97

Sequence No.: 15 Autosampler Location: 12
Sample ID: K0710642-001 Date Collected: 12/19/2007 9:13:44 AM
Analyst: Data Type: Original

Replicate Data: K0710642-001

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 16 Autosampler Location: 13
Sample ID: K0710642-002 Date Collected: 12/19/2007 9:16:04 AM
Analyst: Data Type: Original

Replicate Data: K0710642-002

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 17 Autosampler Location: 14
Sample ID: K0710642-008 Date Collected: 12/19/2007 9:18:26 AM
Analyst: Data Type: Original

Replicate Data: K0710642-008

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 18 Autosampler Location: 15
Sample ID: K0710642-009 Date Collected: 12/19/2007 9:20:51 AM
Analyst: Data Type: Original

Replicate Data: K0710642-009

Table with 10 columns: Repl #, SampleConc ug/L, StndConc ug/L, BlnkCorr Signal, Peak Area, Peak Height, Bkgnd Area, Bkgnd Height, Time, Peak Stored. Contains 3 replicate rows and summary statistics.

Sequence No.: 19

Sample ID: K0710642-010

Analyst:

Autosampler Location: 16

Date Collected: 12/19/2007 9:23:16 AM

Data Type: Original

Replicate Data: K0710642-010

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.359	0.359	0.006	0.054	0.011			09:23:40	Yes
2	0.372	0.372	0.006	0.052	0.011			09:24:15	Yes
3	0.377	0.377	0.006	0.045	0.011			09:24:49	Yes
Mean:	0.369	0.369	0.006						
SD:	0.009	0.009	0.0002						
%RSD:	2.529	2.529	2.53						

Sequence No.: 20

Sample ID: K0710142-001

Analyst:

Autosampler Location: 17

Date Collected: 12/19/2007 9:25:41 AM

Data Type: Original

Replicate Data: K0710142-001

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	6.402	6.402	0.105	0.638	0.110			09:26:01	Yes
2	6.519	6.519	0.107	0.641	0.112			09:26:35	Yes
3	6.506	6.506	0.107	0.648	0.112			09:27:10	Yes
Mean:	6.476	6.476	0.107						
SD:	0.064	0.064	0.0011						
%RSD:	0.988	0.988	0.99						

Sequence No.: 21

Sample ID: K0710142-002

Analyst:

Autosampler Location: 18

Date Collected: 12/19/2007 9:28:00 AM

Data Type: Original

Replicate Data: K0710142-002

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	11.62	11.62	0.191	1.174	0.196			09:28:24	Yes
2	11.70	11.70	0.193	1.151	0.197			09:28:58	Yes
3	11.43	11.43	0.188	1.143	0.193			09:29:32	Yes
Mean:	11.58	11.58	0.191						
SD:	0.140	0.140	0.0023						
%RSD:	1.211	1.211	1.21						

Sequence No.: 22

Sample ID: CCV

Analyst:

Autosampler Location: 5

Date Collected: 12/19/2007 9:30:22 AM

Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	BlkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.066	9.066	0.149	0.836	0.154			09:30:45	Yes
2	9.412	9.412	0.155	0.874	0.160			09:31:18	Yes
3	9.375	9.375	0.154	0.861	0.159			09:31:53	Yes
Mean:	9.284	9.284	0.153						
SD:	0.190	0.190	0.0031						
%RSD:	2.045	2.045	2.05						

QC value within limits for Se 196.03 Recovery = 92.84%
All analyte(s) passed QC.

00047

Sequence No.: 23
Sample ID: CCB
Analyst:

Autosampler Location: 1
Date Collected: 12/19/2007 9:32:45 AM
Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.069	0.069	0.001	0.035	0.006			09:33:06	Yes
2	0.073	0.073	0.001	0.013	0.006			09:33:40	Yes
3	0.113	0.113	0.002	0.031	0.007			09:34:15	Yes
Mean:	0.085	0.085	0.001						
SD:	0.024	0.024	0.0004						
%RSD:	28.52	28.52	28.52						

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Sequence No.: 24
Sample ID: K0710142-003
Analyst:

Autosampler Location: 19
Date Collected: 12/19/2007 9:35:04 AM
Data Type: Original

Replicate Data: K0710142-003

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	8.314	8.314	0.137	0.855	0.142			09:35:25	Yes
2	8.395	8.395	0.138	0.830	0.143			09:35:59	Yes
3	8.342	8.342	0.137	0.820	0.142			09:36:33	Yes
Mean:	8.350	8.350	0.138						
SD:	0.041	0.041	0.0007						
%RSD:	0.489	0.489	0.49						

Sequence No.: 25
Sample ID: K0710142-004
Analyst:

Autosampler Location: 20
Date Collected: 12/19/2007 9:37:23 AM
Data Type: Original

Replicate Data: K0710142-004

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.95	10.95	0.180	1.098	0.185			09:37:46	Yes
2	10.55	10.55	0.174	1.063	0.178			09:38:20	Yes
3	10.93	10.93	0.180	1.095	0.185			09:38:54	Yes
Mean:	10.81	10.81	0.178						
SD:	0.228	0.228	0.0038						
%RSD:	2.106	2.106	2.11						

Sequence No.: 26
Sample ID: K0710142-005
Analyst:

Autosampler Location: 21
Date Collected: 12/19/2007 9:39:44 AM
Data Type: Original

Replicate Data: K0710142-005

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	5.234	5.234	0.086	0.546	0.091			09:40:05	Yes
2	5.424	5.424	0.089	0.525	0.094			09:40:39	Yes
3	5.410	5.410	0.089	0.528	0.094			09:41:13	Yes
Mean:	5.356	5.356	0.088						
SD:	0.106	0.106	0.0017						
%RSD:	1.976	1.976	1.98						

Sequence No.: 27
Sample ID: K0710142-006
Analyst:

Autosampler Location: 22
Date Collected: 12/19/2007 9:42:03 AM
Data Type: Original

Replicate Data: K0710142-006

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	6.790	6.790	0.112	0.662	0.117			09:42:25	Yes
2	6.537	6.537	0.108	0.636	0.112			09:42:59	Yes
3	6.557	6.557	0.108	0.644	0.113			09:43:33	Yes
Mean:	6.628	6.628	0.109						
SD:	0.141	0.141	0.0023						
%RSD:	2.120	2.120	2.12						

Sequence No.: 28
Sample ID: K0710142-007
Analyst:

Autosampler Location: 23
Date Collected: 12/19/2007 9:44:23 AM
Data Type: Original

Replicate Data: K0710142-007

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	6.581	6.581	0.108	0.660	0.113			09:44:45	Yes
2	6.488	6.488	0.107	0.659	0.112			09:45:19	Yes
3	6.528	6.528	0.108	0.643	0.112			09:45:53	Yes
Mean:	6.532	6.532	0.108						
SD:	0.047	0.047	0.0008						
%RSD:	0.713	0.713	0.71						

Sequence No.: 29
Sample ID: K0710142-007D
Analyst:

Autosampler Location: 24
Date Collected: 12/19/2007 9:46:43 AM
Data Type: Original

Replicate Data: K0710142-007D

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	5.772	5.772	0.095	0.601	0.100			09:47:06	Yes
2	5.617	5.617	0.093	0.586	0.097			09:47:40	Yes
3	5.773	5.773	0.095	0.582	0.100			09:48:14	Yes
Mean:	5.721	5.721	0.094						
SD:	0.090	0.090	0.0015						
%RSD:	1.568	1.568	1.57						

Sequence No.: 30
Sample ID: K0710142-007S
Analyst:

Autosampler Location: 25
Date Collected: 12/19/2007 9:49:04 AM
Data Type: Original

Replicate Data: K0710142-007S

Repl #	SampleConc ug/L	StndConc ug/L	BlnkCorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	5.676	5.676	0.093	0.520	0.098			09:49:27	Yes
2	5.791	5.791	0.095	0.519	0.100			09:50:02	Yes
3	5.980	5.980	0.098	0.549	0.103			09:50:37	Yes
Mean:	5.816	5.816	0.096						
SD:	0.154	0.154	0.0025						
%RSD:	2.642	2.642	2.64						

Sequence No.: 31
Sample ID: K0710142-007A
Analyst:

Autosampler Location: 26
Date Collected: 12/19/2007 9:51:28 AM
Data Type: Original

00049

12/19/07

Replicate Data: K0710142-007A

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	11.13	11.13	0.183	1.339	0.188			09:51:51	Yes
2	7.451	7.451	0.123	0.522	0.127			09:52:25	Yes
3	8.818	8.818	0.145	0.877	0.150			09:52:59	Yes
Mean:	9.132	9.132	0.150						
SD:	1.858	1.858	0.0306						
%RSD:	20.35	20.35	20.35						

Sequence No.: 32
Sample ID: CCV
Analyst:

Autosampler Location: 5
Date Collected: 12/19/2007 9:53:50 AM
Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	9.407	9.407	0.155	0.862	0.160			09:54:13	Yes
2	8.735	8.735	0.144	0.757	0.149			09:54:48	Yes
3	9.509	9.509	0.157	0.868	0.161			09:55:22	Yes
Mean:	9.217	9.217	0.152						
SD:	0.421	0.421	0.0069						
%RSD:	4.562	4.562	4.56						

QC value within limits for Se 196.03 Recovery = 92.17%
All analyte(s) passed QC.

Sequence No.: 33
Sample ID: CCB
Analyst:

Autosampler Location: 1
Date Collected: 12/19/2007 9:56:13 AM
Data Type: Original

Replicate Data: CCB

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	0.107	0.107	0.002	0.030	0.006			09:56:34	Yes
2	-0.035	-0.035	-0.001	0.007	0.004			09:57:08	Yes
3	-0.043	-0.043	-0.001	0.011	0.004			09:57:43	Yes
Mean:	0.010	0.010	0.000						
SD:	0.085	0.085	0.0014						
%RSD:	859.1	859.1	859.05						

QC value within limits for Se 196.03 Recovery = Not calculated
All analyte(s) passed QC.

Analysis Begun

Logged In Analyst: ACQMET10
Spectrometer Model: AAnalyst 200, S/N 200S5061701
Technique: AA FIAS-Flame
Autosampler Model: AS-90

Sample Information File: C:\data-AA\ACQMET10\Sample Information\121907-Se.sif
Batch ID: 121907-Se
Results Data Set: 121907-Se
Results Library: R:\ICP\WIP\DATA\K-FLAA-02\Results.mdb

Sequence No.: 34
Sample ID: LCSS DORM
Analyst:

Autosampler Location:
Date Collected: 12/19/2007 9:59:33 AM
Data Type: Original

User canceled analysis.

Sequence No.: 35

Autosampler Location:

Sample ID: LCSS DORM
Analyst:

Date Collected: 12/19/2007 9:59:47 AM
Data Type: Original

Replicate Data: LCSS DORM

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.775	2.775	0.046	0.277	0.050			10:00:03	Yes
2	2.688	2.688	0.044	0.281	0.049			10:00:37	Yes
3	2.664	2.664	0.044	0.277	0.049			10:01:11	Yes
Mean:	2.709	2.709	0.045						
SD:	0.059	0.059	0.0010						
%RSD:	2.163	2.163	2.16						

=====
Sequence No.: 36
Sample ID: K0710642-001
Analyst:

Autosampler Location:
Date Collected: 12/19/2007 10:02:22 AM
Data Type: Original

Replicate Data: K0710642-001

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.206	2.206	0.036	0.231	0.041			10:02:38	Yes
2	2.110	2.110	0.035	0.221	0.039			10:03:13	Yes
3	2.135	2.135	0.035	0.231	0.040			10:03:47	Yes
Mean:	2.150	2.150	0.035						
SD:	0.050	0.050	0.0008						
%RSD:	2.326	2.326	2.33						

Sequence No.: 37
Sample ID: K0710642-010
Analyst:

Autosampler Location:
Date Collected: 12/19/2007 10:05:17 AM
Data Type: Original

Replicate Data: K0710642-010

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	2.351	2.351	0.039	0.323	0.043			10:05:34	Yes
2	1.353	1.353	0.022	0.127	0.027			10:06:08	Yes
3	1.388	1.388	0.023	0.139	0.028			10:06:42	Yes
Mean:	1.698	1.698	0.028						
SD:	0.566	0.566	0.0093						
%RSD:	33.37	33.37	33.37						

Sequence No.: 38
Sample ID: K0710642-010
Analyst:

Autosampler Location:
Date Collected: 12/19/2007 10:07:38 AM
Data Type: Original

Replicate Data: K0710642-010

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	1.269	1.269	0.021	0.116	0.026			10:07:56	Yes
2	1.304	1.304	0.021	0.136	0.026			10:08:30	Yes
3	1.250	1.250	0.021	0.123	0.025			10:09:05	Yes
Mean:	1.274	1.274	0.021						
SD:	0.027	0.027	0.0005						
%RSD:	2.150	2.150	2.15						

Sequence No.: 39
Sample ID: K0710642-001A
Analyst:

Autosampler Location:
Date Collected: 12/19/2007 10:10:21 AM
Data Type: Original

Replicate Data: K0710642-001A

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.831	3.831	0.063	0.399	0.068			10:10:38	Yes
2	3.822	3.822	0.063	0.387	0.068			10:11:12	Yes
3	3.782	3.782	0.062	0.381	0.067			10:11:46	Yes
Mean:	3.812	3.812	0.063						
SD:	0.026	0.026	0.0004						
%RSD:	0.681	0.681	0.68						

12/19/07

Sequence No.: 40

Sample ID: K0710642-010A

Analyst:

Autosampler Location:

Date Collected: 12/19/2007 10:24:28 AM

Data Type: Original

Replicate Data: K0710642-010A

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	3.494	3.494	0.058	0.363	0.062			10:24:44	Yes
2	3.573	3.573	0.059	0.366	0.064			10:25:18	Yes
3	3.466	3.466	0.057	0.369	0.062			10:25:53	Yes
Mean:	3.511	3.511	0.058						
SD:	0.055	0.055	0.0009						
%RSD:	1.577	1.577	1.58						

Sequence No.: 41

Sample ID: K0710642-0002A

Analyst:

Autosampler Location:

Date Collected: 12/19/2007 10:32:19 AM

Data Type: Original

Replicate Data: K0710642-0002A

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	6.813	6.813	0.112	0.630	0.117			10:32:35	Yes
2	8.013	8.013	0.132	0.797	0.137			10:33:09	Yes
3	7.322	7.322	0.121	0.666	0.125			10:33:43	Yes
Mean:	7.383	7.383	0.122						
SD:	0.602	0.602	0.0099						
%RSD:	8.160	8.160	8.16						

Sequence No.: 42

Sample ID: CCV

Analyst:

Autosampler Location:

Date Collected: 12/19/2007 10:34:45 AM

Data Type: Original

Replicate Data: CCV

Repl #	SampleConc ug/L	StndConc ug/L	Blncorr Signal	Peak Area	Peak Height	Bkgnd Area	Bkgnd Height	Time	Peak Stored
1	10.21	10.21	0.168	0.904	0.173			10:35:01	Yes
2	9.796	9.796	0.161	0.907	0.166			10:35:36	Yes
3	10.01	10.01	0.165	0.928	0.170			10:36:10	Yes
Mean:	10.01	10.01	0.165						
SD:	0.206	0.206	0.0034						
%RSD:	2.057	2.057	2.06						

Sequence No.: 43

Sample ID: CCB

Analyst:

Autosampler Location:

Date Collected: 12/19/2007 10:37:05 AM

Data Type: Original

Replicate Data: CCB

Repl #	SampleConc	StndConc	Blncorr	Peak	Peak	Bkgnd	Bkgnd	Time	Peak
	ug/L	ug/L	Signal	Area	Height	Area	Height		Stored

#	ug/L	ug/L	Signal	Area	Height	Area	Height	Stored
1	0.070	0.070	0.001	0.018	0.006		10:37:22	Yes
2	0.122	0.122	0.002	0.029	0.007		10:37:56	Yes
3	0.508	0.508	0.008	0.072	0.013		10:38:30	Yes
Mean:	0.233	0.233	0.004					
SD:	0.239	0.239	0.0039					
%RSD:	102.6	102.6	102.63					

C0053

Columbia Analytical Services

- Cover Page -

INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental

Service Request: K0710142

Project Name:

Project No.:

Sample Name:

Lab Code:

SM1007-CC1A-FT001

K0710142-001

SM1007-CC1A-FT001-Eggs

K0710142-002

SM1007-CC3A-FT101

K0710142-003

SM1007-CC3A-FT101-Eggs

K0710142-004

SM1007-CC3A-FT001

K0710142-005

SM1007-CC3A-FT100

K0710142-006

SM1007-CC3A-FT002

K0710142-007

SM1007-CC3A-FT002D

K0710142-007D

SM1007-CC3A-FT002S

K0710142-007S

Method Blank

K0710142-MB

Comments:

Approved By: _____

Date: _____

Metals

- 1 -

INORGANIC ANALYSIS DATA PACKAGE

Client: New Fields Environmental Service Request: K0710142
Project No.: NA Date Collected: 10/22/07
Project Name: NA Date Received: 10/30/07
Matrix: TISSUE Units: mg/Kg
Basis: DRY

Sample Name: SM1007-CC1A-FT001-Eggs Lab Code: K0710142-002

Analyte	Analysis Method	MRL	MDL	Dilution Factor	Date Extracted	Date Analyzed	Result	C	Q
Selenium	7742	0.50	0.10	10.0	12/12/07	12/19/07	11.5		

Comments:

COLUMBIA ANALYTICAL SERVICES, INC.

Analytical Report

Client: New Fields Environmental
Project: NA
Sample Matrix: Tissue

Service Request: K0710142
Date Collected: 10/22-24/07
Date Received: 10/30/07

Solids, Total

Prep Method: NONE
Analysis Method: Freeze Dry
Test Notes:

Units: PERCENT
Basis: Wet

Sample Name	Lab Code	Date Analyzed	Result	Result Notes
SM1007-CC1A-FT001	K0710142-001	12/05/07	25.8	
SM1007-CC1A-FT001-Eggs	K0710142-002	12/05/07	35.3	
SM1007-CC3A-FT101	K0710142-003	12/05/07	25.2	
SM1007-CC3A-FT101-Eggs	K0710142-004	12/05/07	36.1	
SM1007-CC3A-FT001	K0710142-005	12/05/07	26.7	
SM1007-CC3A-FT100	K0710142-006	12/05/07	26.5	
SM1007-CC3A-FT002	K0710142-007	12/05/07	26.8	

APPENDIX E
EPA's TRAP Software and Summary Output Statistics for Each Model Run

Toxicity Relationship Analysis Program (TRAP) version 1.2 (Erickson 2008) – Data Inputs and Outputs

Logistic regression analysis was performed using USEPA regression-analysis software (TRAP version 1.2; Erickson 2008). This program analyzes the decline of a biological variable (e.g., survival, growth, fecundity) from its control value to zero as chemical exposure (e.g., concentration, log concentration, dose) increases. The software allows for two types of analysis options including Maximum Likelihood Tolerance Distribution Analysis and Least-Squares Nonlinear Regression Analysis.

The logistic regression model in the Least-Squares Nonlinear Regression Analysis option was selected for use consistent with USEPA's (2004) approach. The independent variable (exposure variable) for each of these analyses was egg selenium (mg/kg dw). Separate analyses of the egg selenium data found that it was log normally distributed; therefore all analyses using these data in the logistic regression function were conducted using log transformed egg selenium data. Furthermore, Log transformation of concentrations is often desirable to make the analysis models (which are symmetric) more applicable to toxicological toxicity relationships (which are often skewed) (Erickson 2008). The data were entered as raw values and the program provided the log transformation. Independent variables (effects variables) included:

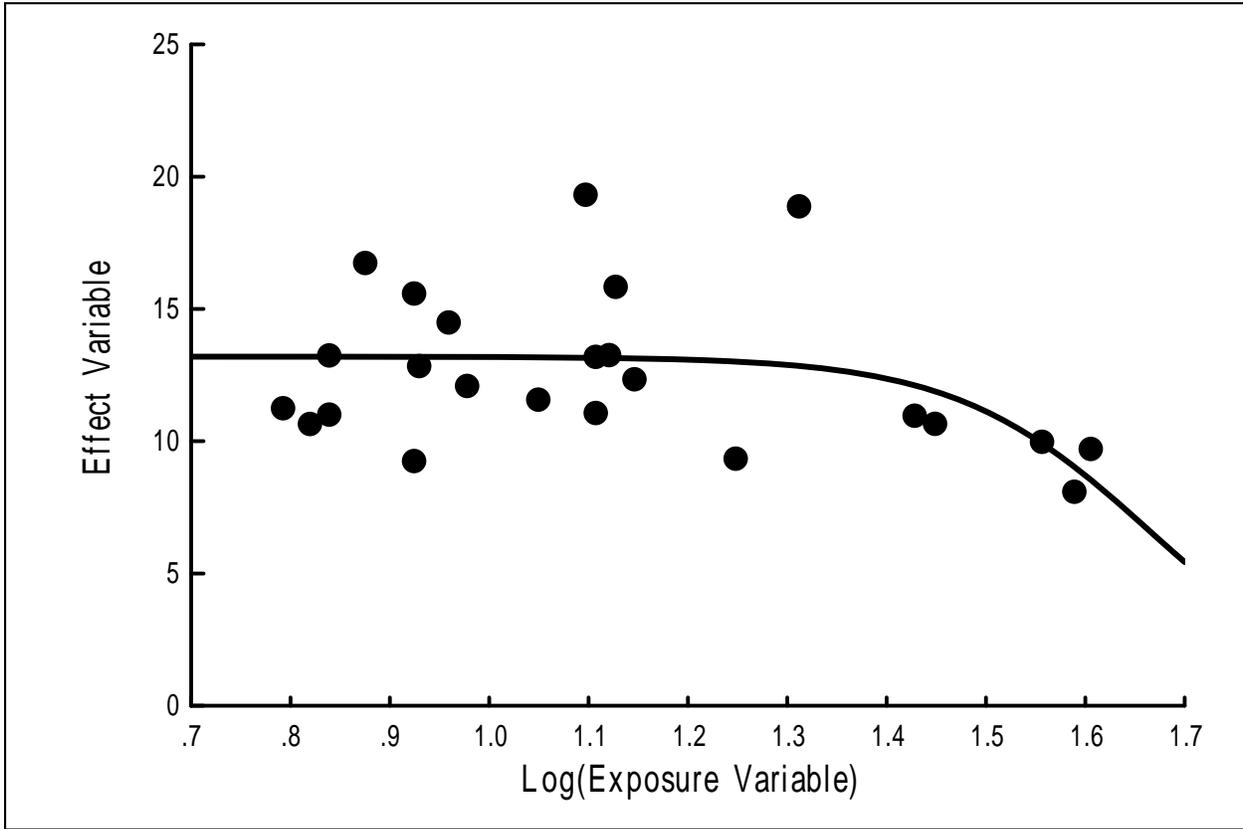
- Larval fish growth
- % Survival 15 day post swim up feeding trial
- % Survival Overall
- % Survival (hatch to test end)
- Cranio facial deformities (fraction normal)
- Skeletal deformities (fraction normal)
- Finfold deformities (fraction normal)
- Edema deformities (fraction normal)
- Sum Fraction normal
- Mean Fraction normal

Data for each of these variables were not transformed.

The model allows for initial guesses of the model parameters X_{50} (effect concentration that causes a 50% reduction relative to the control), Y_0 (control value), and S $(p-50)/(X_p-X_{50})$ which is the steepness. If no initial guessed are made, the model will estimate these parameters. No initial guesses were made and the model was allowed to fill in these parameters.

Independent model runs were made for each pairing of egg selenium data and the dependent variable to obtain a model output which is shown in the remainder of this appendix. The model output illustrates the graphical distribution of the data and the predicted logistic regression line fit to the data. It also includes a parameter summary, predicted effects concentrations and their confidence limits, a regression analysis of variance, a summary of the data, and an error summary. The data summary for each output includes the exposure variable (log transformed egg selenium (mg/kg dw), and the observed effects (raw untransformed input data), predicted effects, and residual error.

Egg Se vs. Growth



Parameter Summary (Logistic Equation Regression Analysis)

Parameter	Guess	FinalEst	StdError	95%LCL	95%UCL
LogX50	1.6132	1.6649	0.1119	1.4321	1.8977
S	2.530	2.546	2.897	-3.478	8.570
Y0	13.454	13.198	0.705	11.731	14.664

Effect Concentration Summary

%Effect	Xp Est	95%LCL	95%UCL
50.0	46.23	27.05	79.01
20.0	33.79	22.84	49.99
10.0	28.13	13.09	60.44
5.0	23.76	7.54	74.80

Egg Se vs. Growth

Regression Analysis of Variance

Source	df	SS	MS	F	Alpha
Total	23	201.5	8.76		
Regression	2	42.0	20.98	2.76	0.0861
Error	21	159.5	7.60		

Data Summary

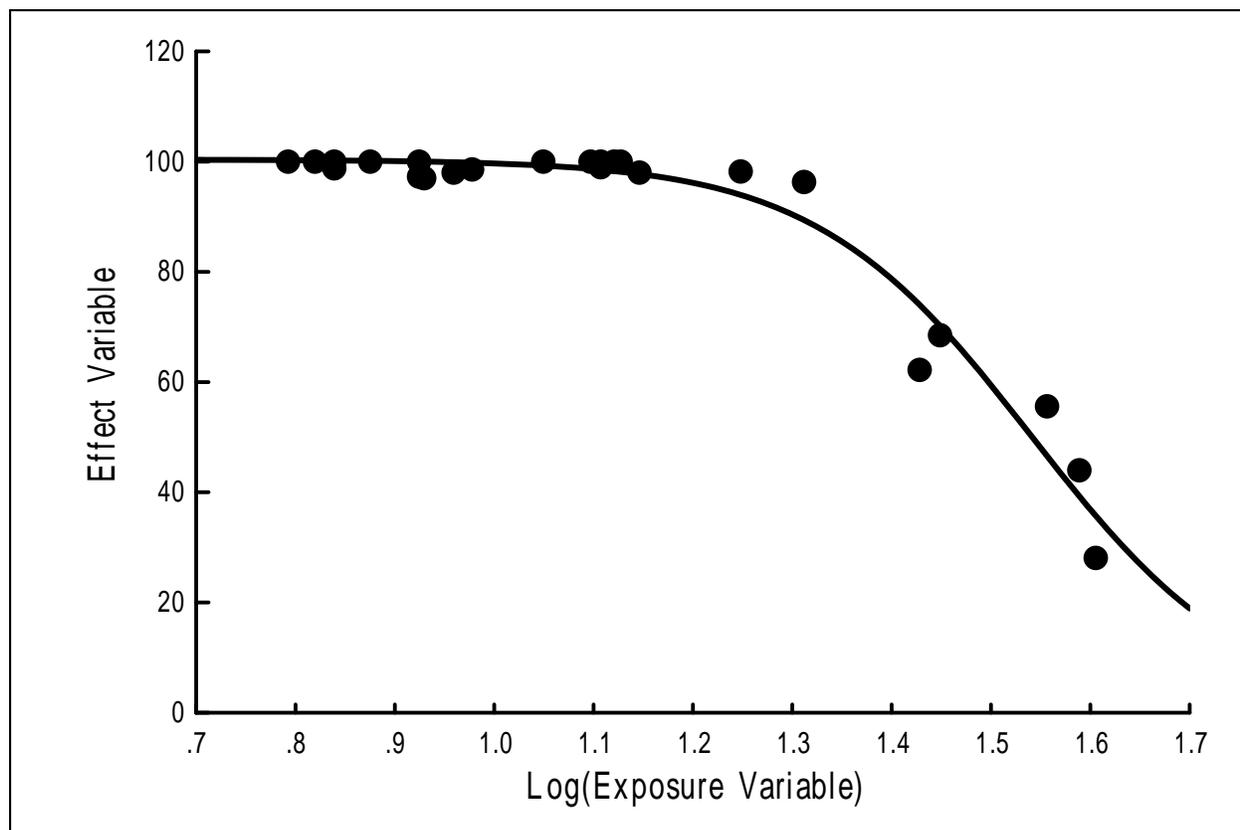
Exposure	Obs Effects	Pred Effects	Residual	Weight
0.7924	11.2480	13.1957	1.9477	1.
0.8195	10.6530	13.1951	2.5421	1.
0.8388	13.2450	13.1946	-0.0504	1.
0.8388	11.0090	13.1946	2.1856	1.
0.8751	16.7360	13.1933	-3.5427	1.
0.9243	15.5830	13.1905	-2.3925	1.
0.9243	9.2485	13.1905	3.9420	1.
0.9294	12.8400	13.1902	0.3502	1.
0.9590	14.4860	13.1876	-1.2984	1.
0.9777	12.0890	13.1855	1.0965	1.
1.0492	11.5730	13.1726	1.5996	1.
1.0969	19.3190	13.1571	-6.1619	1.
1.1072	13.1880	13.1526	-0.0354	1.
1.1072	11.0690	13.1526	2.0836	1.
1.1206	13.2560	13.1461	-0.1099	1.
1.1271	15.8350	13.1426	-2.6924	1.
1.1461	12.3420	13.1309	0.7889	1.
1.2480	9.3420	13.0112	3.6692	1.
1.3118	18.8780	12.8454	-6.0326	1.
1.4281	10.9670	12.1112	1.1442	1.
1.4487	10.6560	11.8832	1.2272	1.
1.5563	9.9790	9.9163	-0.0627	1.
1.5888	8.0918	9.0341	0.9423	1.
1.6053	9.7100	8.5418	-1.1682	1.

Egg Se vs. Growth

Error Summary

Large Standard Error for Steepness

Egg Se vs. Survival (15 day post feeding trial)



Parameter Summary (Logistic Equation Regression Analysis)

Parameter	Guess	FinalEst	StdError	95%LCL	95%UCL
LogX50	1.5396	1.5407	0.0099	1.5200	1.5613
S	1.8091	2.292	0.264	1.742	2.841
Y0	99.16	100.38	1.26	97.76	103.01

Effect Concentration Summary

%Effect	Xp Est	95%LCL	95%UCL
50.0	34.73	33.11	36.42
20.0	24.51	22.26	26.99
10.0	19.997	17.369	23.022
5.0	16.575	13.790	19.921

Egg Se vs. Survival (15 day post feeding trial)

Regression Analysis of Variance

Source	df	SS	MS	F	Alpha
Total	23	9918.	431.23		
Regression	2	9514.	4757.22	247.	0.0000
Error	21	404.	19.23		

Data Summary

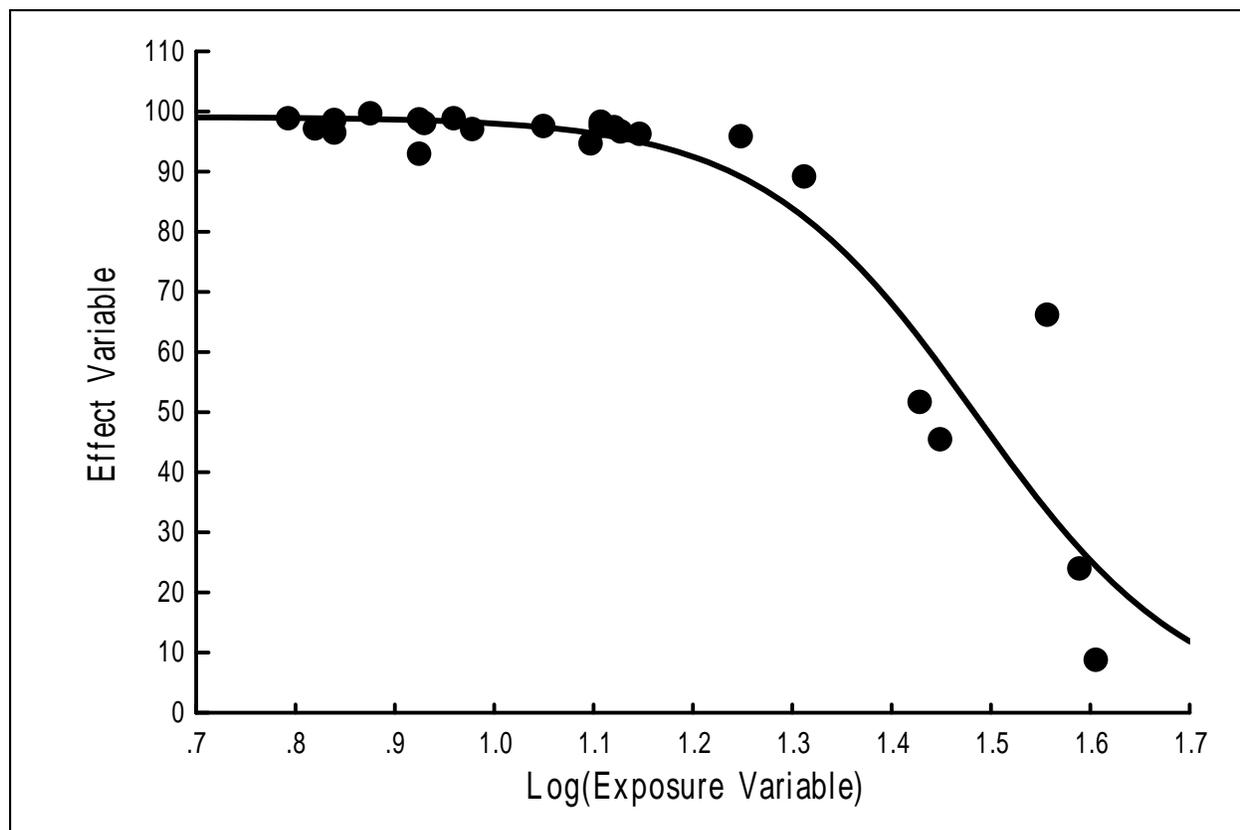
Exposure	Obs Effects	Pred Effects	Residual	Weight
0.7924	100.0000	100.2759	0.2759	1.
0.8195	100.0000	100.2462	0.2462	1.
0.8388	100.0000	100.2201	0.2201	1.
0.8388	98.8000	100.2201	1.4201	1.
0.8751	100.0000	100.1568	0.1568	1.
0.9243	100.0000	100.0294	0.0294	1.
0.9243	97.3000	100.0294	2.7294	1.
0.9294	97.0000	100.0125	3.0125	1.
0.9590	98.0000	99.8980	1.8980	1.
0.9777	98.6000	99.8082	1.2082	1.
1.0492	100.0000	99.2835	-0.7165	1.
1.0969	100.0000	98.6918	-1.3082	1.
1.1072	100.0000	98.5276	-1.4724	1.
1.1072	99.0000	98.5276	-0.4724	1.
1.1206	100.0000	98.2911	-1.7089	1.
1.1271	100.0000	98.1650	-1.8350	1.
1.1461	98.0000	97.7538	-0.2462	1.
1.2480	98.2000	93.9580	-4.2420	1.
1.3118	96.3000	89.4133	-6.8867	1.
1.4281	62.2000	74.0020	11.8020	1.
1.4487	68.5000	70.1752	1.6752	1.
1.5563	55.6000	46.6005	-8.9995	1.
1.5888	44.0000	39.2882	-4.7118	1.
1.6053	28.1000	35.7425	7.6425	1.

Egg Se vs. Survival (15 day post feeding trial)

Error Summary

No Errors

Egg Se vs. Survival (hatch to test end)



Parameter Summary (Logistic Equation Regression Analysis)

Parameter	Guess	FinalEst	StdError	95%LCL	95%UCL
LogX50	1.4841	1.4846	0.0212	1.4406	1.5286
S	2.072	2.316	0.474	1.330	3.301
Y0	97.32	99.10	2.75	93.39	104.82

Effect Concentration Summary

%Effect	Xp Est	95%LCL	95%UCL
50.0	30.52	27.58	33.77
20.0	21.62	17.77	26.32
10.0	17.677	13.440	23.249
5.0	14.681	10.357	20.810

Egg Se vs. Survival (hatch to test end)

Regression Analysis of Variance

Source	df	SS	MS	F	Alpha
Total	23	15331.	666.6		
Regression	2	13611.	6805.3	83.1	0.0000
Error	21	1721.	81.9		

Data Summary

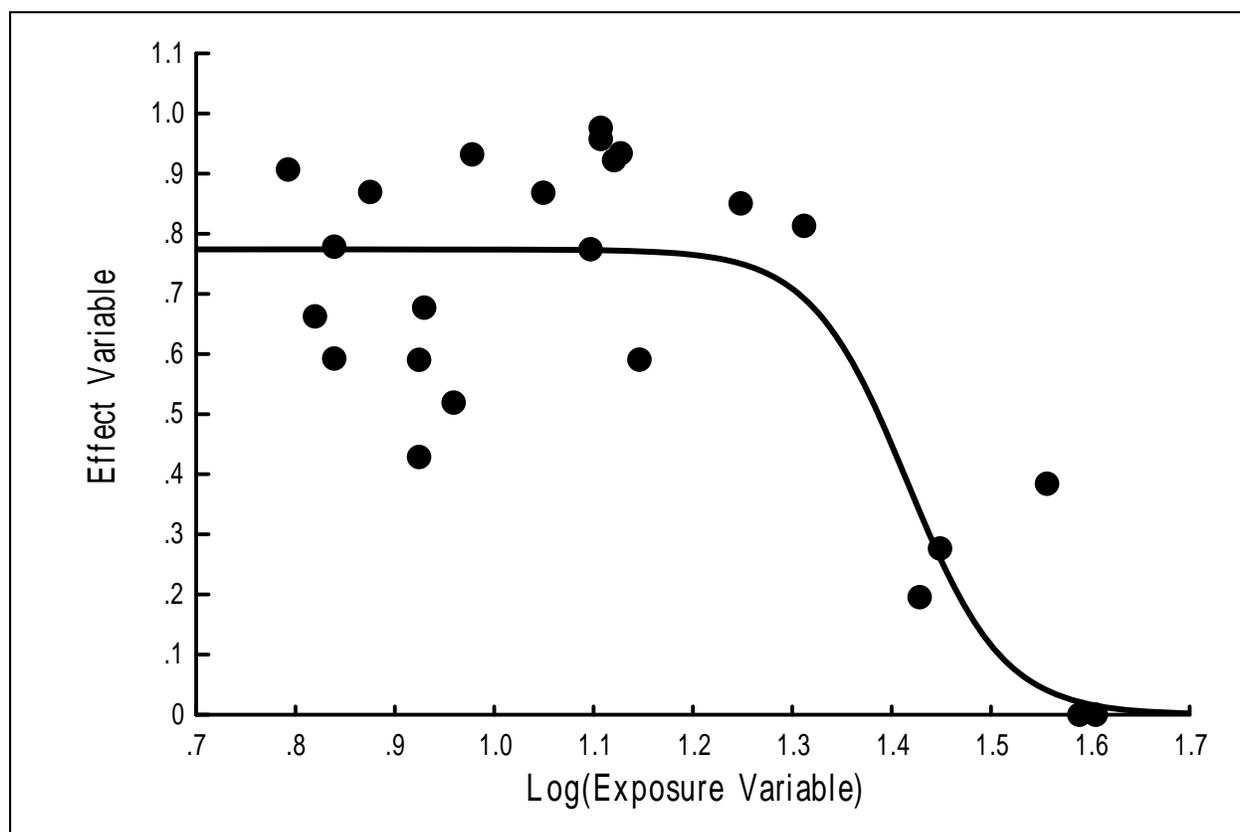
Exposure	Obs Effects	Pred Effects	Residual	Weight
0.7924	98.9000	98.9424	0.0424	1.
0.8195	97.2000	98.8961	1.6961	1.
0.8388	98.6000	98.8553	0.2553	1.
0.8388	96.5000	98.8553	2.3553	1.
0.8751	99.7000	98.7562	-0.9438	1.
0.9243	98.7000	98.5559	-0.1441	1.
0.9243	93.0000	98.5559	5.5559	1.
0.9294	98.1000	98.5293	0.4293	1.
0.9590	98.9000	98.3490	-0.5510	1.
0.9777	97.1000	98.2075	1.1075	1.
1.0492	97.6000	97.3794	-0.2206	1.
1.0969	94.7000	96.4466	1.7466	1.
1.1072	98.3000	96.1883	-2.1117	1.
1.1072	97.6000	96.1883	-1.4117	1.
1.1206	97.4000	95.8167	-1.5833	1.
1.1271	96.7000	95.6189	-1.0811	1.
1.1461	96.3000	94.9751	-1.3249	1.
1.2480	95.9000	89.1474	-6.7526	1.
1.3118	89.2000	82.4730	-6.7270	1.
1.4281	51.7000	62.2247	10.5247	1.
1.4487	45.5000	57.7159	12.2159	1.
1.5563	66.2000	33.6760	-32.5240	1.
1.5888	24.0000	27.3308	3.3308	1.
1.6053	8.8000	24.4157	15.6157	1.

Egg Se vs. Survival (hatch to test end)

Error Summary

No Errors

Egg Se vs. Cranio Facial Fraction Normal



Parameter Summary (Logistic Equation Regression Analysis)

Parameter	Guess	FinalEst	StdError	95%LCL	95%UCL
LogX50	1.4405	1.4157	0.0347	1.3436	1.4878
S	2.739	5.153	3.811	-2.773	13.079
Y0	0.7802	0.7740	0.0430	0.6845	0.8634

Effect Concentration Summary

%Effect	Xp Est	95%LCL	95%UCL
50.0	26.04	22.06	30.75
20.0	22.31	15.91	31.27
10.0	20.37	12.78	32.47
5.0	18.742	10.402	33.769

Egg Se vs. Cranio Facial Fraction Normal

Regression Analysis of Variance

Source	df	SS	MS	F	Alpha
Total	23	2.035	0.0885		
Regression	2	1.381	0.6903	22.2	0.0000
Error	21	0.654	0.0312		

Data Summary

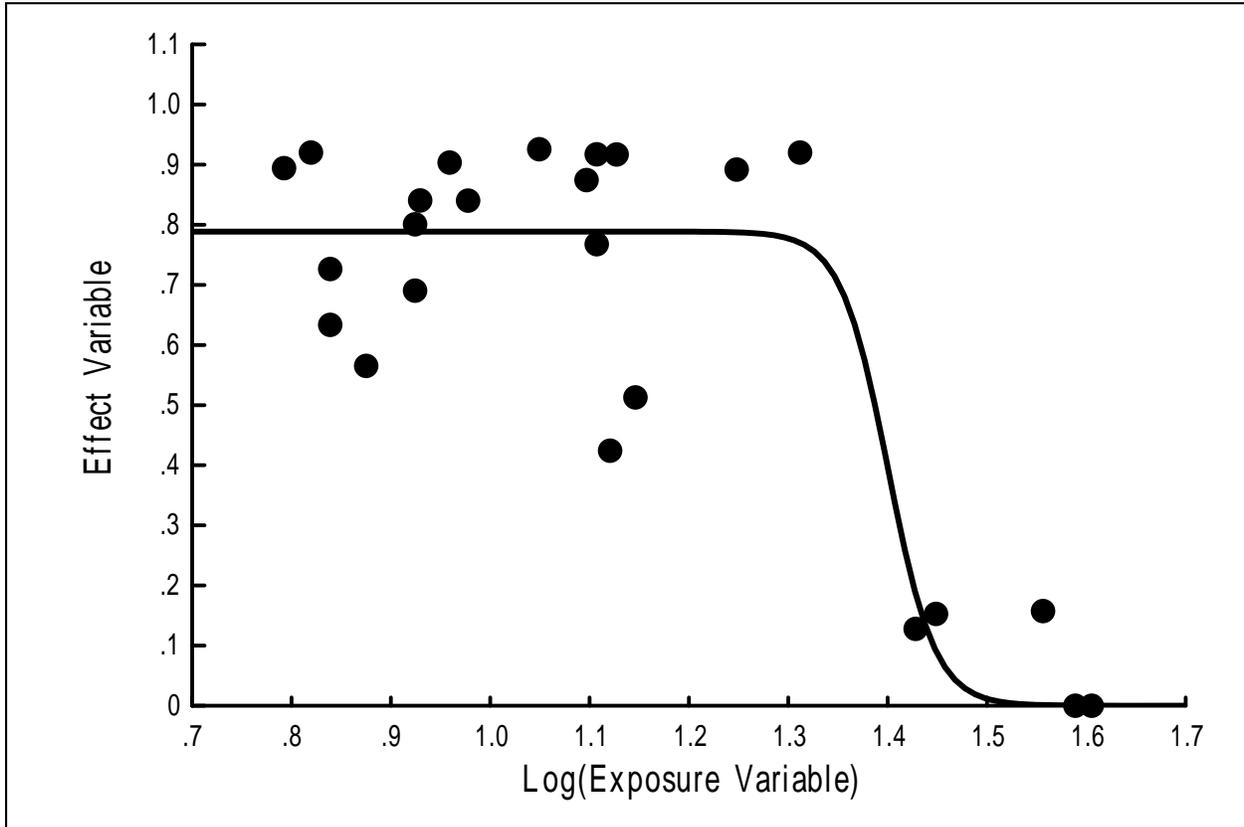
Exposure	Obs Effects	Pred Effects	Residual	Weight
1.1072	0.9577	0.7739	-0.1838	1.
0.9243	0.4286	0.7739	0.3454	1.
0.9294	0.6773	0.7739	0.0966	1.
0.9243	0.5903	0.7739	0.1836	1.
0.9590	0.5191	0.7739	0.2548	1.
0.8751	0.8696	0.7739	-0.0956	1.
0.8195	0.6626	0.7739	0.1113	1.
0.8388	0.5926	0.7739	0.1813	1.
0.7924	0.9068	0.7739	-0.1329	1.
1.1461	0.5907	0.7739	0.1832	1.
0.8388	0.7786	0.7735	-0.0051	1.
0.9777	0.9320	0.7729	-0.1591	1.
1.1072	0.9761	0.7726	-0.2035	1.
1.6053	0.0000	0.7726	0.7726	1.
1.5563	0.3841	0.7722	0.3880	1.
1.4281	0.1957	0.7719	0.5763	1.
1.2480	0.8505	0.7710	-0.0795	1.
1.5888	0.0000	0.7503	0.7503	1.
1.1206	0.9224	0.6926	-0.2297	1.
1.1271	0.9340	0.3376	-0.5964	1.
1.3118	0.8133	0.2602	-0.5532	1.
1.0969	0.7744	0.0404	-0.7339	1.
1.0492	0.8682	0.0212	-0.8470	1.
1.4487	0.2765	0.0152	-0.2612	1.

Egg Se vs. Cranio Facial Fraction Normal

Error Summary

Large Standard Error for Steepness

Egg Se vs. Skeletal Fraction Normal



Parameter Summary (Logistic Equation Regression Analysis)

Parameter	Guess	FinalEst	StdError	95%LCL	95%UCL
LogX50	1.4219	1.4002	0.0489	1.2986	1.5019
S	3.519	10.557	14.610	-19.827	40.940
Y0	0.7936	0.7887	0.0351	0.7157	0.8617

Effect Concentration Summary

%Effect	Xp Est	95%LCL	95%UCL
50.0	25.13	19.89	31.76
20.0	23.30	15.01	36.18
10.0	22.29	12.68	39.20
5.0	21.40	10.84	42.25

Egg Se vs. Skeletal Fraction Normal

Regression Analysis of Variance

Source	df	SS	MS	F	Alpha
Total	23	2.397	0.1042		
Regression	2	1.929	0.9644	43.3	0.0000
Error	21	0.468	0.0223		

Data Summary

Exposure	Obs Effects	Pred Effects	Residual	Weight
0.7924	0.8943	0.7887	-0.1056	1.
0.8195	0.9202	0.7887	-0.1316	1.
0.8388	0.7263	0.7887	0.0624	1.
0.8388	0.6336	0.7887	0.1551	1.
0.8751	0.5652	0.7887	0.2235	1.
0.9243	0.8008	0.7887	-0.0121	1.
0.9243	0.6903	0.7887	0.0984	1.
0.9294	0.8404	0.7887	-0.0517	1.
0.9590	0.9034	0.7887	-0.1147	1.
0.9777	0.8402	0.7887	-0.0515	1.
1.0492	0.9257	0.7887	-0.1370	1.
1.0969	0.8744	0.7887	-0.0857	1.
1.1072	0.9173	0.7887	-0.1286	1.
1.1072	0.7676	0.7887	0.0211	1.
1.1206	0.4242	0.7887	0.3645	1.
1.1271	0.9170	0.7887	-0.1283	1.
1.1461	0.5130	0.7887	0.2757	1.
1.2480	0.8918	0.7874	-0.1043	1.
1.3118	0.9200	0.7703	-0.1497	1.
1.4281	0.1278	0.1856	0.0578	1.
1.4487	0.1527	0.0902	-0.0625	1.
1.5563	0.1575	0.0011	-0.1564	1.
1.5888	0.0000	0.0003	0.0003	1.
1.6053	0.0000	0.0001	0.0001	1.

Egg Se vs. Skeletal Fraction Normal

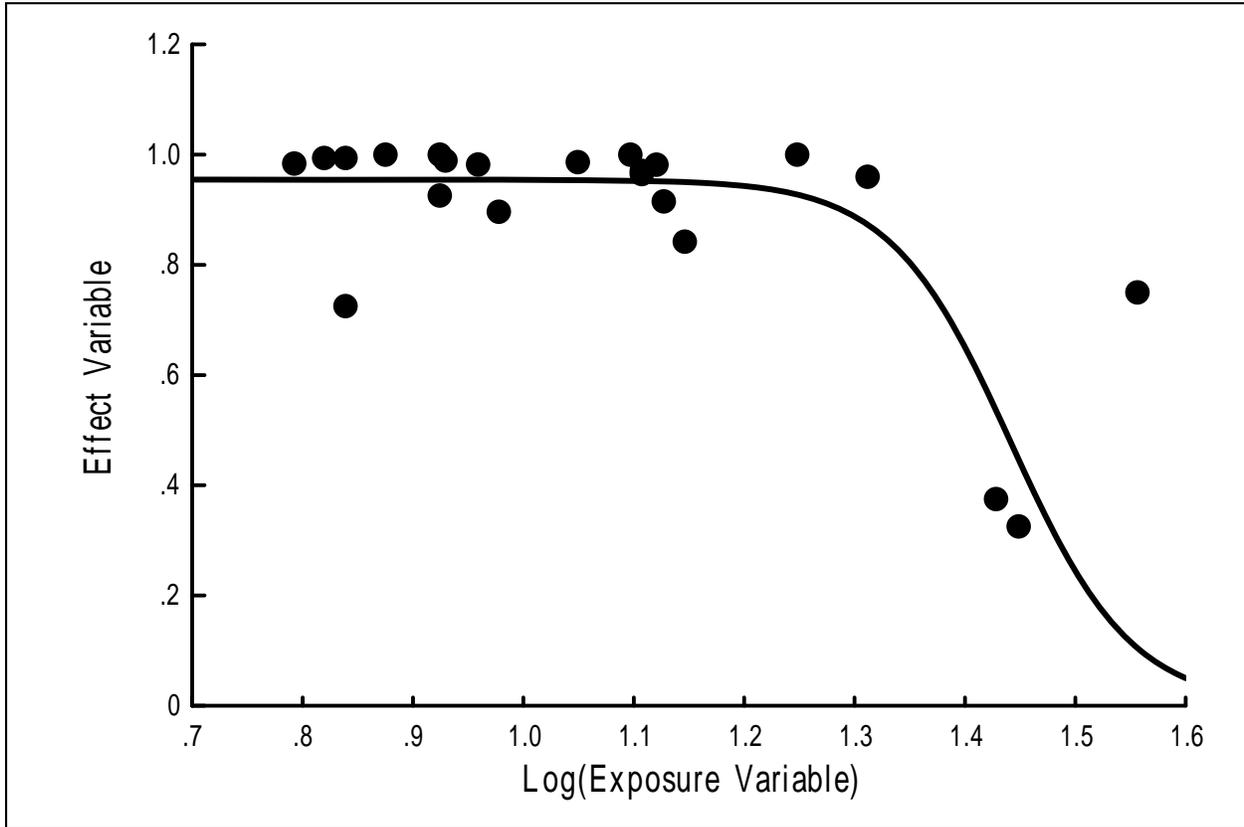
Error Summary

Maximum Iterations Reached Without Convergence

Steepness At Maximum or Minimum Limit

Large Standard Error for Steepness

Egg Se vs. Finfold Fraction Normal



Parameter Summary (Logistic Equation Regression Analysis)

Parameter	Guess	FinalEst	StdError	95%LCL	95%UCL
LogX50	1.4119	1.4416	0.0270	1.3851	1.4982
S	4.979	4.568	2.808	-1.310	10.446
Y0	0.9576	0.9546	0.0416	0.8675	1.0417

Effect Concentration Summary

%Effect	Xp Est	95%LCL	95%UCL
50.0	27.65	24.27	31.49
20.0	23.22	17.85	30.19
10.0	20.96	14.30	30.73
5.0	19.077	11.591	31.398

Egg Se vs. Finfold Fraction Normal

Regression Analysis of Variance

Source	df	SS	MS	F	Alpha
Total	21	0.767	0.0365		
Regression	2	0.212	0.1062	3.64	0.0459
Error	19	0.554	0.0292		

Data Summary

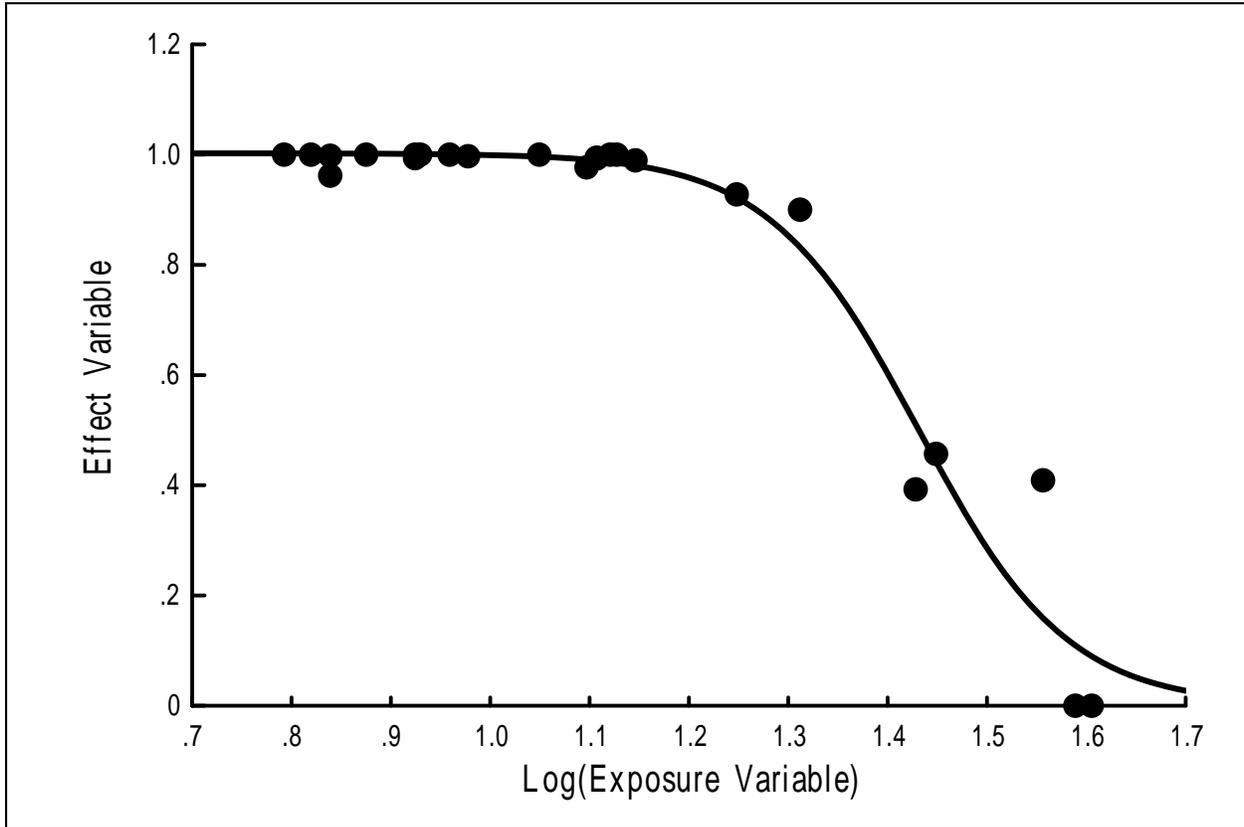
Exposure	Obs Effects	Pred Effects	Residual	Weight
0.7924	0.9839	0.9546	-0.0293	1.
0.8195	0.9939	0.9545	-0.0393	1.
0.8388	0.9938	0.9545	-0.0393	1.
0.8388	0.7252	0.9545	0.2294	1.
0.8751	1.0000	0.9545	-0.0455	1.
0.9243	1.0000	0.9545	-0.0455	1.
0.9243	0.9258	0.9545	0.0287	1.
0.9294	0.9894	0.9545	-0.0349	1.
0.9590	0.9820	0.9544	-0.0276	1.
0.9777	0.8964	0.9544	0.0579	1.
1.0492	0.9865	0.9538	-0.0327	1.
1.0969	1.0000	0.9528	-0.0472	1.
1.1072	0.9706	0.9524	-0.0181	1.
1.1072	0.9648	0.9524	-0.0123	1.
1.1206	0.9819	0.9519	-0.0301	1.
1.1271	0.9151	0.9515	0.0364	1.
1.1461	0.8420	0.9503	0.1083	1.
1.2480	1.0000	0.9276	-0.0724	1.
1.3118	0.9600	0.8732	-0.0868	1.
1.4281	0.3750	0.5359	0.1609	1.
1.4487	0.3253	0.4465	0.1212	1.
1.5563	0.7500	0.1046	-0.6454	1.

Egg Se vs. Finfold Fraction Normal

Error Summary

Maximum Iterations Reached Without Convergence
Large Standard Error for Steepness

Egg Se vs. Edema Fraction Normal



Parameter Summary (Logistic Equation Regression Analysis)

Parameter	Guess	FinalEst	StdError	95%LCL	95%UCL
LogX50	1.4299	1.4310	0.0133	1.4033	1.4587
S	2.116	3.328	0.525	2.236	4.420
Y0	0.9953	1.0024	0.0186	0.9638	1.0410

Effect Concentration Summary

%Effect	Xp Est	95%LCL	95%UCL
50.0	26.98	25.31	28.75
20.0	21.23	18.95	23.77
10.0	18.448	15.818	21.515
5.0	16.211	13.361	19.670

Egg Se vs. Edema Fraction Normal

Regression Analysis of Variance

Source	df	SS	MS	F	Alpha
Total	23	2.3608	0.10264		
Regression	2	2.2570	1.12852	228.	0.0000
Error	21	0.1038	0.00494		

Data Summary

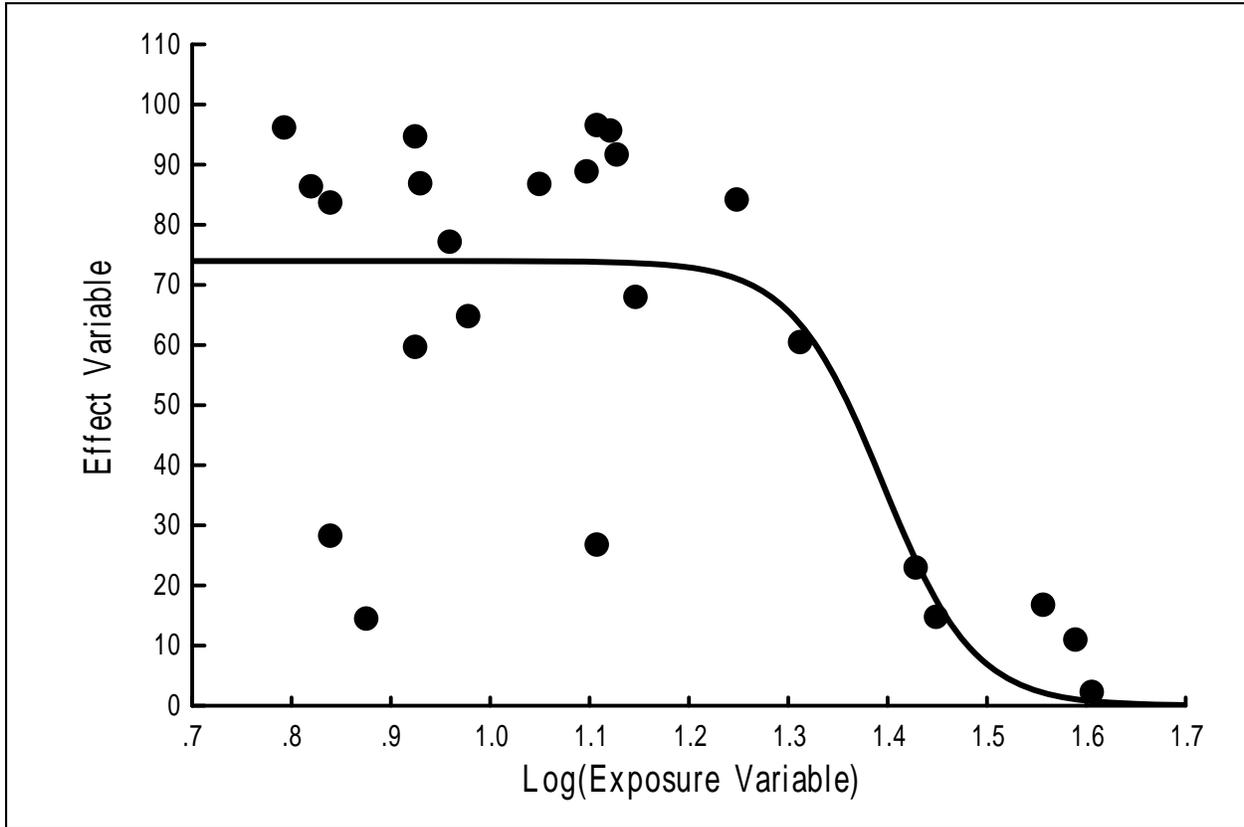
Exposure	Obs Effects	Pred Effects	Residual	Weight
0.7924	1.0000	1.0022	0.0022	1.
0.8195	1.0000	1.0021	0.0021	1.
0.8388	0.9979	1.0021	0.0041	1.
0.8388	0.9618	1.0021	0.0402	1.
0.8751	1.0000	1.0018	0.0018	1.
0.9243	1.0000	1.0013	0.0013	1.
0.9243	0.9935	1.0013	0.0077	1.
0.9294	1.0000	1.0012	0.0012	1.
0.9590	1.0000	1.0006	0.0006	1.
0.9777	0.9970	1.0000	0.0030	1.
1.0492	1.0000	0.9963	-0.0037	1.
1.0969	0.9769	0.9908	0.0139	1.
1.1072	0.9945	0.9892	-0.0053	1.
1.1072	0.9930	0.9892	-0.0038	1.
1.1206	1.0000	0.9866	-0.0134	1.
1.1271	1.0000	0.9852	-0.0148	1.
1.1461	0.9896	0.9803	-0.0093	1.
1.2480	0.9278	0.9218	-0.0060	1.
1.3118	0.9000	0.8323	-0.0677	1.
1.4281	0.3925	0.5108	0.1182	1.
1.4487	0.4570	0.4424	-0.0146	1.
1.5563	0.4091	0.1591	-0.2500	1.
1.5888	0.0000	0.1092	0.1092	1.
1.6053	0.0000	0.0897	0.0897	1.

Egg Se vs. Edema Fraction Normal

Error Summary

No Errors

Egg Se vs. Overall Survival



Parameter Summary (Logistic Equation Regression Analysis)

Parameter	Guess	FinalEst	StdError	95%LCL	95%UCL
LogX50	1.4184	1.3950	0.0530	1.2848	1.5051
S	2.683	5.419	5.029	-5.039	15.877
Y0	75.75	73.98	5.79	61.93	86.03

Effect Concentration Summary

%Effect	Xp Est	95%LCL	95%UCL
50.0	24.83	19.27	32.00
20.0	21.43	13.60	33.77
10.0	19.661	10.745	35.978
5.0	18.161	8.594	38.380

Egg Se vs. Overall Survival

Regression Analysis of Variance

Source	df	SS	MS	F	Alpha
Total	23	25808.	1122.		
Regression	2	14085.	7042.	12.6	0.0003
Error	21	11724.	558.		

Data Summary

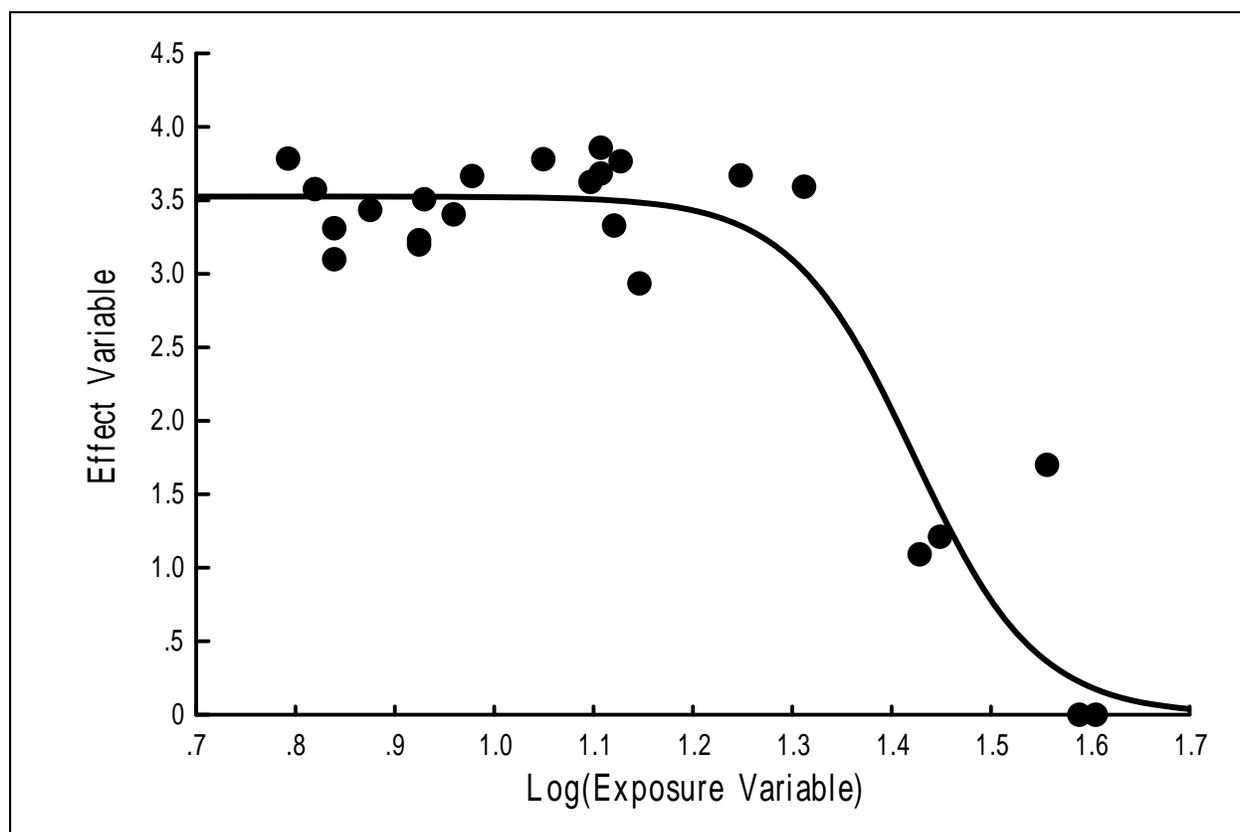
Exposure	Obs Effects	Pred Effects	Residual	Weight
0.7924	96.2000	73.9772	-22.2227	1.
0.8195	86.4000	73.9771	-12.4229	1.
0.8388	83.7000	73.9770	-9.7230	1.
0.8388	28.3000	73.9770	45.6770	1.
0.8751	14.5000	73.9765	59.4765	1.
0.9243	94.7000	73.9747	-20.7253	1.
0.9243	59.7000	73.9747	14.2747	1.
0.9294	86.9000	73.9743	-12.9257	1.
0.9590	77.2000	73.9716	-3.2284	1.
0.9777	64.8000	73.9687	9.1687	1.
1.0492	86.8000	73.9363	-12.8637	1.
1.0969	88.9000	73.8619	-15.0381	1.
1.1072	96.6000	73.8331	-22.7669	1.
1.1072	26.8000	73.8331	47.0331	1.
1.1206	95.7000	73.7847	-21.9153	1.
1.1271	91.7000	73.7555	-17.9445	1.
1.1461	68.0000	73.6428	5.6428	1.
1.2480	84.2000	71.0420	-13.1580	1.
1.3118	60.5000	63.5194	3.0194	1.
1.4281	23.0000	24.2424	1.2424	1.
1.4487	14.8000	17.5956	2.7956	1.
1.5563	16.8000	2.1755	-14.6245	1.
1.5888	11.0000	1.0911	-9.9089	1.
1.6053	2.3000	0.7668	-1.5332	1.

Egg Se vs. Overall Survival

Error Summary

Large Standard Error for Steepness

Egg Se vs. Sum fraction normal



Parameter Summary (Logistic Equation Regression Analysis)

Parameter	Guess	FinalEst	StdError	95%LCL	95%UCL
LogX50	1.4419	1.4222	0.0207	1.3791	1.4652
S	2.733	4.041	1.266	1.409	6.672
Y0	3.516	3.526	0.108	3.301	3.751

Effect Concentration Summary

%Effect	Xp Est	95%LCL	95%UCL
50.0	26.43	23.94	29.19
20.0	21.70	18.09	26.02
10.0	19.329	15.072	24.789
5.0	17.377	12.696	23.785

Egg Se vs. Sum fraction normal

Regression Analysis of Variance

Source	df	SS	MS	F	Alpha
Total	23	32.34	1.4060		
Regression	2	28.46	14.2307	77.1	0.0000
Error	21	3.88	0.1846		

Data Summary

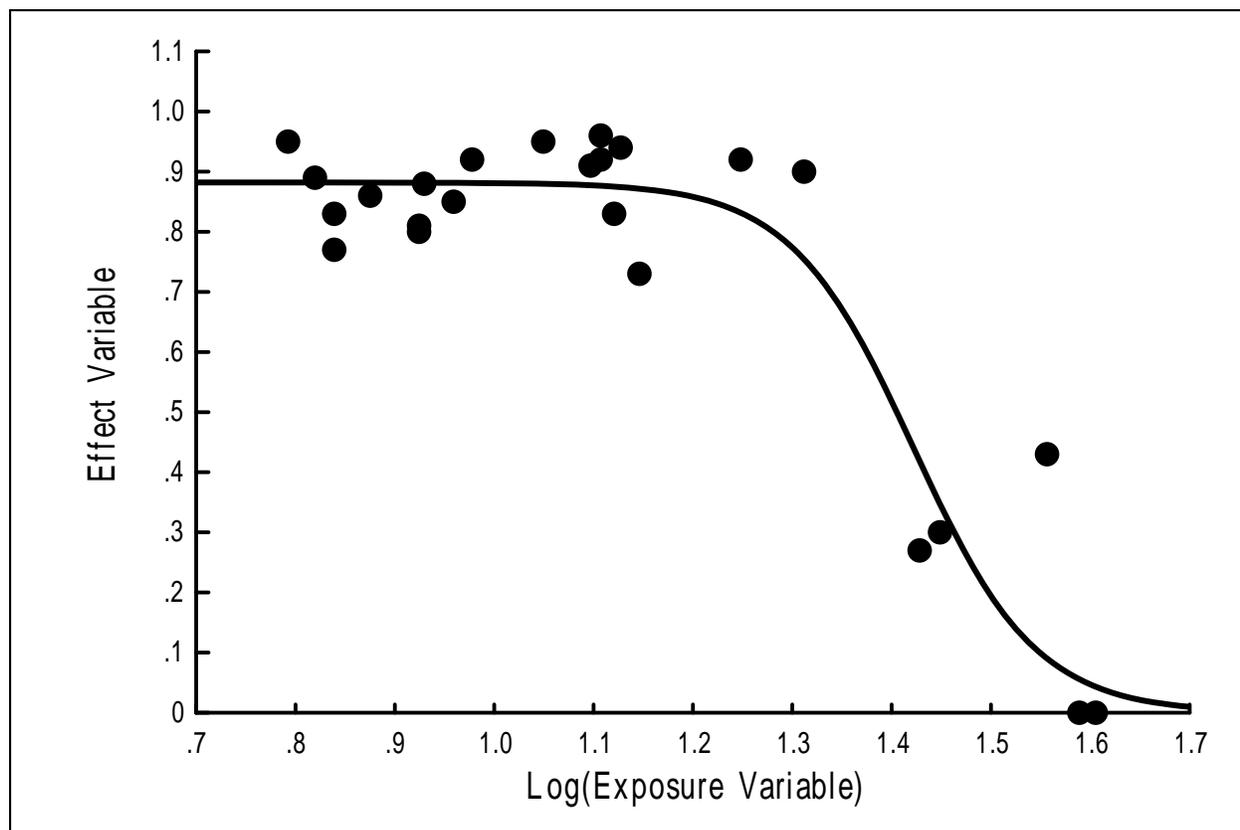
Exposure	Obs Effects	Pred Effects	Residual	Weight
0.7924	3.7849	3.5262	-0.2587	1.
0.8195	3.5767	3.5261	-0.0505	1.
0.8388	3.3107	3.5261	0.2154	1.
0.8388	3.0992	3.5261	0.4269	1.
0.8751	3.4348	3.5258	0.0911	1.
0.9243	3.2293	3.5252	0.2959	1.
0.9243	3.2000	3.5252	0.3252	1.
0.9294	3.5071	3.5251	0.0180	1.
0.9590	3.4045	3.5244	0.1199	1.
0.9777	3.6657	3.5237	-0.1420	1.
1.0492	3.7804	3.5179	-0.2625	1.
1.0969	3.6256	3.5081	-0.1175	1.
1.1072	3.8585	3.5048	-0.3537	1.
1.1072	3.6831	3.5048	-0.1783	1.
1.1206	3.3285	3.4996	0.1711	1.
1.1271	3.7660	3.4967	-0.2693	1.
1.1461	2.9352	3.4861	0.5509	1.
1.2480	3.6701	3.3271	-0.3430	1.
1.3118	3.5933	3.0194	-0.5739	1.
1.4281	1.0910	1.6780	0.5870	1.
1.4487	1.2114	1.3906	0.1792	1.
1.5563	1.7007	0.3620	-1.3387	1.
1.5888	0.0000	0.2233	0.2233	1.
1.6053	0.0000	0.1737	0.1737	1.

Egg Se vs. Sum fraction normal

Error Summary

No Errors

Egg Se vs. Mean Fraction normal



Parameter Summary (Logistic Equation Regression Analysis)

Parameter	Guess	FinalEst	StdError	95%LCL	95%UCL
LogX50	1.4418	1.4218	0.0210	1.3780	1.4656
S	2.723	4.039	1.283	1.371	6.707
Y0	0.8797	0.8820	0.0275	0.8248	0.9391

Effect Concentration Summary

%Effect	Xp Est	95%LCL	95%UCL
50.0	26.41	23.88	29.21
20.0	21.68	18.02	26.07
10.0	19.310	15.000	24.859
5.0	17.359	12.622	23.874

Egg Se vs. Mean Fraction normal

Regression Analysis of Variance

Source	df	SS	MS	F	Alpha
Total	23	2.028	0.08816		
Regression	2	1.778	0.88898	74.8	0.0000
Error	21	0.250	0.01189		

Data Summary

Exposure	Obs Effects	Pred Effects	Residual	Weight
0.7924	0.9500	0.8819	-0.0681	1.
0.8195	0.8900	0.8819	-0.0081	1.
0.8388	0.8300	0.8819	0.0519	1.
0.8388	0.7700	0.8819	0.1119	1.
0.8751	0.8600	0.8818	0.0218	1.
0.9243	0.8100	0.8817	0.0717	1.
0.9243	0.8000	0.8817	0.0817	1.
0.9294	0.8800	0.8816	0.0016	1.
0.9590	0.8500	0.8815	0.0315	1.
0.9777	0.9200	0.8813	-0.0387	1.
1.0492	0.9500	0.8798	-0.0702	1.
1.0969	0.9100	0.8773	-0.0327	1.
1.1072	0.9600	0.8765	-0.0835	1.
1.1072	0.9200	0.8765	-0.0435	1.
1.1206	0.8300	0.8752	0.0452	1.
1.1271	0.9400	0.8745	-0.0655	1.
1.1461	0.7300	0.8718	0.1418	1.
1.2480	0.9200	0.8318	-0.0882	1.
1.3118	0.9000	0.7544	-0.1456	1.
1.4281	0.2700	0.4184	0.1484	1.
1.4487	0.3000	0.3466	0.0466	1.
1.5563	0.4300	0.0901	-0.3399	1.
1.5888	0.0000	0.0556	0.0556	1.
1.6053	0.0000	0.0433	0.0433	1.

Egg Se vs. Mean Fraction normal

Error Summary

No Errors

APPENDIX F
Agency Comments on February 5, 2009 Draft Report

IDEQ – Pocatello Comments on
Draft Brown Trout Laboratory Reproduction Studies Conducted in Support of
Development of a Site-Specific Selenium Criterion
18 Feb 09

General Comments

- The initial power analysis suggested 40 fish were necessary to cover the expected range of maternal selenium. The number of fish in the mid-range (i.e., effects level) may not provide a sufficient sample size to completely resolve this question.
- Be consistent with references, e.g., use of period after et al, separating citations with comma or semi-colons.

Specific Comments

- Page 4, Section 1.2, Bullet 3. Was this relationship graphically represented in the report? If not, it needs to be.
- Page 7, Section 2.1.2, Paragraph 2. Why were adult carcasses shipped to ENSR?
- Page 12, Section 3.2, Paragraph 1. Eggs were collected from 26 fish yet Table 5 shows that 34 ripe females were retained. Two of those females (from Stump Creek) were released. What was the disposition of the other 6 fish?
- Page 12, Section 3.2.1, Paragraph 3. Please identify the previous field monitoring data to which you refer.
- Page 15, Section 3.4.3, Paragraph 2, Line 8. Please remind the reader, maybe in parentheses, the change in fertilization technique.
- Page 16, Section 3.4.4, Paragraph 1, Line 9. Please reword the sentence beginning with “As noted previously . . .” as I could not understand the point as written.
- Page 21, Section 4.1, Paragraph 2, Last line. Site LSV-2C is not depicted in Figure 24. Please do so.
- Page 24, Section 4.3.1, Paragraph 3, Line 1. Please explain what is meant by “variability of the overall mortality endpoint.”
- Page 25, Section 4.4, Paragraph 1. It was mentioned previously that the data are not normally distributed. Please explain how the TRAP model is able to deal with data which are not normally distributed.
- Page 27, Section 4.4.2, First partial paragraph, Line 1. Please explain what is meant when it is said that no errors were reported in the TRAP software output.

Editorial Comments

- Page 1, Introduction, Paragraph 3, Line 5. Change *compliment* to complement.
- Page 2, Introduction, Bullet 9, Line 1. Eliminate the s from cutthroat.
- Page 3, Section 1.1, Paragraph 1, Line 1. Add for selenium to the end of the first sentence to read, “. . . water quality standard for selenium.”

- Page 3, Section 1.1, Paragraph 1, Line 2. Add downstream to read, “. . . exceedances decline downstream with tributary inflows.”
- Page 3, Section 1.1, Paragraph 2, Line 1. Consider adding gravid and deleting *ready to spawn* to read, “. . . testing used gravid adult wild fish captured at . . .”
- Page 7, Section 2.1.3, Paragraph 2, Line 2. The town is Lewistown not *Lewiston*. Also occurs on Page 13, Section 3.3, Paragraph 2.
- Page 10, Section 2.4, Paragraph 1, Line 2. Consider using hatching rate or hatching success rather than just *hatch*.
- Page 10, Section 2.4, Paragraph 1, Line 5. Change *changed* to change.
- Page 10, Section 2.4, Paragraph 2, Last line. Change *EC10* to EC₁₀ to be consistent with EC₂₀ and later usage.
- Page 12, Section 3.2.1, Paragraph 2, Line 6. Eliminate the comma following *fish*.
- Page 15, Section 3.4.3, Paragraph 2, Line 16. Please consider revising the sentence to “As shown in Figure 12, the eggs from those five LSV-2C samples had the highest . . .”
- Page 21, Section 4.1, Paragraph 2, Line 5. Eliminate the s on *terms*.
- Page 24, Section 4.3.1, Paragraph 1, Line 5. Add a comma between *analyze* and *for*.
- Page 26, Section 4.4.2, Paragraph 2, Line 6. Delete *does*.
- Page 28, Section 4.4.3, Paragraph 4, Line 10. Consider changing the sentence to read, “. . . not only a good fit, but a low variability as well.”
- Page 30, Section 4.5, Paragraph 2, Line 3. Change *later* to latter.
- Page 30, Section 4.5, Paragraph 3, Line 4. Change *increase* to increased or increasing.
- Page 30, Section 4.5, Paragraph 4, Line 8. Eliminate one of the two ands.

Table & Figure Comments

- Tables 1 and 5. Add zeros in blank cells where appropriate to be consistent with first row.
- Tables 2, 3, and 4. Put in the footnotes what the shaded cells represent (e.g., October 2007 sampling).
- Table 5. The table notes that 8 ripe females were retained from CC-150, but there were 9 total egg samples submitted. What accounts for this difference?
- Table 6. Change *dwt* to dw in the legend and column headings.
- Table 7. Put in the footnote what the shaded cells represent (e.g., October/November 2007 sampling).
- Table 8. Change *Ec_x* to EC_x in legend and column heading. Change *Comcentration* to Concentration in column heading.
- Figures 10, 11, 14, 15, 16, 17, 18, and 23. Please denote that the order of fish as represented on the X-axis is in order of egg selenium concentration.
- Figure 30. Is the heavy black line for both wild and hatchery fish? If so please identify accordingly.
- Figure 33. Eliminate the period that precedes Percent.

Forest Service Comments

Brown Trout Laboratory Reproduction Studies Conducted in Support of a Site-specific Criterion (Newfields 2009)

Mary E. Kauffman

February 25, 2009

Comments provided by Steve Bauer under contract to the Forest Service

Figure 2. Fish Collection vs Exposure. The adult brown trout used in the study are grouped by Background (CC-150), Low Exposure (CC-350), and High Exposure (LSV2C). It should be noted that no adult spawners were collected from Hoopes Springs, the highest exposure area, because Hoopes Springs does not provide spawning habitat and that this is one of the reasons that the adult spawner selenium concentrations are lower than the total brown trout population selenium concentrations as summarized in Figure 47.

Figure 3 and 4. Selenium Concentration vs. Size. These figures include both hatchery and wild exposed fish. The illustration of the relationship between size and selenium concentration would be improved by including only the wild fish so that the Y-scale can be adjusted appropriately.

Figure 5 and Figure 47. Range of selenium exposure. These two figures show the same information. I suggest using only Figure 47 since it contains more data points for the same information. The sufficiency of the adult fish used in the lab study in representing the full range of selenium exposure below Hoopes Springs is an important issue. Figure 47 displays the 90th percentile (I assume of all fish from the watershed, but not the adult spawners for this study). It is apparent that there are numerous brown trout in Hoopes and Sage Creek that exceed the range of selenium concentration in the adult spawners used in the brown trout lab study.

The question of adequate sample size is discussed on page 31, 3rd paragraph, of the report. The report reiterates that the target tissue concentration for the study was the 90th percentile, 20 mg/kg dw. Two adult female brown trout were captured that fit the criteria, with a max of 22.6 mg/kg. The report concludes that the adult study fish span the range of whole body tissue concentration for brown trout from the watershed. It also notes, more importantly, that the resulting range of egg selenium spans the full range of effects.

This issue of whether the fish captured meets the target deserves additional analysis and discussion. A visual inspection of Figure 47 indicates a much lower range of selenium in the test fish in comparison to the range of selenium in the fish population in the watershed. A box plot of the adult study fish vs all other fish collected would provide a screening assessment. Descriptive statistics (mean, sd, median, interquartile range, etc.) and statistical tests may help to answer this question.

Whether the egg selenium concentration spans the full range of effects is a separate question that also deserves some further explanation. The TRAP model figures, Figure 37 to Figure 46 illustrate similar relationships among the endpoints. Would these relationships be improved by including more female spawners with a higher body burden, and would that affect the outcome, that is the 13.35 mg/kg concentration endpoint? Several of the models exhibit poor R^2 values and wide confidence intervals. One could speculate that the inflection point would be steeper, and the confidence intervals would be tighter if more data from the higher selenium range were available.

Number of High Exposure Fish. The outcome of the TRAP models depend on the influence of 5 fish (egg batch per adult spawner). The conclusion of the report relies primarily on two relationships, illustrated in Figure 40 and Figure 46, evaluated by logistic regression: 1) Egg selenium vs percent survival, and 2) Egg selenium vs mean fraction normal fish larvae – the deformities metric. The R^2 for these equations is 0.89 and 0.88, indicating a good fit for the data. Does the high R^2 alone indicate that there is good fit in the model and therefore a valid conclusion? Or would this conclusion be altered if there was a larger sample of spawners with high whole body selenium?

On the call we asked the EPA toxicologists to address this issue in their written comments since they have specific experience in conducting these kinds of tests.

Figure 10 and 11. There are several figures that include hatchery fish with the wild exposed fish. The fish from the Saratoga Hatchery, labeled SC, exhibited high egg mortality and poor hatch. One would assume the fish were included in the study as a control in evaluation endpoints. It turns out that the hatchery fish are not used as controls for effects evaluation. I suggest revising these and other figures either to remove these from the data set with the explanation of why and/or move the information into an appendix. Including these fish seems an unnecessary distraction.

A footnote should be added to explain reasons for the mortality if known for the egg batches that are labeled “None fertilized” and “Dead upon arrival”.

Figure 12. Five egg batches from LSV2C did not swim up. It would be useful to indicate in parentheses what the average selenium concentration was in these eggs.
Response:

Figure 14, Figure 18. The authors should consider removing the SC Hatchery fish from these figures. Mortality is apparently related to factors outside of the test for selenium and therefore this set of fish does not function as a control. The other set of hatchery fish should be adequate for the intended purpose.

Figure 25 vs Figure 26. Figure 26 illustrates the relationship between the selenium in egg vs whole body in the wild fish. This is the relationship that is later used appropriately to calculate the whole body selenium tissue value from the egg selenium concentration. Figure 25, which includes the hatchery fish, is extraneous.

Figure 27. This graph shows a weak relationship between fecundity (number of eggs) and egg selenium concentration. It may not add to the project outcome, but it seems that the variation introduced by body size should be removed prior running the regression. A variable such as Number of eggs/length or eggs/weight could be used as the dependent variable.

Even though there is a weak relationship, the graph illustrates that a female spawner with a higher body burden produces a lower number of eggs. One would expect the R^2 to improve if additional female spawners with a higher body burden had been captured, this would then be more representative of the selenium body burden of the total brown trout population. Is fecundity an issue that reduces the population in Sage and Crow Creeks?

Deformities metric. After examination of the data Newfields chose to use the fraction normal versus the GSI scores in the TRAP model. The GSI scores necessarily depend on the judgment of the analyst in deciding the category score, a procedure which introduces a source of variability. Evaluating only “normal” vs “non-normal” provides a more robust that would expected to be highly repeatable between different observers, thereby increasing method precision.

Selecting Biological Endpoints. Newfields used logistic regression to evaluate a number of different biological endpoints (summarized in Table 8). These endpoints are consistent with the literature in evaluating the most sensitive life stage. Selecting the two endpoints with the best fit; hatch to test end and overall fraction of deformities normal; is logical and captures the range of response in all the endpoints.

EC 20 vs. EC 10. Charlie Delos pointed out that EPA policy would be to use the EC 10 vs the EC 20 concentration for this type of contaminant. This decreases the EC for the selected endpoints: 21.6 mg/kg to 17.7 mg/kg for the survival endpoint, and 21.7 mg/kg to 19.3 mg/kg for the deformities endpoint. Using the EC 10 would consequently lower the calculated whole body tissue concentration. We would assume that Newfields will revise the toxicity level to be consistent with EPA policy.

Approach. The methods used were consistent with the study approach previously reviewed by the work group and with the literature. Using numerous endpoints and several test species is a strength of these studies. Combining the results of this lab study with the Yellowstone cutthroat trout study will provide several lines of evidence that will be useful in proposing a site-specific criteria.

Documents: I would like to see the Parmetrix (2009) and Erickson (2008) documents referenced in this study posted on the website. I sent this request to Sean Covington by e-mail after our meeting.

APPENDIX G
EPA Comments on ECx Values

EPA's Draft Comments on Formation's August 2010 Draft Interpretive Findings for Field and Laboratory Studies and Literature Review in Support of a Site-Specific Selenium Criterion, Smoky Canyon Mine

December 21, 2010

The overall study is comprehensive and provides information needed to assess the nature of the selenium toxicity problem and determine the selenium effects concentration threshold in eggs, with potential to translate that to water concentrations, when the project reaches that stage.

Toxicity Study Conduct

Maternal Transfer Reproductive Studies (with offspring of wild-caught brown trout and Yellowstone cutthroat trout) – We have not identified any issues with the conduct of these studies.

We have incorporated the brown trout data in our internal draft national criterion document. This study is of outstanding quality in terms numbers of individual fish, comprehensiveness of endpoints evaluated, and chemical measurements taken. Although the more recent cutthroat trout study appears to be of comparable quality to the brown trout study, we have not yet had time to consider it for the national document, and as a result, our review of that data has not been as thorough as for the brown trout study.

Non-Reproductive (Non Maternal Transfer) Early Life Stage Survival – Yellowstone cutthroat trout only. Control survival, measured from start to finish, was low. If we were to assume that dissolved selenium has little propensity to enter eggs, such that hatchability is not an issue, survival is still somewhat low, around 70% measured from Day 6 (hatch) to the end of the test.

We have evaluated the results as Day 71 (end) survival divided by Day 6 (hatch) survival, as well as Day 71 divided by Day 38 (Se diet start) survival. Although survival at all Se-spiked exposures were lower than for control, there was no recognizable response to incrementally increased selenium concentrations over a 16 fold range, and no ECs can be calculated. It appears that the threshold for Se effects to this life stage, sans maternal transfer of Se, is higher than the highest tested concentration.

Our hypothesis prior to this test was that it would not involve a sensitive endpoint, and the results do not counter the hypothesis. This test, in spite of its difficulties with the health of the controls, probably should not be considered an issue.

Calculation of Brown Trout Effect Concentrations

Formation examined a number of different effects and chose survival from hatch to test end (Formation's Figure 5-5) as the best endpoint for deriving a criterion. Examining the graphed data independent of the fitted line, we agree with this choice. This is the endpoint we selected in the internal draft national criterion document. In contrast, the high background response variability at low selenium exposures makes total survival including hatch (Formation's Figure 5-4), facial-cranial deformities, and skeletal deformities (Figures 5-7 and 5-8) less desirable. Fraction normal (Figures 5-11 and 5-12), finfold deformities (Figure 5-9), and survival 15-days post feeding (Figure 5-6) show less effects at around 20 mg/kg, suggesting that they are slightly less sensitive measures. Edema (Figure 5-10) shows a pattern similar to survival from hatch to test end, but its measurement is more subjective. Because the test was so long, the concern about irreversible edema is already accounted for in the measurement of survival at test end. Consequently, independent of particular estimates of EC10s for all these endpoints, the data themselves favor survival to test end (Figure 5-5) as the endpoint of concern.

Formation used EPA's TRAP program to calculate the ECs. TRAP's help screens provide the following guidance for its application:

In the end, to effectively use this (or any similar) program, the user should examine the fitted curve relative to the data and decide if the various parameter estimates and confidence limits appear reasonable. The value of this type of toxicity relationship analysis is to provide some quantitative objectivity and assessment of uncertainty to the estimation of parameters of interest that the user already can approximate by inspection of the data. The computed toxicity relationship should be close to what someone could get by just "eyeballing" the data; otherwise, some aspect of the data, model, or analysis might be causing problems. This kind of analysis demands some judgment from the user - if the results don't look good, they probably aren't and more evaluation is needed.

TRAP has many options for calculating the EC, and Formation presented the one option that is the same as what EPA used in the internal draft national criterion document: nonlinear regression comparing the untransformed effect with the log of the exposure concentration, and fitting to a logistic S-curve. For this application there are nine options that could be considered: three options for transforming the effect scale, times three options for the generic shape of the fitted S-curve. The logistic S-curve that EPA commonly uses for calculating the ECs of various pollutants is ordinarily the most environmentally conservative approach because it has a long and relatively thick tail that never reaches zero effect – there is always some effect at any nonzero concentration. The other two shapes available are “threshold sigmoid” a curving S-shape with a nonzero threshold, and “piecewise linear”, more commonly called hockey stick. The non-zero threshold of these latter two shapes might be more consistent with idea that selenium is a necessary nutrient.

The EC50s, EC20s, and EC10s for these nine reasonable options are shown in Table 1. The values for R^2 should only be compared for the three Curve Shapes within Effect Transform groups, not between groups.

Table 1. Summary of alternative TRAP analyses for fitting the all brown trout data for Crow Creek and Lower Sage Creek (and excluding hatchery fish per both the Formation report and the EPA internal draft national criteria document).

Effect Transform	Curve Shape	EC50	EC20	EC10	R ²	
None	Logistic	30.52	21.62	17.677	0.888	See Fig. 1
	Threshold Sigmoid	30.69	21.8	18.344	0.888	
	Piecewise Linear	30.49	21.84	19.547	0.900	
SqRt	Logistic	30.08	21.91	18.205	0.858	See Fig. 2
	Threshold Sigmoid	30.17	22.08	18.865	0.858	
	Piecewise Linear	30.07	22.4	20.31	0.871	
ArcsinSqRt	Logistic	30.75	22.98	19.384	0.904	
	Threshold Sigmoid	30.73	22.91	19.764	0.904	
	Piecewise Linear	30.03	21.74	19.522	0.913	
Overall Average		30.39	22.14	19.07		
Coefficient of Variation		0.010	0.023	0.045		
Max/Min		1.024	1.063	1.149		

All nine models fit the complete dataset reasonably well. The EC50s vary negligibly. The EC20s vary slightly, with Max 6% greater than Min, and the EC10s vary slightly more with Max=20.31 mg/kg being 15% greater than Min=17.67 mg/kg. The EC10 presented in the Formation document is the most environmentally conservative choice. Its graph is presented in Figure 1.

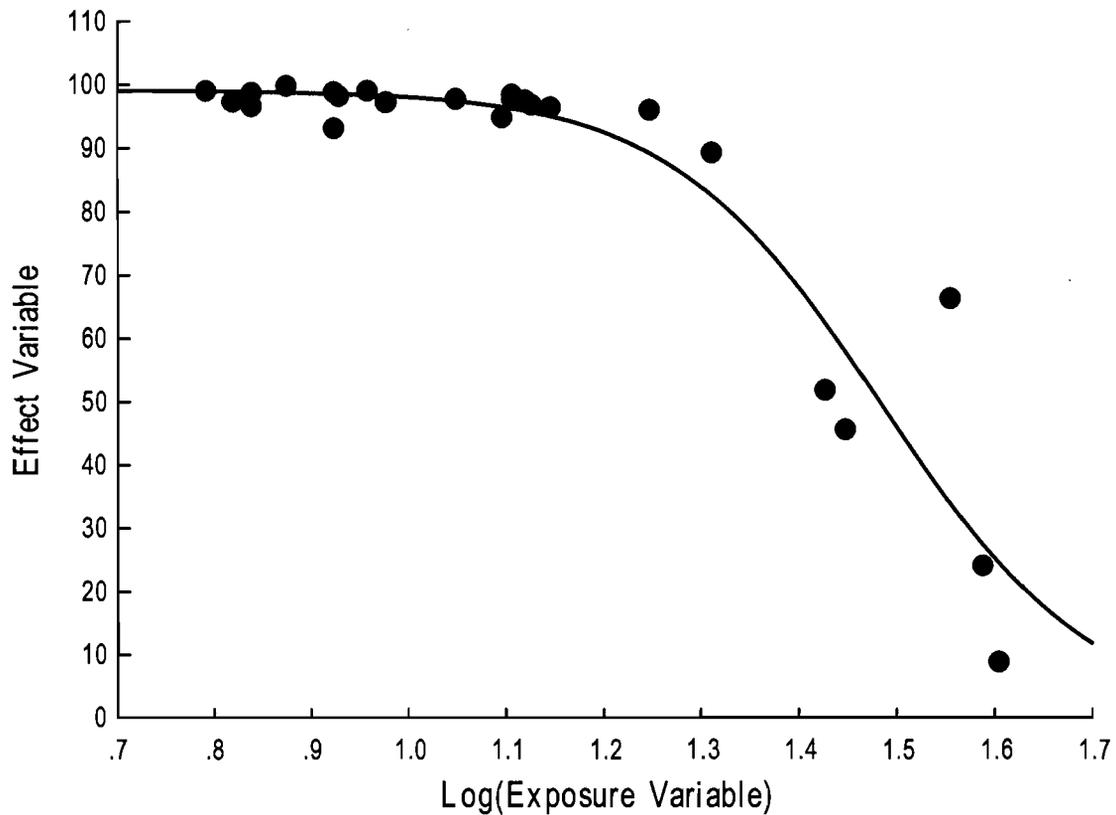


Figure 1. Brown trout nonlinear regression, logistic curve, effect not transformed, EC10=17.67 mg/kg.

X-Axis Log Scale Translator:

Log	1.1	1.2	1.3	1.4	1.5	1.6
Value	12.6	15.8	20.0	25.1	31.6	39.8

The Figure 1 curve can be seen to slightly overstate the observed effects at y-values near 90% (10% effect). In contrast to the line there is a point showing 8% (Abbott adjusted) reduction from background survival at a concentration of 20.5 mg/kg (log≈1.3), whereas the curve yields 10% reduction in survival at 17.67 mg/kg.

Figure 2 shows the curve yielding the highest EC10 in Table 1. Examining the data points nearest the beginning of the downward effect on survival, its fit to the observed data points is better than that shown in Figure 1, and likewise better than any of the other Table 1 models.

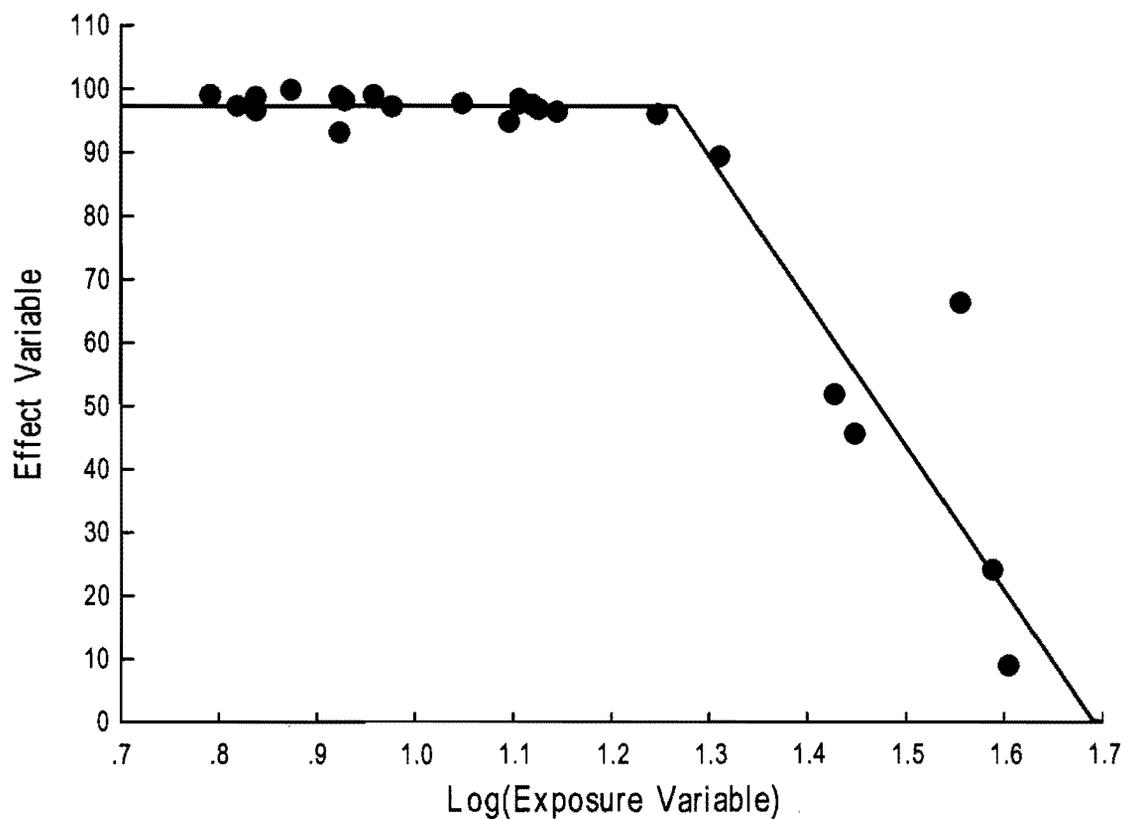


Figure 2. Brown trout nonlinear regression, piecewise linear (hockey stick) curve, square root effect transformation, EC10=20.3 mg/kg.

X-Axis Log Scale Translator:

Log	1.1	1.2	1.3	1.4	1.5	1.6
Value	12.6	15.8	20.0	25.1	31.6	39.8

It should be noted that the TRAP program does not know what percent effect the user is interested in: e.g., EC10, EC50, or EC90. In attempting to fit all the points as well as possible, it compromises the fit in the region we are most interested in, 15-30 mg/kg ($\log=1.2-1.5$). Consequently, one additional approach for estimating the EC10 is worth noting. Eliminating the three points above 30 mg/kg ($\log>1.5$) allows TRAP to focus on the threshold region we are interested in, as shown in Figure 3. This yields an EC10 of 20.7 mg/kg, thus indicating that the Formation's EC10 of 17.7 mg/kg is environmentally conservative.

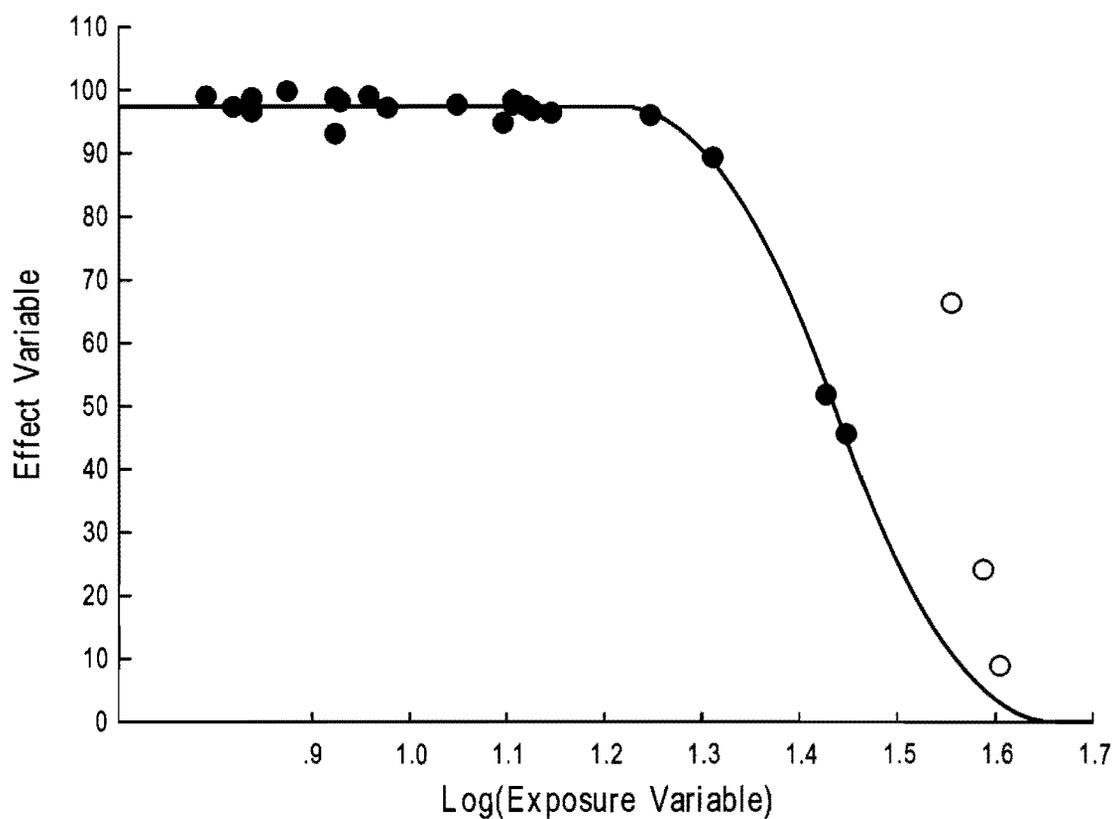


Figure 3. Brown trout nonlinear regression, sigmoid threshold, effect not transformed, observations above 30 mg/kg ($\log=1.5$) ignored. EC10=20.7 mg/kg. Replacing the sigmoid threshold shape with the logistic shape yields an essentially identical curve, with EC10=20.8 mg/kg.

X-Axis Log Scale Translator:

Log	1.1	1.2	1.3	1.4	1.5	1.6
Value	12.6	15.8	20.0	25.1	31.6	39.8

Translation to Whole Body Concentration

For translation from egg to whole body, Formation used log-log regression: $\log(\text{Egg Se})$ vs. $\log(\text{Whole Body Se})$, for *combined brown and cutthroat trout*, excluding hatchery fish. Because the log-log slope is slightly different from 1.0, the relationship is not quite a direct proportion, and the Egg/WB ratio varies slightly, from 1.53 to 1.54 in the range of the brown trout calculated EC10 and EC20.

For brown trout in the current internal draft national criteria document, we used the Smoky Canyon brown trout data, including hatchery fish, plus a few other measurements from elsewhere. We used arithmetic scale (not log scale) single-parameter regression (where the regression slope is estimated after forcing the line through the origin). We thereby obtained an Egg/WB ratio of 1.74, which yields a slightly more conservative WB tissue criterion translation. For application to Smoky Canyon, Formation's log-log approach might well be judged as good or better than the linear approach we used. However, the combining of brown and cutthroat data might or might not be viewed as appropriate, since the WB translation yields a slightly lower value using brown trout alone than using the combined data, although there are advantages to increasing the number of data points in the analysis. Considering other strains of cutthroat trout, external to the Smoky Canyon dataset, there is no evidence that mixing in cutthroat trout data would inherently bias the translated WB benchmark upward, since egg/WB ratios implied by the Hardy, Kennedy, and Rudolph data would tend to lower the WB translated benchmark.

Translation to Water

EPA is unable to comment on the water translation until the work progresses to that stage.